

**EXTRACTION OF ANTI-TUBERCULOSIS PROPERTIES FROM SELECTED  
MEDICINAL PLANT BY DEEP EUTECTIC SOLVENTS (DES)**

**ALI SAMI DHEYAB**

A thesis submitted in  
fulfilment of the requirements for the award of the  
Doctor of Philosophy in Science



**Faculty of Applied Sciences and Technology  
Universiti Tun Hussein Onn Malaysia**

**FEBRUARY 2023**

## **DEDICATION**

To my beloved father and mother for their immeasurable support and care. This thesis is also dedicated to my brothers and Sisters who have continually supported and believed in me. I dedicate this thesis especially to my wife, the best in the world, without your support, none of the positive things I have accomplished when we've been together would have been possible. This thesis is as much yours as it is mine. To my wonderful son who is the lights of my life and inspire me every moment of every day.



PTTAUTHN  
PERPUSTAKAAN TUNKU TUN AMINAH

## ACKNOWLEDGEMENT

All praises and thanks to the Almighty Allah (SWT), for His countless showers of blessings throughout my research work to begin and complete this study work successfully.

I am extremely grateful to my supervisor Assoc. Prof. Dr. Mohd Fadzelly Abu Bakar and co-supervisors Dr. Siti Fatimah Sabran and Dr. Mohamed Khalid Alomar for teaching and guiding me throughout the academic period of this research, and for the valuable comments and suggestions that streamlined the conduct of the study, I am extremely grateful for their vision, dynamism, sincerity, and whatever they offered me during this research work. Thanks for everything you have done for me. You have made my family proud of me and you have made me believe in myself. I will never forget your kindness and assistance. I am quite thankful to my family for their support during my Ph.D. journey they inspired me to finish this gigantic task of PhD a timely.

I also would like to express thanks to the family for their prayers and both moral and financial support. This journey would not have been completed without the support and help of my friends and colleagues.

I would like to pay gratitude to Universiti Tun Hussein Onn Malaysia (UTHM) for providing me with the opportunity, excellent environment, and support to complete this journey of research work. Lastly, I would also thank all those people who supported me and helped me directly or indirectly in any way light a candle for me along the road to accomplishing my objective.

May Almighty Allah in his infinite mercy reward everyone abundantly.

## ABSTRACT

Globally, the threat of tuberculosis (TB) continues to grow, requiring innovative techniques to find more effective treatment options. Medicinal plants of Malaysia, Middle East, and North Africa (MENA region) following: Algeria, Bahrain, Djibouti, Egypt, Iran, Iraq, Jordan, Kuwait, Lebanon, Libya, Malta, Morocco, Oman, Qatar, Saudi Arabia, Syria, Tunisia, Turkey, United Arab Emirates, Palestine, Yemen, Ethiopia, and Sudan have a significant role in providing new therapeutic remedies. Locals reported that indigenous formulations of some herbs are utilized to treat respiratory disease symptoms relate to TB disease. Deep eutectic solvents (DESs) are considered green solvents and have more significant than traditional solvents in ecosystem. The present study aims to evaluate the potential anti-TB from plants of Malaysia and MENA region extracted by DESs and exploit the traditional medicinal practices of the local people. Aqueous extracts and different types of deep eutectic solvents extracts of 5 kind plant species (*Moringa oleifera* Lam, *Premna odorata* Blanco, *Ocimum basilicum* L, *Rosmarinus officinalis* L and *Eucalyptus camaldulensis*) were screened using different assay against multidrug-resistant (MDR) *Mycobacterium tuberculosis* strain isolated from a clinical sample. The effect of the most active extracts on mycobacterial cells at the cellular level was investigated upon treatment with the crude extracts via time-kill analysis, leakage of compound was absorbing at 280nm. The *in vitro* cytotoxicity of most active extracts was evaluated against Rat Embryonic Fibroblasts (REF) cell line using MTT assay. Phytochemical profiling of the extracts was investigated by using liquid chromatography-mass spectrometry (LC-MS) in order to identify the chemical constituents present in the most active extracts. The findings revealed that DES2: Tailor extract of *R. officinalis* displayed the largest zone of inhibition (DIZ=  $17.33 \pm 1.15$  mm) and exhibited the best antimycobacterial activity with minimum inhibitory concentration (3.12 mg/mL). Followed by DES3: CHGl of *R. officinalis* showed the good minimum inhibitory concentration (MIC= 6.25 mg/mL). DES2: Tailor extract of *R. officinalis* showed the lowest minimum bactericidal concentration (MBC= 12.5 mg/mL). At 3-fold of MIC,

DES2: Tailor extract of *R. officinalis* killed the entire bacterial cell within 48 h of exposure by causing the cell lysis and releases of cell constituents when measured cell wall integrity. Cytotoxicity assay revealed that the most active DES extract was non-toxic to the REF cell line. The LC-MS analysis revealed the presence of phytoconstituents like: camphor,  $\alpha$ -pinene, quercetin, kaempferol and oleic acid that potentially contribute to the antimycobacterial activity. In conclusion, the results demonstrated that the extracts obtained by DESs have incredible potency in TB treatment and are a greener procedure than conventional extraction solvents. Further studies on the use of DESs of plant extract could lead to the development of new anti-TB drugs.



