EXTRACTION AND CHARACTERIZATION OF HALAL GELATIN FROM WASTE BY-PRODUCT (BLACK TILAPIA SKIN)

NORHASIKIN BINTI ISMAIL

A thesis submitted in fulfillment of the requirement for the award of the Doctor of Philosophy in Mechanical Engineering

Faculty of Mechanical and Manufacturing Engineering Universiti Tun Hussein Onn Malaysia

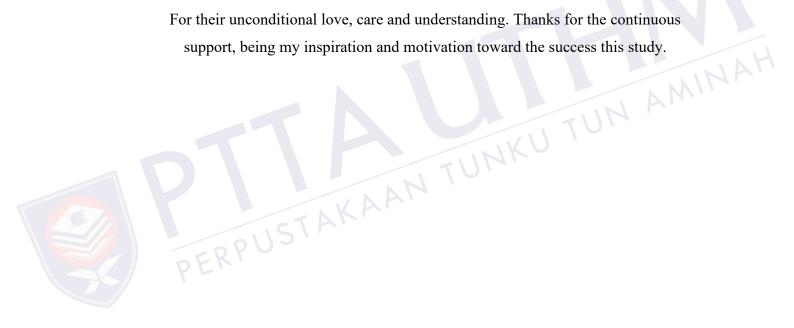
OCTOBER, 2022

To my precious Allah S.W.T., who gave me new life, hope and purpose of life.

To my beloved Prophet Muhammad S.A.W., who gave a guidance thru his Sunnah and Hadis.

Special thanks to my beloved family, Especially my mother, in laws, my husband, Shukur Saleh and my son, Muhammad Shafwan Naufal,

For their unconditional love, care and understanding. Thanks for the continuous



ACKNOWLEDGEMENT

Above all, I thank to Allah S.W.T. for his grace and strength that given to me during the study which make the research possible. Secondly, I would like to extend my sincere gratitude to my Ph.D supervisor, Prof. Dr. Hasan Zuhudi Bin Abdullah for his guidance, continuous motivation and encouragement in this study. I also would like to thank my co-supervisors, Assoc. Prof. Dr. Maizlinda Izwana Binti Idris and Assoc. Prof. Dr. Sufizar Binti Ahmad for their valuable encouragement and support in completion of my study.

I would like to express my acknowledgement to the Ministry of Higher Education Malaysia for funding scholarship through Skim Latihan Akademik Bumiputera (SLAB) and research financial support under Multidisciplinary Research Grant (MDR) vot 1313 during my study.

I would also like to thank to Centre of Graduate Studies (CGS) and Research Management Centre (RMC), Universiti Tun Hussein Onn Malaysia (UTHM) for their financial support via grant TIER 1 (vot H135) for paper publications and conference fees.

Nevertheless, I would also like to thank the entire laboratory Assistant Engineer especially for Materials Science, Polymer Ceramic, Faculty of Mechanical and Manufacturing Engineering and laboratory of Microbiology and Food Instrumentation, Faculty of Applied Science and Technology for their guidance and technical support to utilize the laboratory equipment in accomplishing this research.

Finally, special thanks to my postgraduate colleagues and fellow friends for their sharing knowledge and encouragement during my study.



ABSTRACT

Gelatin is one of the most widely used biopolymers, derived from collagen of mammalian species that commonly apply in food and non-food industries. Since the porcine and bovine are mostly sources used from the total of the production, there is significant interest in the Halal or Kosher market. Thus, fish and other marine sources are being exploited simultaneously. However, fish gelatin has certain inferiorities compared to mammalian gelatin in terms of low gel strength and viscosity. Therefore, this study aims to extract gelatin from Black Tilapia fish skins, with a focus on extraction methods for producing high quality gelatin properties includes gel strength, viscosity, melting temperature and wettability. Gelatin was extracted by using hydrochloric acid (HCl) as a pre-treatment and thermal extraction with controlling acid concentration (0.03, 0.05, 0.1, 0.15 and 0.2 M), pre-treatment time (4, 16 and 24 h), thermal extraction temperature (45, 65 and 85 °C) and extraction time (4, 16 and 24 h). The effects of ultraviolet (UV) light on the properties of gelatin has been studied. Gelatin has been exposed to Ultraviolet A (UVA) and Ultraviolet C (UVC) for 0.5 to 2 h. The gelatin yield, gel strength, viscosity, melting temperature and amino acid composition were determined. From the obtained results, the suitable condition for pre-treatment is at 0.03 M HCl with 4 h duration. The gelatin yield, gel strength, viscosity and melting temperature of gelatin extracted at 45 °C for 4 h were found 26.35 ± 0.11 %, 286.7 ± 2.52 g, 8.43 ± 0.3 cP and 29.5 ± 0.8 °C, respectively. Gelatin has been successfully extracted from Black Tilapia skin with acid extraction. Gelatin treated with UVA and UVC has improved the gel strength (22.67 %) and viscosity (18.03 %) simultaneously. Results indicated that employing UV treatment can enhance the properties of gelatin.



ABSTRAK

Gelatin merupakan salah satu biopolymer yang paling banyak digunakan yang berasal dari kolagen spesis mamalia yang biasanya digunakan dalam industri makanan dan bukan makanan. Oleh kerana kebanyakan sumber yang digunakan dari pengeluaran gelatin adalah daripada babi dan lembu, terdapat keperluan yang besar di pasaran Halal dan Kosher. Oleh itu, ikan dan sumber laut dieksploitasi secara serentak. Walau bagaimanapun, gelatin ikan mempunyai kekurangan yang tertentu berbanding gelatin mamalia dari segi sifat seperti kekuatan gel dan kelikatan yang rendah. Oleh itu, tujuan kajian ini adalah untuk mengekstrak gelatin dari kulit ikan tilapia hitam, dengan memberi fokus kepada kaedah pengekstrakan untuk menghasilkan sifat gelatin yang berkualiti tinggi merangkumi kekuatan gel, kelikatan, suhu lebur dan kebolehbasahan. Gelatin diekstrak dengan menggunakan asid hidroklorik (HCl) sebagai pra-rawatan dan pengekstrakan terma dengan pengawalan kepekatan asid (0.03, 0.05, 0.1, 0.15 dan 0.2 M), masa pra-rawatan (4, 16 dan 24 jam), suhu pengekstrakan (45, 65 dan 85 °C) dan masa pengekstrakan (4, 16 dan 24 jam). Kesan sinar ultraungu (UV) terhadap sifat gelatin telah dikaji. Gelatin telah di dedahkan pada UVA dan UVC selama 0.5 hingga 2 jam. Keputusan hasil gelatin, kekuatan gel, kelikatan, suhu lebur dan komposisi asid amino telah ditentukan. Dari hasil yang diperolehi, keadaan yang sesuai untuk prarawatan adalah pada 0.03 M HCl dengan jangkamasa 4 jam. Hasil gelatin, kekuatan gel, kelikatan dan suhu lebur gelatin yang diekstrak pada 45 °C selama 4 jam didapati masing-masing menunjukkan 26.35 ± 0.11 %, 286.7 ± 2.52 g, 8.43 ± 0.3 cP dan 29.5 \pm 0.8 °C. Gelatin telah berjaya di ekstrak daripada kulit ikan tilapia hitam dengan pengekstrakan asid. Gelatin yang dirawat dengan UVA dan UVC dapat meningkatkan kekuatan gel (22.67 %) dan kelikatan (18.03 %) secara serentak. Hasil menunjukkan bahawa dengan menggunakan rawatan UV dapat meningkatkan sifat gelatin.



TABLE OF CONTENTS

	TITI	E		i	
	DEC	ARATION		ii	
	DED	CATION		iii	
	ACK	NOWLEDGEMENT		iv	
	ABS	RACT		v	
	TAB	E OF CONTENTS		vii	
	LIST	OF TABLES		xi	
	LIST	OF FIGURES		xiii	
	LIST	OF SYMBOLS AND	O ABBREVIATIONS	xvii	
	LIST	OF APPENDICES		XX	
	СНА	PTER 1 INTRODUC	CTION		
	1.1	Background of Stud	y	1	
	1.2	Problem Statements		3	
	1.3	Objectives		4	
	1.4	Scope of Study		4	
	1.5	Significance of Stud	У	5	
	СНА	PTER 2 LITERATU	RE REVIEW		
	2.1	Introduction		7	
	2.2	Gelatin		7	
	2.3	Halal Gelatin		8	
	2.4	The Gelatin Precurso	or Collagen	10	
	2.5	Sources of Gelatin		12	
	2.6	Collagen-Gelatin Tra	ansformation	14	
		2.6.1 Chemical Pro	ocess	14	
		2.6.2 Enzymatic Pr	rocess	16	

2.7	Gelati	n Applications	18
	2.7.1	Food Industry	20
	2.7.2	Pharmaceutical	22
	2.7.3	Cosmetic	24
	2.7.4	Engineering and Technical	24
	2.7.5	Biomedical	26
2.8	Types	of Gelatin	27
	2.8.1	Type-A Gelatin	27
	2.8.2	Type-B Gelatin	28
2.9	Proper	rties of Gelatin	29
	2.9.1	Yield	29
	2.9.2	Chemical Properties	31
	2.9.3	Physical Properties	34
		2.9.3.1 Color	34
		2.9.3.2 Viscosity	34
		2.9.3.3 Gel Strength (Bloom Value)	36
	2.9.4	Melting Temperature	38
	2.9.5	Water Contact Angle (WCA)	39
2.10	Enhan	cement Properties of Fish Gelatin	40
	2.10.1	Ultraviolet (UV) Treatment on Gelatin	40
2.11	Produc	ction of Gelatin	42
	2.11.1	Pre-treatment Process	42
		2.11.1.1 Acidic Pre-treatment	42
		2.11.1.2 Alkaline Pre-treatment	43
	2.11.2	Extraction Process	44
	2.11.3	Recovering Process	47
2.12	Types	of Fish	47
2.13	Tilapia	a Fish	48
2.14	Black	Tilapia Fish	50
2.15	Skin		51
2.16	Gelati	n from Fish	52
2.17	Extrac	tion Gelatin from fish Skin	53
	2.17.1	Extraction using Alkaline	53
	2.17.2	Extraction using Acid	54

viii



CHAPTER	3	METHO	DOLOGY
----------------	---	--------------	--------

3.1	Introdu	action	57
3.2	Raw M	laterials	57
3.3	Metho	dology Flowchart	58
3.4	Prepara	ation of Black Tilapia Fish Skin	61
3.5	Pre-tre	atment Process	62
	3.5.1	Preparation of Hydrochoric (HCl)	
		Acid Solution	63
3.6	Extract	tion Process	64
3.7	Filterir	ng Process	65
3.8	Ultravi	olet (UV) Irradiation	66
3.9	Testing	g and Characterization	67
	3.9.1	Gelatin Yield	67
	3.9.2	Viscosity Measurement	67
	3.9.3	Gel Strength / Bloom value	68
	3.9.4	Gel Melting Temperature	69
	3.9.5	Moisture Content	70
	3.9.6	Ash Content	71
	3.9.7	Protein Content	71
	3.9.8	Amino Acid Analysis	73
	3.9.9	Morphology Test	74
	3.9.10	Thermal Analysis	74
	3.9.11	Fourier Transform Infrared (FTIR)	
		Spectroscopy	75
	3.9.12	Water Contact Angle (WCA)	75

CHAPTER 4 CHARACTERIZATION OF BLACK TILAPIA SKINS

4.1	Introduction	77
4.2	Parts of Black Tilapia Fish Skin	78
	4.2.1 Skin from Wastes	80
4.3	Microstructure of fish Skins	82
4.4	Proximate Composition of Black Tilapia Fish Skins	84
	4.4.1 Moisture Content	85

	4.4.2	Protein Content	86
	4.4.3	Ash Content	87
4.5	Therm	ogravimetric Analysis (TGA)	88
4.6	Effect	s of Pre-treatment Conditions on Black	
	Tilapia	a Skins	89
	4.6.1	Protein Analysis	89

CHAPTER 5 EFFECTS OF THERMAL EXTRACTION ON PROPERTIES OF FISH GELATIN

5.1	Introduction	94
5.2	Gelatin Yield	95
5.3	Color of Gelatin	95
5.4	Gel Strength (Bloom Value) after Thermal Extraction	98
5.5	Viscosity of Gelatin after Thermal Extraction	101
5.6	Gel Melting Temperature	103
5.7	Bonding Analysis after Thermal Extraction	105
5.8	Amino Acid Composition	110

CHAPTER 6 EFFECTS OF ULTRAVIOLET (UV)

TREATMENT

6.1	Introduction	113
6.2	S Gel Strength after UV Treatment	114
6.3	Viscosity of Gelatin after UV Treatment	117
6.4	Melting Temperature of Gelatin after UV Treatment	119
6.5	Bonding Analysis of Gelatin after UV Treatment	120
6.6	Water Contact Angle (WCA) after UV Treatment	122

CHAPTER 7 CONCLUSION AND RECOMMENDATIONS

7.1	Conclusion	129
7.2	Recommendations	130
REFE	RENCES	131
APPE	NDICES	144
LIST OF PUBLICATIONS		149
LIST	OF COMPETITIONS	151
VITA		152

LIST OF TABLES

1.1	Sources of marine collagen and gelatin	3
2.1	Classification of collagen	10
2.2	Sources of gelatin	13
2.3	Types of chemicals used for acid and alkaline process	16
2.4	Types of chemicals used for enzymatic process	17
2.5	Applications of gelatin in the market	18
2.6	The recommended of gelatin use and bloom value in	
	food ingredients	21
2.7	Functional properties of gelatin in foods	22
2.8	Applications of gelatin in the pharmaceutical industry	23
2.9	Applications of gelatin in engineering industry	25
2.10	Differentiations of gelatin for type A and type B	27
2.11	Types of chemicals used for gelatin type A for	
	mammalian and fish	27
2.12	Types of acid used for gelatin type B for mammalian	
	and fish	28
2.13	Yield of fish gelatin from various type of fish	30
2.14	Classifications of amino acids	31
2.15	Amino acid composition of collagen and gelatin from	
	fish skin	32
2.16	Viscosity of various fish gelatin	35
2.17	Gel strength of various fish gelatin	37
2.18	Melting point of various fish gelatin	38
2.19	Water contact angle of various fish gelatin	40
2.20	Method used for extraction process of fish gelatin	46



2.21	Properties of fish layer	52
2.22	Gelatin yield obtained from selected fish skin	53
2.23	Alkaline extraction	54
2.24	Acid extraction	55
3.1	Description of process of producing fish gelatin in this	
	study	60
3.2	Summary of parameters for pre-treatment process	62
3.3	Volume of concentrated HCl needed to achieve desired	
	HCl solution	63
3.4	Parameter for extraction process	64
3.5	Parameters irradiation treatment for UV	66
3.6	Parameters for gel strength analysis	69
3.7	Operating parameters for SEM	74
4.1	Comparison characteristic of raw fish skin	82
4.2	Comparison summary of moisture content	86
4.3	Comparison summary of protein content	87
4.4	Comparison summary of ash content	88
5.1	Color of black tilapia skin at different conditions	97
5.2	FTIR spectra peak locations for black tilapia skin	
	gelatin extracted at various conditions	109
5.3	Amino acids composition of black tilapia skin gelatin	111
5.4	Properties of black tilapia skin gelatin extracted at	
	45°C for 4 h	112
6.1	Comparison of gel strength exposed UV irradiation for	
	mammalian and fish gelatin	117
6.2	FTIR spectra peak locations for black tilapia skin	
	gelatin treated with UVA at different irradiation time	121
6.3	FTIR spectra peak locations for black tilapia skin	
	gelatin treated with UVC at different irradiation time	122
6.4	Water droplet image for gelatin treated with UVA and	
	UVC at various irradiation time	124