PTTA UTHM  
PERPUSTAKAAN TUNKU TUN AMINAH

## ABSTRAK

Di peringkat global, ancaman tuberkulosis (TB) terus meningkat, teknik inovatif untuk mencari rawatan yang lebih berkesan amat diperlukan. Tumbuhan ubatan Malaysia, Timur Tengah dan Afrika Utara (rantau MENA) memainkan peranan penting dalam menyediakan ubat terapeutik baharu. Penduduk tempatan juga telah melaporkan bahawa formulasi asli beberapa herba telah digunakan untuk merawat gejala penyakit pernafasan yang berkaitan dengan penyakit TB. Pelarut eutektik dalam (DES) dianggap sebagai pelarut hijau dan lebih penting daripada pelarut tradisional dalam mesra alam. Kajian ini bertujuan untuk mencari potensi tumbuhan anti-TB dari Malaysia dan rantau MENA melalui ekstrak pelarut hijau DESs dan mengeksplorasi amalan perubatan tradisional penduduk tempatan. Ekstrak akueus dan pelbagai jenis ekstrak pelarut eutektik daripada 5 jenis spesies tumbuhan telah disaring menggunakan ujian penyebaran cakera agar untuk mendapat aktiviti antimikobakteria, kepekatan perencatan minimum. Juga diukur secara spektrofotometri pada 570 nm dan kepekatan bakteria minimum terhadap strain *Mycobacterium tuberculosis* tahan multidrug (MDR) juga diasingkan daripada sampel klinikal. Kesan ekstrak paling aktif pada sel mikobakteria telah disiasat semasa rawatan dengan ekstrak mentah melalui analisis masa-mematiakan, kebocoran sebatian menyerap pada 280nm di peringkat sel. Sitotoksiti in vitro untuk kebanyakan ekstrak aktif telah dinilai terhadap garis sel Embryonic Fibroblasts Tikus (REF) melalui ujian MTT. Pemprofilan fitokimia bagi ekstrak yang telah disiasat menggunakan spektrometri jisim kromatografi cecair (LC-MS) untuk mengenal pasti juzuk kimia yang terdapat dalam ekstrak yang paling aktif. Hasil menunjukkan bahawa DES2: Ekstrak Tailor *R. officinalis* memaparkan zon perencatan terbesar ( $DIZ = 17.33 \pm 1.15$  mm) dan menujukkan aktiviti antimikobakteria terbaik dengan kepekatan perencatan minimum ( $3.12$  mg/mL). Diikuti dengan DES1: LGH dan DES3: CHGl of *M. oleifera*, DES3: CHGl of *P. odorata*, DES2: Tailor of *O. basilicum* and DES3: CHGl of *R. officinalis* menunjukkan kepekatan perencatan minimum yang baik ( $MIC = 6.25$  mg/mL). DES2: Ekstrak Tailor *R. officinalis* menunjukkan kepekatan bakteria minimum yang paling

rendah ( $MBC = 12.5 \text{ mg/mL}$ ). Pada 3 kali ganda MIC, DES2: Ekstrak Tailor *R. officinalis* membunuh keseluruhan sel bakteria dalam pendedahan selama 48 jam dengan menyebabkan lisis sel dan pelepasan konstituen sel semasa integriti dinding sel diukur. Ujian sitotoksik menunjukkan bahawa ekstrak DES yang paling aktif tidak toksik kepada garis sel REF. Analisis LC-MS mendedahkan kehadiran kandungan fitokimia: camphor,  $\alpha$ -pinene, quercetin, kaempferol dan asid oleik yang mempunyai potensi aktiviti antimikobakteria. Kesimpulannya, hasil yang didapati menunjukkan bahawa ekstrak yang diperoleh dari DES mempunyai potensi besar dalam rawatan TB dan merupakan prosedur yang lebih hijau daripada menggunakan pelarut. Kajian lanjut mengenai penggunaan DES dalam ekstrak tumbuhan boleh membawa kepada pembangunan ubat anti-TB baru.