LIST OF FIGURES

2.1	Malaysia Halal Logo	8
2.2	Halal certification process guide	9
2.3	Collagen structure (polypeptide strands) in fish gelatin	11
2.4	Collagen structure	11
2.5	Global gelatin market by source	13
2.6	Denaturation process of collagen into gelatin	14
2.7	General methods for acid and alkaline of gelatin	
	extraction	15
2.8	General methods of enzymatic process for fish gelatin	15 17
	extraction	17
2.9	Global fish gelatin market size by product 2013-2024	19
2.10	Global fish gelatin market size by applications	20
2.11	Gelatin as texturing agent in confectionery	21
2.12	Gelatin capsules (a) Hard capsule (b) Soft capsule	23
2.13	Cosmetic products	24
2.14	Gelatin as a binder	25
2.15	Gelatin for wound dressing	26
2.16	Four most amino acids present in gelatin	33
2.17	Basic structure of amino acid	33
2.18	Effects of gel strength (g) by gelatin concentration (%)	
	at 10°C	36
2.19	Electromagnetic radiation spectrum with visible and	
	UV light division	41
2.20	Mechanism of denaturation triple helix structure of	
	collagen to random coil of gelatin	45
2.21	Black Tilapia	49
2.22	Red Tilapia	49



2.23	Blue Tilapia	50
2.24	Parts of Black Tilapia	50
2.25	Layers of fish skins	51
3.1	Black tilapia fish skin from fillet factory after cleaning	
	process	58
3.2	Experimental flowchart of producing fish gelatin	59
3.3	Experimental flowchart for preparation of black tilapia	
	fish skin for characterization	61
3.4	Schematic for pre-treatment process	63
3.5	Schematic for extraction process	64
3.6	Schematic for filtering process	65
3.7	Schematic diagram of UV treatment	66
3.8	Schematic of texture analyzer for gel strength test	68
3.9	Position of dye at initial temperature and melting	
	temperature	70 72
3.10	Schematic of Kjeldahl digestion method	72
3.11	Schematic of distillation and titration method	73
3.12	Schematic of water contact angle machine	75
3.13	Schematic of water drop for contact angle	76
4.1	Structure of collagen fibers, fibrils, triple helical and	
	amino acids chains	78
4.2	Fresh fish of black tilapia with average weight around	
	1.2 kg	79
4.3	Weight percentage of black tilapia fish parts (wet	
	basis)	79
4.4	Black tilapia skin	80
4.5	Raw fish skin (a) Before drying (b) After drying	81
4.6	Thickness changes of fish skins	82
4.7	SEM image for external surface of tilapia skin	83
4.8	SEM image of collagen fibers	84
4.9	Proximate composition of black tilapia skin	85
4.10	TGA graph of tilapia skin	89



4.11	Effects of protein content to HCl concentration and	
	pre-treatment conditions on black tilapia skin	91
5.1	Effects of extraction time and temperature on yield of	
	black tilapia skin gelatin	94
5.2	Commercial bovine bone gelatin	96
5.3	Typical gel strength of bovine gelatin	98
5.4	Effects of extraction temperature and time on gel	
	strength for black tilapia skin gelatin	99
5.5	Gelatin renaturation process mechanism which affected	
	to extraction temperature	100
5.6	Comparison effects of thermal extraction and time on	
	yield and gel strength for black tilapia skin gelatin	101
5.7	Effects of extraction temperature and time on viscosity	
	for black tilapia skin gelatin	102
5.8	Comparison effects of thermal extraction temperature	102
	and time on yield and viscosity for black tilapia skin	103
	gelatin	
5.9	Effects of extraction temperature and time on melting	
	temperature for black tilapia skin gelatin	104
5.10	Comparison effects of extraction temperature and time	
	on melting temperature and gel strength for black	105
	tilapia skin gelatin	
5.11	FTIR spectra of black tilapia fish skin gelatin extracted	
	at various conditions	108
6.1	Gel strength of black tilapia fish skin gelatin treated	
	with UVA and UVC at different irradiation time	114
6.2	Schematic of effects for UVA and UVC irradiation on	
	skin gelatin	116
6.3	Viscosity of black tilapia skin gelatin treated with UVA	
	and UVC at different irradiation time	118
6.4	Melting temperature of black tilapia skin gelatin treated	
	with UVA and UVC at different irradiation time	120
6.5	Water droplet image before UV treatment	123

xv

6.6	Water contact angle for gelatin treated with UVA and	
	UVC at different time irradiation	125
6.7	Relationship between gel strength and contact angle for	
	gelatin treated with UVA and UVC	126
6.8	Schematic effects of crosslinking formation due to UV	
	irradiation on the gelatin surface	127
6.9	Schematic diagram of UVA and UVC penetration on	
	the gelatin surface	128

LIST OF SYMBOLS AND ABBREVIATIONS

%	-	Percentage
°C	-	Degree Celcius
AOAC	-	Association of Official Analytical Chemists
ATR	-	Attenuated Total Reflectance
BSE	-	Bovine Spongiform Encephalopathy
BSI	-	British Standard Instituition
$C_2H_4O_2$	-	Acetic Acid
$C_3H_6O_3$	-	Lactic Acid
$C_6H_8O_7$	-	Citric Acid
CaO	-	Citric Acid Calcium Oxide
Ca(OH) ₂	-	Lime solution
CH ₃ COOH	-	Acetic Acid
СООН	-	Carboxyl
cm		Centimeter
cP	<u>p.U</u>	Centipoise
EDX	-	Energy Dispersive X-Ray Spectroscopy
FAO	-	Food and Agriculture Organization
FMD	-	Foot and Mouth Disease
FTIR	-	Fourier Transform Infrared Spectroscopy
g	-	Gram
gL ⁻¹	-	Gram per liter
GME	-	Gelatin Manufactures of Europe
GMIA	-	Gelatin Manufactures Institute of America
h	-	hour
H_2O_2	-	Hydrogen Peroxide
H_2SO_4	-	Sulphuric Acid
H ₃ PO ₄	-	Phosporic Acid

	٠	٠	٠
XV	1	1	1

HC1	-	Hydrochloric Acid
HPLC	-	High Performance Liquid Chromatography
JAKIM	-	Jabatan Kemajuan Islam Malaysia
kDa	-	kilodaltons
kg	-	Kilogram
kV	-	kilo Volts
L	-	Liter
М	-	Molar
min	-	Minute
mm	-	Millimeter
mM	-	Millimolar
MPa	-	Mega Pascal
MgSO ₄	-	Magnesium Sulfate
Ν	-	Normality
NaCl	-	Sodium Chloride
NaOH	-	Sodium Chloride Sodium Hydroxide No Carbon Required Amino
NCR	-	No Carbon Required
NH ₂	-	Amino Nanometer
nm	-	Nanometer
рН		Potential of hydrogen
rpm	-19	Revolutions per Minute
SEM	201	Scanning Electron Microscopy
Т	-	Temperature
TGA	-	Thermogravimetric Analysis
UK	-	United Kingdom
USA	-	United States of America
UTHM	-	Universiti Tun Hussein Onn Malaysia
UV	-	Ultraviolet
UVA	-	Ultraviolet A
UVB	-	Ultraviolet B
UVC	-	Ultraviolet C
WCA	-	Water contact angle
w/v	-	Weight per volume
μL	-	Microliter



μm	-	Micrometer
μΜ	-	Micromolar

LIST OF APPENDICES

APPENDIX

TITLE

PAGE

А	Table A1: Weight percentage of black tilapia	
	fish parts (wet basis)	136
	Table A2: Percentage of protein content for	
	treated skins at various conditions	136
В	Table B1: Yield of black tilapia skin gelatin	
	extracted at various conditions	137
	Table B2: Gel strength of black tilapia skin	
	gelatin extracted at various conditions	137
	Table B3: Viscosity of black tilapia skin gelatin	
	extracted at various conditions	138
	Table B4: Melting temperature of black tilapia	
	skin gelatin extracted at various conditions	138
GPU	Table C1: Gel strength of black tilapia skin	
	gelatin exposed with UVA and UVC at different	
	irradiation time	139
	Table C2: Viscosity of black tilapia skin	
	gelatin exposed with UVA and UVC at different	
	irradiation time	139
	Table C3: Melting temperature of black tilapia	
	skin gelatin exposed with UVA and UVC at	
	different irradiation time	140

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Gelatin is one of the biopolymers that widely used in food, cosmetic, pharmaceutical and photographic industries. It has the characteristic of being colourless, translucent, brittle, edible and flavourless (Arpi *et al.*, 2018). In the food industry, gelatin is mainly used in confections that provide chewiness, texture and foam stabilization while in the pharmaceutical industry, it is used in the manufacture of capsules including hard and soft capsules (Karim and Bhat, 2009). Most of the worldwide gelatin production is contributed to the food industry which consists of about 70% while 15% is used in the pharmaceutical industry, thus 10% is taken by the photographic industry and the rest is used for other industry such as cosmetics and technical applications (Ofari, 1999).