PTTA UTHM  
PERPUSTAKAAN TUNKU TUN AMINAH

## TABLE OF CONTENTS

<b>TITLE</b>	<b>i</b>
<b>DECLARATION</b>	<b>ii</b>
<b>DEDICATION</b>	<b>iii</b>
<b>ACKNOWLEDGEMENT</b>	<b>iv</b>
<b>ABSTRACT</b>	<b>v</b>
<b>ABSTRAK</b>	<b>vii</b>
<b>TABLE OF CONTENTS</b>	<b>ix</b>
<b>LIST OF TABLES</b>	<b>xiii</b>
<b>LIST OF FIGURES</b>	<b>xv</b>
<b>LIST OF SYMBOLS AND ABBREVIATIONS</b>	<b>xvii</b>
<b>LIST OF APPENDICES</b>	<b>xx</b>
<b>LIST OF PUBLICATIONS</b>	<b>xxi</b>
<b>CHAPTER 1 INTRODUCTION</b>	<b>1</b>
1.1    Research background	1
1.2    Problem statements	4
1.3    Research objectives	5
1.4    Scope of the study	5
1.5    Significance of the study	5
<b>CHAPTER 2 LITERATURE REVIEW</b>	<b>7</b>
2.1    Tuberculosis (TB) disease	7
2.1.1    TB disease history	7
2.1.2    Classification of <i>M. tuberculosis</i>	9
2.1.3    Morphology of <i>M. tuberculosis</i>	10
2.1.4    Culture characteristics of <i>M. tuberculosis</i>	11
2.1.5    TB etiologic agent	12
2.1.6    TB disease transmission	12
2.1.7    Pathogenesis of <i>M. tuberculosis</i>	13
2.1.8    Clinical symptoms of <i>M. tuberculosis</i>	14

2.1.9	Treatment of TB	15
2.1.10	Model strain for discovery of <i>M.</i> <i>tuberculosis</i> inhibitors	17
2.1.11	Epidemiology of TB	18
2.2	Herbal medicinal plants	21
2.2.1	Ethnobotanical research	22
2.2.2	Drugs recently extracted from natural products	26
2.2.3	The major compounds from medicinal plants	34
2.2.4	Plant crude extraction	38
2.2.5	Conventional plant extraction methods	38
2.3	Deep eutectic solvents (DES)	41
2.3.1	A brief Overview	41
2.3.2	Advanced plant extraction techniques by DES	42
2.3.3	DES for extraction of bioactive compounds	46
2.3.4	Biological application of plant extracts obtained using DES	47
2.4	Phytochemical analysis techniques	49
2.4.1	High-Performance Liquid Chromatography (HPLC)	49
2.4.2	Gas Chromatography-Mass Spectrometry (GC-MS) analysis	50
2.4.3	Liquid Chromatography-Mass Spectrometry (LC-MS)	50
2.5	Bioassay Guidance for evaluating the antimycobacterial activity	51
2.5.1	Agar diffusion	51
2.5.2	Macro and micro agar dilution	52
2.5.3	Micro broth dilution	52
2.6	Effect of the extracts at a cellular level	53
2.6.1	Time-kill assay	53
2.6.2	Membrane integrity	54

2.6.3 <i>In-vitro</i> cytotoxicity	55
<b>CHAPTER 3 METHODOLOGY</b>	<b>57</b>
3.1 Selection of medicinal plant samples	57
3.2 Sample collection and preparation	58
3.3 Selection of deep eutectic solvent types	59
3.4 Deep Eutectic Solvents preparations	60
3.4.1 Chemical materials	60
3.4.2 Preparations	60
3.5 Extraction process	61
3.6 Determine of total phenolic and flavonoid contents (TPC and TFC).	61
3.7 Preparation of plant extracts/drug standard concentrations	61
3.8 Antimycobacterial bioassay	62
3.8.1 Multi drugs resistance - <i>Mycobacterium</i> <i>tuberculosis</i> strain	62
3.8.2 Determination of diameter of inhibition zones	62
3.8.3 Antimycobacterial screening by micro dilution tube test	65
3.9 Effect of the extracts at the cellular level	66
3.9.1 Time-kill assay	66
3.9.2 Cell membrane integrity analysis	66
3.10 Evaluation <i>in-vitro</i> cytotoxicity of the extracts	67
3.10.1 Cell lines and cell cultures	67
3.10.2 Cytotoxicity assay	67
3.11 Phytochemical Analysis by Liquid Chromatography-Mass Spectrometry (LC-MS)	68
3.12 Statistical analysis	68
<b>CHAPTER 4 RESULTS AND DISCUSSION</b>	<b>70</b>
4.1 Yield of the crude extract from through quantitative analysis	70
4.2 Biocompatibility	72
4.3 Antimycobacterial activity	73