According to Tkaczewska *et al.*, 2018, the usage of gelatin in industrial applications has raised the global demand for gelatin enormously in recent years, with an expected gelatin output of 450,000 tons in 2018. Currently, only 1.5 % of gelatin is produced from fish collagen-containing raw materials, while 41 % is produced from pig-skin, 28.5 % from bovine hides and 29.5 % from bovine bones (Milovanovic, 2018). However, the use of gelatin extracted from porcine and bovine sources is restricted due to the religious issue and consider on serious risk for human health effect such as foot-and-mouth disease (FMD) and outbreaks of bovine spongiform encephalopathy (BSE) called "mad cow disease" (Ratnasari *et al.*, 2013).



Porcine and bovine gelatin sources are commonly used around the world, accounting for 98.5 % of the world's gelatin supply (Sow and Yang, 2015). These existing gelatins in Islamic countries do not comply with the criteria for the Halal market as for use or consumption of the pork-related product is prohibited (Sanaei *et al.*, 2013; Riquelme *et al.*, 2015). Issues of non-Halal gelatin in packed food and cosmetic products have recently brought up sensitive conflicts among Malaysians. Malaysia is a multiracial country in which Islam is the largest community followed by Buddhism, Christianity, Hinduism and other religion. Muslims are prohibited to consume pork-related products while Hindus and Buddhists are forbidden to consume any cow-related products due to their belief. Hence, both porcine and bovine gelatins have limitations of consumption and utilization due to religious outlook.

Due to that issue, many researchers nowadays explore other sources that can produce gelatin such as marine by-products (fish) including skins, scales and bones which can be acceptable, especially for Islam and also can be used with minimally restricted for Judaism and Hinduism (Nik Aisyah *et al.*, 2014, in *et al.* 2017). Thus, collagen and gelatin extracted from marine by-products, especially from fish waste give a better advantage and increase the economic return for the fish-processing industry (Regenstein & Zhou, 2006). Fish gelatin is free from disease and is also recognized by all religions in Malaysia as Halal gelatin.



Fish resources in Malaysia are well-known categorized into freshwater and saltwater. More than 100 million tons of various species of fish have been harvested annually throughout the world. From the data reported, tilapia fish was second harvesting about 6 million tons per year throughout the world (FAO, 2018). Fish-based industries in Malaysia, such as canned fish and fillet processing, are rapidly growing due to high consumer demands. These fish processing industries produce a high quantity of fish waste as much as 70 - 85 % of the total catch, which leads to environmental issues (Kristinsson and Rasco, 2000). Since fish skins and bones contribute almost 30 % of the total fish weight, the use of these wastes from the fishery industry is excellent for the preparation of high protein food especially, gelatin. Conversion of these wastes into value-added products has both economic and waste management benefits for the fish industry (Choi and Regenstein, 2000). The production of gelatin from fish wastes has gained considerable attention recently (Etxabide *et al.*, 2016).

Table 1.1 shows several types of sources that can be extracted to produce collagen and gelatin. There are three types of sources of marine collagen and gelatin: marine invertebrates, sea mammals and fishes.

Types of Marine Species	Species	Tissues
Invertebrate	Cuttlefish, octopus and squid Jellyfish Sea urchin Sea cucumber	Outer skin and cartilaginous tissue Exumbrella and mesogela Body wall Test (or shell) Body wall
Sea mammal	Seal and whale	Skin
Fish	Lawless fish Cartilaginous fish Bony fish	Skin, notochord and cartilaginous tissues Skin and cartilage Skin, scale, bone and swim bladder

Table 1.1: Sources of marine collagen and gelatin (Regenstein and Zhou, 2006)

This study was focused on the Black Tilapia fish which is categorized as a freshwater fish to extract the gelatin. The skins of fish were used as sources of gelatin focusing on the method of extraction and the characterization of the gelatin produced. The aim of this study is to produce a better quality of gelatin with effective methods.

1.2 Problem Statements



Gelatin extracted from fish is one of the alternative sources besides porcine and bovine. Gelatin from fish has an advantage as they are not associated with the risk of FMD and BSE. Besides, fish gelatin is acceptable for Muslims and also for Judaism and Hinduism. For example, in Malaysia, there was an issue regarding the Cadbury chocolate that contains non-halal gelatin from porcine which affected to the business and reputation of the company. Due to those issues, the demand for halal foods has created fish gelatin which can apply to related industries.

Badii and Howell, (2006) reported that the industry of fish processing produced a bulk of waste including skin, bones and scales that were cause waste and pollution (Badii and Howell, 2006). The processing of fish leads to enormous amounts of waste. About 25% of the total caught fish weight was used for the filleting industry and the rest was considered discard (Gomez-Guillen *et al.*, 2002).

The total world volume of fishing in 2018 is estimated at 179 million tons, and the figure is increasing every year (FAO, 2020). The waste from fish processing can

increase up to 85 % of the total catch as estimated (Wasswa *et. al,* 2007 and Salvatore *et. al,* 2020). According to Ahmed *et. al,* 2020, a significant percentage of waste includes skin (about 30 %), bones and scales which contain a high of collagen. Therefore, the production and use of fish as a source of gelatin seem to be very promising.

Due to that problem, this research has used waste by-products of black tilapia fish skin as a source to become a valuable halal gelatin product that helps to eliminate harmful environment and improve the quality of the fish industry. Fish skin also has received attention as an alternative raw material that has the potential for producing high-quality gelatin.

1.3 Objectives

- i. To identify suitable pre-treatment parameters (hydrochloric acid concentration and time) and thermal extraction parameters (temperature and time) for extraction of gelatin from black tilapia skin
- ii. To investigate the influence of thermal extraction time and temperature on the properties of gelatin from black tilapia skin
- iii. To determine the effects of UVA and UVC irradiation in enhancing the properties of gelatin from black tilapia skin

1.4 Scope of Study

The scopes of this research are:

The first stage involved the procedure on study the characteristics of the raw materials (Black Tilapia skins);

- i. Raw material: waste by-product (Black Tilapia skin) from fillet industry [Trapia (M) Berhad].
- Surface morphology of raw material was undergoing Scanning Electron Microscopy (SEM).
- iii. Proximate composition of raw material was analyzed according to AOAC (2000) methods.

iv. The thermal stability of raw material was analyzed by Thermogravimetric Analysis (TGA).

The second stage in this study involved the characterization of gelatin which includes the preparation of gelatin extraction;

- i. Pre-treatment process by using hydrochloric acid with concentration of 0.03, 0.05, 0.1, 0.15 and 0.2 M at various times of 4, 16 and 24 hours.
- ii. Thermal extraction of gelatin with temperatures of 45, 65 and 85°C and duration time of 4, 16 and 24 hours.
- iii. Filtration process by using Whatman filter paper No. 4.
- iv. Drying process for gelatin with oven at a temperature of 60°C for 48 h.
- v. Color determination via visual observation.
- vi. Gel strength of gelatin was determined according to British Standard (BS 757:1975).
- vii. Viscosity of gelatin was determined according to Killekar et al., 2012 method.
- v. Melting temperature of gelatin was determined according to Koli *et al.*, 2012 method.
- vi. Amino acid composition of gelatin was analyzed according to AOAC (2000) method.

The third stage in this study was involved the treatment to enhance the properties of gelatin, which includes;

- i. Irradiation via Ultraviolet A (UVA) and Ultraviolet C (UVC)
- ii. Irradiation time of 0.5, 1.0, 1.5 and 2.0 hours.

1.5 Significance of Study

This study is focused on extracting Halal gelatin which is not only acceptable to Muslim consumers, but also to all Malaysian. Fish gelatin can be used as an alternative to replace mammalian gelatin, especially gelatin that extracted from bovine, which we consider the issues of an outbreak of bovine spongiform encephalopathy (BSE) and foot-and-mouth disease (FMD). BSE are progressive neurological disorders affecting

REFERENCES

- Abdelmalek, B. E., Gomez-Estaca, J., Sila, A., Martinez-Alvarez, O., Gomez-Guillen,
 M. C., Chaabouni-Ellouz, S., and Bougatef, A. (2016). Characteristics and functional properties of gelatin extracted from squid (*Loligo vulgaris*) skin. *LWT- Food Science and Technology*, 65, 924-931.
- Actionable Data Insight and Global Market Research Reports (2019). Retrieve on November 17, 2019, from http://marketintellica.com
- Agheb, M., Dinari, M., Rafienia, M., and Salehi, H. (2017). Novel electrospun nanofibers of modified gelatin-tyrosine in cartilage tissue engineering. *Materials Science and Engineering*, C71, 240-251.
- Ahmad, M. and Benjakul, S. (2011). Characteristics of gelatin from the skin of unicorn leatherjacket (*Aluterus monocerus*) as influenced by acid pretreatment and extraction time. *Food Hydrocolloids*, 18, 100315.
- Ahmed, M., Verma, A. K., and Patel, R. (2020). Collagen extraction and recent biological activities of collagen peptides derived from sea-food waste: A review. *Sustainable Chemistry Pharmaceutical*, C71, 240-251.
- Aldana, A. A., and Abraham, G. A. (2017). Current advances in electrospun gelatinbased scaffolds for tissue engineering applications. *International Journal of Pharmaceutics*, 523(2), 441-453.
- Alfaro, A. T., Fonseca, G. G., Balbinot, E., Machado, A. and Prentice, C. (2013). Physical and chemical properties of wami tilapia skin gelatin. *Food Science and Technology*, 33(3), 592-595.
- Al-Saidi, G. S., Al-Alawi, A., Rahman, M. S. and Guizani, N. (2012). Fourier transform infrared (FTIR) spectroscopic study of extracted gelatin from shaari (*Lithrinusmicrodon*) skin: effects of extraction conditions. *Int. Food Research*, 19(3), 1167-1173.

- Ambak, M. A., Mat Isa, M., Zakaria, M. Z. and Abd Ghaffar. (2012) M. Fishes of Malaysia 2nd Edition. Terengganu: Penerbit UMT. pp. 193-194.
- Ardekani, V. S., Mahmoodani, F., See, S. F., Yusop, S. M. and Babji, A. S. (2013). Processing optimization and characterization of gelatin from catfish (*Clarias gariepinus*). Sains Malaysiana, 42(12), 1697-1705.
- Arnesen, J. A. and Gildberg, A. (2007). Extraction and characterisation of gelatin from Atlantic salmon (Salmo salar) skin. *Bioresource Technology*, 98, 53-57.
- Arpi, N., Fahrizal, Martunis and Hardianti, E. (2018). Preparation and characterization of biodegradable film based on skin and bone fish gelatin. *IOP Conference Series: Earth Environment Science*, 207, 012050.
- Avena-Bustillus, R. J., Chiou, B., Olsen, C. W., Bechtel, P. J., Olson, D. A. and McHugh, T. H. (2011). Gelation, oxygen permeability and mechanical properties of mammalian and fish gelatin films. *Food Science*, 76(7), E519-E524.
- Badii, F. and Howell, N. K. (2006). Fish gelatin: structure, gelling properties and interaction with egg albumen proteins. *Food Hydrocolloids*, 20, 630-640.
- Balti, R., Jridi, M., Sila, A., Soussi, N., Nedjar-Arroume, N., Guillochon, D. and Nasri
 M. (2011). Extraction and functional properties of gelatin from the skin of cuttlefish (*Sepia officinalis*) using smooth hound crude asid protease-aided process. *Food Hydrocolloids*, 25, 943-950.
- Benbettaieb, N., Karbowiak, T., Brachais, C. H. and Debeaufort, F. (2016). Impact of electron beam irradiation on fish gelatin film properties. *Food Chemistry*, 196, 11-18.
- Bhat, R. and Karim, A. A. (2009). Ultraviolet irradiation improves gel strength of fish gelatin. *Food Chemistry*, 113, 1160-1164.
- Bhat, R. and Karim, A. A. (2012). Towards producing novel fish gelatin films by combination treatments of ultraviolet radiation and sugars (ribose and lactose) ad cross-linking agents. *Food Science Technology*, 51(7), 1326-1333.
- Boran, G. and Regenstein, J. M. (2009). Optimization of gelatin extraction from silver carp skin. *Food Science*, 74(8), 432-441.
- British Standard Institution (1975). *Methods for Sampling and Testing Gelatin* (*Physical and Chemical Methods*). London: BSI 757:1975.

- Chandra, M. V., and Shamasundar, B. A. (2015). Rheological properties of gelatin prepared from the swim bladders of freshwater fish Catla catla. *Food Hydrocolloids*, 48, 47-54.
- Chen, Y., Lu, W., Guo, Y., Zhu, Y., Lu, H. and Wu, Y. (2018). Superhydrophobic coatings on gelatin-based films: fabrication, characterization and cytotoxicity studiest. *The Royal Society of Chemistry*, 8, 23712-23719.
- Choi, S. S. and Regenstein, J. M. (2000). Physicochemical and sensory characteristics of fish gelatin. *Food Science*, 65, 279-286.
- Cihar, J.(1991). Freshwater Fish. Czech Republic: Aventium Publishing House.
- de Wolf, F. A. (2010). Collagen and gelatin. Biotechnology, 133-218.
- Da Silva, R. S. G. and Pinto, L.A.A. (2012). Physical cross-linkers: Alternatives to improve the mechanical properties of fish gelatin. *Food Engineering Reviews*, 4, 165-170.
- Damrongsakkul, S., Ratanathammapan, K., Komolpis, K. and Tanthapanichakoon, W. (2008). Enzymatic hydrolysis of rawhide using papain and neutrase. *Journal of Industrial and Engineering Chemistry*, 14, 202-206.
- Derkach, S. R, Kuchina, Y. A., Baryshnikov, A. V., Kolotova, D. S., and Voron'ko, N. G. (2019). Tailoring Cod gelatin structure and physical properties with acis and alkaline extraction. *Journal of Polymers*, 11, 1724-1741.
- Elliot, D. G. Functional morphology of the integumentary system in fishes. In Farrell,A. P., (2011). (Ed.). Encyclopedia of fish physiology: From genome toenvironment. USA: Elsevier Inc. pp. 476-488.
- Erge, A., and Zorba, O. (2018). Optimization of gelatin extraction from chicken mechanically deboned meat residue using alkaline pre-treatment. *Journal of Food science and Technology*, 97, 205-212.
- Etxabide, A.,Leceta, I., Cabezudo, S., Guerrero, P and de la Caba, K. (2016). Sustainable fish gelatin fils: from food processing waste to compost. *Sustainable Chemistry and Engineering*, 65, 279-286.
- FAO, Food and Agriculture Organisation of the United Nations. (2006). The Total Fish Production Statistics of the World. Fishery information, data and statistics unit. Rome, Italy.
- FAO, Food and Agriculture Organisation of the United Nations. (2019a). FAO GlobalFishery and Aquaculture Production Statistics 1950-2017. Rome, Italy.

- FAO, Food and Agriculture Organisation of the United Nations. (2019b). Top 10 Species Groups in Global Aquaculture 2017. Rome, Italy.
- FAO, Food and Agriculture Organisation of the United Nations. (2020). The State of World Fisheries and Aquaculture 2020. Sustainability in Action. Rome, Italy.
- Fernandez-Diaz, M. D., Montero, P. and Gomez-Guillen, M. C. (2003). Effect of freezing fish skins on molecular and rheological properties of extracted gelatin. *Food Hydrocolloids*, 17, 281-286.
- Giminez, B., Turnay, J., Lizarbe, M. A., Montero, P. and Gomez-Guillen, M. P. (2005). Use of lactic acid for extraction of fish skin gelatin. *Food Hydrocolloid*, 19, 941-950.
- GMIA, (2017). *Gelatin Handbook*, Gelatine Manufacturers Institute of America, Inc., New York, NY.
- Gelatin Manufacturers Association of Asia Pasific (2020). Retrieve on August 9, 2020, from http://gmap-gelatin.
- Gomez-Guillen, M. C. and Montero, P., (2001). Extraction of gelatin from Megrim (*Lepidorhombusboscii*) skins with different organic acids. *Food Science*. Vol. 66(2): 213-216.
- Gomez-Guillen, M. C., Turnay, J., Fernandez-Diaz, M. D., Ulmo, N., Lizarbe, M. A. and Montero, P., (2002). Structural and physical properties of gelatin extracted from different marine species: a comparative study. *Food Hydrocolloids*. Vol. 16: 25-34.
- Gomez-Guillen, M. C., Giminez, B. and Montero, P., (2005). Extraction of gelatin from fish skins by high pressure treatment. *Food Hydrocolloids*. Vol. 19: 923-928.
- Gomez-Guillen, M. C., Giminez, B., Lopez-Caballero, M. E. and Montero, P., (2011). Functional and bioactive properties of collagen and gelatin from alternative sources: A review. *Food Hydrocolloids*, 25, 1813-1827.
- Grossmann, S. and Bergmann, M. (1992). Process for the production of gelatin from fish skins. U.S. Patent 5,093,474.
- Gudmundsson, M. (2002). Rheological properties of fish gelatin. *Food Science*, 67, 2172-2176.
- Gudmundsson, M., and Hafsteinsson, H. (1997). Gelatin from cod skins as affected by chemical treatments. *Food Science*, 62, 37-47.

- Halal Development Corporation Berhad (2020). *Halal Certification*. Retrieve on March 16, 2017, from http://www.hdcglobal.com
- Haryati, D., Nadhifa, L., Humairah., and Abdullah, N. (2019). Extraction and characterization gelatin from Rabbitfish skin (*Siganus canaliculatus*) with enzymatic method using bromelin enzyme. *Earth and Environmental Science*, 355, 012095-012101.
- Hashim, D. M., Che Man, Y. B., Norakasha, R., Shuhaimi, M., Salmah, Y., and Shahariza, Z. A. (2010). Potential use of Fourier transform infrared spectroscopy for differentiation of bovine and porcine gelatins. *Food Chemistry*, 118, 856-860.
- Herpandi, Huda, N. and Adzitey, E. (2019). The effects of variation of acetic acid concentration on characteristic of gelatin from Milkfish skin (*Chanoschanos*). *Journal of Agriculture and Veterinary Science*, 12(5), 52-56.
- Ismail, S. and Suprayitno, F. (2011). Fish bone and scale as a potential source of Halal gelatin. *Fisheries and Aquatic Science*, 6(4), 379-389.
- Jabatan Kemajuan Islam Malaysia (JAKIM). *Manual Prosedur Pensijilan Halal Malaysia*. Third Edition. Putrajaya: Firdaus Press. 2014.
- Jafari, H., Lista, A., Siekapen, N. N., Lei Nei, P. G., Alimoradi, H. and Shavandi, A. (2020). Fish collagen: Extraction, characterization, and applications for biomaterials Engineering. *Polymers*, 12(10), 2230-2265.
- Jakhar, J. K., Reddy, A. D., Maharia, S., Devi, M., Reddy, G. V. S. and Venkateshwarlu, G. (2012). Characterizations of fish gelatin from Blackspotted Croaker (Protonibea diacanthus). *Applied Science Research*, 4(3), 1353-1358.
- Jamilah, B. and Harvinder, K. G. (2002). Properties of gelatins from skins of fish black tilapia (Oreochromismosammbicus) and red tilapia (Oreochromisnilotica). Food Chemistry, 77, 81-84.
- Jamilah, B., Tan, K. W., Umi Hartina, M. R. and Azizah, A. (2011). Gelatins from three cultured freshwater fish skins obtained by liming process. *Food Hydrocolloids*, 25, 1256-1260.
- Jang, H. J., Kim, Y. M., Yoo, B. Y. and Seo, Y. K. (2017). Wounds healing effects of human dermal components with gelatin dressing. *Journal of Biomaterials Appliations*, 0 (0), 1-9.

- Jongjareonrak, A., Benjakul, S., Visessanguan, W. and Tanaka, M. (2006). Skin gelatin from big eye snapper and brownstripe red snapper: chemical compositions and effects of microbial transglutaminase on gel properties. *Food Hydrocolloids*, 20, 1216-1222.
- Kaewruang, P., Benjakul, S., Prodpran, T. and Nalinanon, S. (2013). Psycochemical and functional properties of gelatin from the skin of Unicorn leatherjacket (*Aluterus monoceros*) as affected by extraction conditions. *Food Biosciences*, 2, 1-9.
- Karim, A. A. and Bhat, R. (2009). Fish gelatin: properties, challenges and prospects as an alternative to mammalian gelatins. *Food Hydrocolloids*, 20, 563-576.
- Kchaou, H., Benbettaieb, N., Jridi, M., Abdelhedi, O., Karbowiak, T., Brachais, C. H., Leonard, M. L., Debeaufort, F. and Nasri, M. (2018). Enhancement of structural, functionall and antioxidant properties of fish gelatin films using Maillard reactions. *Food Hydrocolloids*, 83, 326-339.
- Khiari, Z., Rico, D., Martin-Diana, A. B. and Barry-Ryan, C. (2011). The extraction of gelatin from mackerel (Scomber scombrus) heads with the use of different organic acids. *Fisheries Sciences*, 5(1), 52-63.
- Kim, T-K., Ham, Y-K., Shin, D-M., Kin, H-W., Jang, H. W., Kim, Y-B. and Choi, Y-S. C. (2019). Extraction of crude gelatin from duck skin: effects of heating methods on gelatin yield. *Poultry Sciences*, 0, 1-7.
- Kim, D. and Min, S. C. (2012). Trout skin gelatine-based edible film development. *Food Sciences*, 77(9), E240-E246.
- Kittiphattanabawon, P., Benjakul, S., Sinthusamran, S. and Kishimura, H. (2016). Gelatin from clown featherback skin: Extraction conditions. *LWT-Food Science and Technology*, 66, 186-192.
- Koli, J. M., Sagar, B. V., Kamble, R. S. and Sharangdhar, S. T. (2014). Functional properties of gelatin extracted from four different types of fishes: a comparative study. *Fundamental Applied Science*, 4, 322-327.
- Kong, J., and Yu, S. (2007). Fourier Transform Infrared Spectroscopic Analysis of Protein Secondary Structures Protein FTIR Data Analysis and Band Assignment, 39 (8), 549-559.



- Kouhdasht, A. M., Moosavi-Nasab, M., and Aminlari, M. (2018). Gelatin production using fish wastes by extracted alkaline protease from *Bacillus licheniformis*. *Journal Food Science and Technology*, 55, 5175-5180.
- Koutchma, T. N., Forny, L. J., and Moraru, C. I. (2009). Ultraviolet Light in Food Technology; Principles and Applications, 1st ed. USA: CRC Press.
- Kristinsson, H. G., and Rasco, B. A. (2020). Biochemical and functional properties of Atlantic salmon (*Salmo salar*) muscle hydrolyzed with various alkaline ptoteases. *Journal Food Agriculture and Food Chemistry*, 48, 657-666.
- Kumar, D. P., Chandra, M.V., Elavarasan, K. and Shamsasundar, B. A. (2017). Structural properties of gelatin extracted from croaker fish (Johnius sp) skin waste. *International Journal of Food Properties*, 20(3), S2612-S2625.
- Li, S. T. (2007). Biologic Biomaterials: Tissue-Derived Biomaterials (Collagen), in. Wong, J. Y., and Bronzino, J. D. *Biomaterials*. London: CRC Press. pp. 7-18.
- Limpisophon, K., Tanaka, M., Weng, W., Abe, S. and Osako, K. (2009). Characterization of gelatin films prepared under-utilized blue shark (Prionacegaluca) skin gelatin. *Food Hydrocolloids*, 23(7), 1993-2000.
- Lin, L., Regenstein, J. M., Lv, S., Lu, J. and Jiang, S. (2017). An overview of gelatin derived from aquatic animals: Properrties and modification. *Trends in Food Science and Technology*, 68, 102-112.
- Linden, G. and Lorient, D. (1999). New ingredients in food processing biochemistry and agriculture, England: Woodhead publishing Limited. pp. 201-210.
- Liu, H., Li, D. and Guo, S. (2008). Rheological properties of channel catfish (*Ictalurus punctaus*) gelatin from fish skins preserved by different methods. *Food Science and Technology*, 41(8), 1423-1430.
- Liu, L., Zhou, X., Yuan, C., Bai, Y., Lai, K., Yang, F. and Huang, Y. (2013). Characterization of tilapia (Orechromis niloticus) skin gelatin extracted with alkaline and different acid pretreatments. *Food Hydrocolloids*, 33, 336-341.
- Mahjoorian, A., Mortazavi, S. A., Tavakolipour, H., Motamedzadegan, A. and Askari,
 B. (2013). Rheological properties of skin gelatin of Beluga sturgeon (Huso Huso) from the Caspian sea. *Annals of Biological Research*, 4(7), 227-234.
- Mahmood, K., Muhammad, L., Ariffin, L., Abdul Razak, H. K. and Sulaiman, S., (2016). Review of fish gelatine extraction, properties and packaging applications. *Food Science and Quality Management*. Vol. 56: 47-59.

- Mariod, A. P., and Adam, H. F., (2013). Review: Gelatin, source, extraction and industrial applications. *Acta Sci. Pol.*. Vol. 12(2): 135-147.
- Masuelli, M. A. P., and Sansone, M. G., (2009). Hydrodynamic properties of gelatine studies from Intrinsic Viscosity Measurements. In Vebeck. J. (Ed.). Products and Applications of Biopolymers. Pp. 85-115.
- Milovanovic, I., (2018). Marine gelatin from rest raw materials. *Applied Science*.. Vol. 8: 2807.
- Mohtar, N. F., Perera, C. and Quek, S. Y., (2010). Optimization of gelatine extraction from hoki (*Macruronus novaezelandiae*) skins and measurement of gel strength and SDS-PAGE. *Food Chemistry*. Vol. 122: 307-313.
- Mohajer, S., Razaei, M. and Hosseini, S. F., (2017). Physico-chemical and microstructural properties of fish gelatin/agar bio-based blend films. *Carbohydrate Polymers*. Vol. 157: 784-793.
- Montero, P. and Gomez-Guillen, M. C., (2000). Extracting conditions for Megrim (*Lepidorhombus boscii*) skin collagen affect functional properties of the resulting gelatin. *Food Science*. Vol. 65: 434-438.
- Muyonga, J. H., (2003). *Nile perch collagen and gelatin extraction and physicchemical characterization*. University of Pretoria: PhD Thesis.
- Muyonga, J. H., Cole, C. G. B. and Duodu, K. G. (2004). Extraction and physicochemical characterization of Nile perch (*Lates niloticus*) skin and bones bone gelatin. *Food Hydrocolloids*, 18(4), 581-592.
- Muyonga, J. H., Cole, C. G. B. and Duodu, K. G. (2004b). Fourier transform infrared (FTIR) spectroscopic study of acid soluble collagen and gelatin from skins and bones of young and adult nile perch (*Lates niloticus*). *Food Chemistry*, 86(3), 325-332.
- Nagai, T. and Suzuki, N. (2000). Isolation of collagen from fish waste material skin, bone and fins. *Food Chemistry*, 68, 277-281.
- Nagarajan, M., Benjakul, S., Prodpran, T. and Songtipya, P. (2012). Characteristics and functional properties of gelatin from splendid squid (*Loligo formosana*) skin as affected by extraction temperature. *Food Hydrocolloids*, 29(2), 389-397.

- Nhari, R. M., Ismail, A. and Che Man, Y. B. (2012). Analytical methods for gelatin differentiation from bovine and porcine origins and food products. *Food Sciences*, 77(1), R42-R46.
- Niculescu, M., Gaidau, C., Chen, W., Gavrila, R., Ignat, M. and Epure, D. G. (2017). Study on obtaining and characterization of collagen films with agricultural applications. *Health and the Environment*, 5, 37-49.
- Nik Aisyah, N. M., Nurul, H., Azhar, M. E. and Fazilah, A. (2014). Poultry as an alternative source of gelatin. *Health and the Environment*, 5, 37-49.
- Ninan, G., Jose, J. and Abubacker, Z. (2011). Preparation and characterization of gelatin extracted from the skins of Rohu (*Labeo Rohita*) and Common Carp (*Cyprinus Carpio*). Food Processing and Preservation, 35(2), 143-146.
- Ninan, G., Jose, J. and Abubacker, Z. (2014). A Comparative study on the physical, chemical and functional properties of carp skin and mammalian gelatins. *Food Science Technology*, 51(9), 2085-2091.
- Nikoo, M., Xu, X., Benjakul, S., Xu, G., Ramirez-suarez, J. C., Ehsani, A. and Abbar, S. (2011). Characterization of gelatin from the skin of farmed Amur sturgeon Acipenser schrenckii. *International Aquatic Journal*, 135-145.
- Niu, L.,Zhou, X., Yuan, C., Bai, Y., Lai, K., Yang, F. and Huang, Y. (2013). Characterization of tilapia (*Oreochromis nilotica*) skin gelatin extracted with alkaline and different acid pretreatments. *Food Hydrocolloids*, 33, 336-341.

Norland, R. E. Fish gelatin, technical aspects. In Voight, M. N. and Botta, J. K. (1990).

- (Eds.). Advances in fisheries and biotechnology for increase profitability. USA: Technomic Publishing Co. pp. 325-332.
- Norziah, M. H., Kee, H. Y. and Norita, M. (2014). Response surface optimization of bromelain-assisted gelatin extraction surimi processing wastes. *Food Biosience*, 5, 9-18.
- Nurilmala, M., Jacoeb, A. M., Hidayat, T. and Suryamarevita, H. (2020b). Gelatin production from fish skin. Indonesian Patent. P00201709752.
- Ofori, R. A., (1999). *Preparation of gelatin from fish by enzyme aided process*. McGill University Canada: Master Thesis.
- Okuda, M., Takeguchi, M., Tagaya, M., Tonegawa, T., Hashimoto, A., Hanagata, N., and Ikoma, T. (2009). Elemental distribution analysis of type I collagen fibrils

in tilapia fish scale with energy-filtered transmission electron microscope. *Micron (Oxford, England : 1993),* 40(5-6), 665-668.

- Otoni, C. G., Avena-Bustillos, J., Chiou, B. S., Bilbao-Sainz, C., Bechtel, P. J., and McHugh, T. H. (2012). Ultraviolet-B radiation induced cross-linking improves physical properties of cold- and warm-water fish gelatin gels and films. *Food Science*, 77(9), E215-E223.
- Pranoto, Y., Lee, C. M., and Park, H. J. (2007). Characterization of fish gelatin films added with gellan and κ-carragenan. *Food Science and Technology*, 40(5), 766-774.
- Pranoto, Y., Marseno, D. W., and Rahmawati, H. (2011). Characteristic of gelatins extracted from fresh and sun-dried seawater fish in Indonesia. *International Food Research*, 18(4), 1335-1341.
- Rasli, H. I. and Sarbon, N. M. (2018). Optimization of enzymatic hydrolysis conditions and characterization of Shortfin scad (*Decapterus* Macrosoma) skin gelatin hydrosylate using response surface methodology. *International Food Research Journal*, 25(4), 1541-1549.
- Ratnasari, I., Yuwono, S. S., Nusyam, H. and Widjanarko, S. B. (2013). Extraction and characterization of gelatin from different fresh water fishes as alternative sources of gelatin. *International Food Research*, 20, 3085-3091.
- Regenstein, J. M. and Zhou, P. (2006). Collagen and gelatin from marine by-products, in. Shahidi, F. *Maximising the value of marine by-products*. England: Woodhead publishing Limited. pp. 279-303.
- Riaz, M. N. and Chaudry, M. M. (2003). *Halal Food Production*. CRC Press USA pp 1-103.
- Riquelme, N., Diaz-Calderon, P., Enrione, J. and Matiacevich, S. (2015). Effect of physical state of gelatin-plasticizer based films on the occurrence of Maillard reactions. *Food Chemistry*, 175, 478-484.
- Salvatore, L., Gallo, N., Natalli, M. L., Campa, L., Lunetti, P., Madaghiele, M., Blasi, F. S., Carolla, A., Capobianco, L., and Sannino, A. (2020). Marine collagen and its derivatives: Versatile and sustainable bio-resources for healthcare. *Materials Science Engineering*, 113, 110963.

- Sanaei, A., Mahmoodani, F., See, S., Yusop, S. and Babjil, A. (2013). Optimization of gelatin extraction and physico-chemical properties of catfish (*Carias gariepinus*) bone gelatin. *International Food Research Journal*, 20(1), 423-430.
- Sarabia, A. I., Gomez-Guillen, M. C. and Montero, P. (2000). The effect of added salts on the viscoelastic properties of fish skin gelatin. *Food Chemistry*, 70(1), 71-76.
- Sarbon, N. M., Cheow, C. S., Kyaw, Z. W. and Howell, N. K. (2014). Effects of different types and concentration of salts on the rheological and thermal properties of sin croaker and shortfin scad skin gelatin. *Int. Food Research*, 21(1), 317-324.
- Senthil R., I., Vedakumari, S. W., Hemalatha T., Sumathi V., Gobi N. and Sastry, T.P. (2016). New approaches for the effective utilization of molecular fish skin wastes of Aluterus monoceros. *Earth Environment and Health Science*, 2, 50-55.
- Shen X. R., I., Kurihara, H. and Takahashi, K. (2007). Characterization of molecular species of collagen in scallop mantle. *Food Chemistry*, 102(4), 1187-1191.
- Shyni, K., Hema, G. S., Ninan, G., Mathew, S., Joshy, C. G. and Lakshmanan, P. T.
 (2014). Isolation and characterization of gelatin from the skins of skipjack tuna (*Katsuwonuspelamis*), dog shark (*Scoliodonsorrakowah*) and rohu (*Labeorohita*). Food Hydrocolloids, 39, 68-76.
- Skomal, G. (1997). External features of a freshwater fish, in. *Setting up a Freshwater Aquarium*. New York: Howell Book House. pp. 4
- See, S. F., Hong, P. K., Ng, K. L., Wan Aida, W. M. and Babji, A. S. (2010). Physicochemical properties of gelatins extracted from skins of different freshwater fish species. *Int. Journal of Biological Macromolecules*, 73C, 146-153.
- Sinthusamran, S., Benjakul, S. and Kishimura, H. (2014b). Molecular characteristics and properties of gelatin from skin of seabass with different sizes. *Int. Food Research*, 17, 809-816.
- Sockalingam, K. Extraction and Characterizations of Halal Gelatin Biopolymer from Black Tilapia Bones and Scales. Thesis Ph.D. Universiti Tun Hussein Onn Malaysia; 2017.

- Sow, L. C. and Yang, H. (2015). Effects of salt and sugar addition on the physiochemical properties and nanostructure of fish gelatin. *Food Hydrocolloids*, 45, 72-82.
- Tabarestani, H. S., Maghsoudlou, Y., Motamedzadegan, A. and Mahoonak, A. R. S (2010). Optimization of physic-chemical properties of gelatin extracted from fish skin of rainbow trout (*Onchorhynchus mykiss*). *Bioresource Technology*, 101, 6207-6214.
- Tkaczewska, J., Morawska, M., Kulawik, P. and Zajac, M. (2018). Characterization of carp (*Cyprinus carpio*) skin gelatin extracted using different pretreatment methods. *Food Hydrocolloids*, 81, 169-179.
- Tong, Y. and Ying, T. (2013). Gelling strength improvement and characterization of a gelatin from (Aristichthys nobilis). *Food, Agriculture and Environment*, 11(1), 146-150.
- Trakul, P. and Patcharin, R. (2012). Qualities of gelatin from Thai panga fish skin as affected by skin pretreatment. *Asian Food and Agro-Industry*, 5(06), 512-520.
- Tronci, G. (2010). Synthesize, Characterization, and Biological Evaluation of Gelatin-based Scaffolds. University of Postdam Germany: Ph.D. Thesis.
- Tumerkan, E. T. A., Cansu, U., Boran, G., Regenstein, J. M. and Ozogul, F. (2019). Physiochemical and functional properties of gelatin obtained from tuna, frog and chicken skins. *Food Chemistry*, 287, 273-279.
- Vaz, C. M., De Graaf, D., Reis, R. L. and Cunha, A. M. (2003). Effects of crosslinking thermal treatment and UV irradiation on the mechanical properties and in vitro degradation behavior of several natural proteins aimed to be used in the biomedical field. *Materials Science: Materials in Medicine*, 14, 789-796.
- Wang, H. A. (2021). A review of the effects of collagen treatment in clinical studies. *Polymers*, 13, 3868-3887.
- Weng, W., Zheng, H., and Su, W. (2014). Characterization of edible films based on tilapia (*Tilapia-zillii*) scale gelatin with different extraction pH. Food Hydrocolloids, 41, 19-26.
- Wu, C. K., Tsai, J. S., and Sung, W. C. (2019). Functional characteristics of ultravioletirradiated tilapia fish skin gelatin. *Molecules*, 24(2), E254-E265.



- Wu, C. K., Tsai, J. S., Chen, Z. Y., and Sung, W. C. (2014). UV treatments on the physicochemical properties of tilapia skin and pig skin gelatin. *International Food Science and Technology*, 0(0), 1-11.
- Yang, H., Wang, Y., Zhou, P., and Regenstein, J. M. (2008). Effects of alkaline and acid pretreatment on the physical properties and nanostructures of the gelatin from channel catfish skins. *Food Hydrocolloids*, 22, 1541-1550.
- Yang, J. -I., Liang, W. -S., Chow, C. -J. and Siebert, K. J. (2009). Process for the production of tilapia retorted skin gelatin hydrolysates with optimized antioxidative properties. *Process Biochemistry*, 44(10), 1152-1157.
- Yao, Y., Ding, D., Shao, H., Peng, Q., and Huang, Y. (2017). Antibacterial activity and physical properties of fish gelatin-chitosan edible films supplemented with D-limonene. *Int. Journal of Polymer Science*, 2017, 1-9
- Yudhistira, B., Palupi, E., and Atmaka, W. (2019). The effect of acid concentration and duration of submersion toward the characteristics gelatin Eel fish bone (*Anguillar bicolor*) produced through acid process. *Earth and Environmental Science*, 246, 012046-012052
- Zhang, S., Li, L., and Kumar, A. (2008). *Material Characterization Techniques*. CRC Press.
- Zhou, P., and Regenstein, J. M. (2005). Effects of alkaline and acid pre-treatments on Alaska Pollock skin gelatin extraction. *Journal of Food Science*, 70, 392-396.

VITA

The author was born on 1st May 1981 in Muar, Johor. She went for secondary education in Sekolah Menengah Teknik Johor Baru, Johor. She pursued her higher education at the Universiti Teknologi MARA (UiTM), Selangor, and graduated with the Diploma in Industrial Chemistry in 2000. Upon graduation, she continue her degree and graduated in B. Eng (Hons) in Chemical Engineering at the same institution in October 2005. After graduation, she worked as a Laboratory Executive in Cylin Cosmetic Sdn. Bhd., Muar and then moved to Kotra Pharma (M) Sdn. Bhd. in Melaka worked as a Chemist. In June 2009 she got the opportunity to work at Universiti Tun Hussein Onn Malaysia (UTHM), Johor as a tutor at Faculty of Mechanical & Manufacturing Engineering. Realized the importance of knowledge level as an academician, she then pursued her studies at the University Kebangsaan Malaysia (UKM), Selangor and she was awarded the M. Eng. in Mechanics in 2012. In June 2014, she decided to further pursue her studies at UTHM to complete her Ph.D. in Mechanical Engineering under Department of Materials Engineering and Design. Her field of studies is in biomaterials engineering which focusing in extraction and characterization of gelatin from fish wastes. During her Ph.D. journey, she had produces 3 technical papers. She is a currently a member of Board of Engineers Malaysia (BEM) and Institute of Engineers Malaysia (IEM).