4.3.1	Mycobacterial inhibition by agar disk diffusion assay	74
4.3.2	Minimum inhibition concentration (MIC) and minimum bactericidal concentration (MBC)	76
4.4	Effect of the extracts at the cellular level	81
4.4.1	Effect of selected crude extracts on growth of MDR- <i>Mycobacterium tuberculosis</i> using time kill assay	81
4.4.2	Effect of selected active extracts on cell membrane integrity of MDR- <i>Mycobacterium tuberculosis</i>	86
4.5	<i>In-vitro</i> cytotoxicity	89
4.6	Phytochemical analysis of the selected plant's crude extracts	92
4.6.1	Identification of bioactive constituents using Liquid Chromatography-Mass Spectroscopy (LC-MS) analysis	92
4.6.2	Antimycobacterial activity of the compounds identified from selected plants extract	99
<b>CHAPTER 5 CONCLUSION AND RECOMMENDATION</b>		<b>102</b>
5.1	Conclusion	102
5.2	Recommendations	103
<b>REFERENCES</b>		<b>104</b>
<b>APPENDICES</b>		<b>104</b>
<b>VITA</b>		<b>173</b>

## LIST OF TABLES

2.1	Year of discovery, characteristics and most frequent adverse effects reported of the first line and some of second line anti-TB agents	17
2.2	Some medicinal plants used against <i>M.tuberculosis</i> .	25
2.3	Different between advanced and conventional plant extraction techniques (Belwal <i>et.,al</i> 2018 )	45
3.1	List of the medicinal plants used in the study	59
3.2	List of the chemicals materials used for DESs preparations	60
3.3	Type of DESs used in this study	60
3.4	Guide for the Preparation of McFarland standards	63
4. 1	Phytochemical contents from the plants extracted through used various types of DESs	72
4.2	Influence of DESs on the MDR- <i>Mycobacterium tuberculosis</i> Inhibition (cm)	73
4.3	Diameter (mm) of inhibitory zone (DIZ) of plant crude extracts against MDR- <i>Mycobacterium tuberculosis</i>	75
4.4	Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC)	78
4.5	Phytochemical compounds identified in the DES1:LGH extracts of <i>M. oleifera</i> using LC-MS analysis	95
4.6	Phytochemical compounds identified in the DES3: CHGl extracts of <i>M. oleifera</i> using LC-MS analysis	96
4.7	Phytochemical compounds identified in the DES3: CHGl extract of <i>P. odorata</i> using LC-MS analysis	97
4.8	Phytochemical compounds identified in the DES2: Tailor extract of <i>O. basilicum</i> using LC-MS analysis	97

4.9	Phytochemical compounds identified in the DES2:Tailor extracts of <i>R. officinalis</i> using LC-MS analysis	98
4.10	Phytochemical compounds identified in the DES3:CHGl extracts of <i>R. officinalis</i> using LC-MS analysis	98
4.11	All plants extract with various DES and 3 major constituents	99



PTTA UTHM  
PERPUSTAKAAN TUNKU TUN AMINAH

## LIST OF FIGURES

2.1	<i>M. tuberculosis</i> morphology: Science Photo Library/ Alamy Stock Photo (Kessel, Rana, & Boehme, 2019).	10
2.2	Schematic representation of cell envelope proteins embedded in the <i>M. tuberculosis</i> complex cell wall (Forrellad <i>et al.</i> , 2013).	11
2.3	Shows how the transmission of tuberculosis from an infected (A) to a healthy (B) person.	13
2.4	Countries where <i>Moringa oleifera</i> has been recorded as either native or naturalized (Gandji <i>et al.</i> , 2018).	27
2.5	Geographic distribution of <i>Rosmarinus officinalis</i> according to the Global Biodiversity Information Facility (GBIF)	31
2.6	<i>Eucalyptus camaldulensis</i> (river red gum) on the Murchison River in Western Australia (by courtesy of Prof. Stephen D. Hopper).	33
2.7	Schematic diagram for some advanced extraction techniques (Belwal <i>et.,al</i> 2018 ).	45
2.8	Schematic diagram for mode of action (Kaur <i>et al.</i> , 2022).	55
3.1	Research flowchart	58
4.1	Percentage of antimycobacterial activity of crude plant extracts against MDR- <i>Mycobacterium tuberculosis</i>	77
4.2	Time-kill curves of the extracts (A) DES1: LGH extracts of <i>M. oleifera</i> , (B) DES3: CHGl extracts of <i>M. oleifera</i> , (C) DES3: CHGl extract of <i>P. odorata</i> , (D) DES2: Tailor extract of <i>O. basilicum</i> , (E) DES2: Tailor <i>R. officinalis</i> , (F) DES3: CHGl extracts of <i>R. officinalis</i> , (G) RIF at	

	X1, X2, and X3 MIC against <i>MDR-Mycobacterium tuberculosis</i>	84
4.3	The effect of selected active extracts on release of cell constituents absorbing at 280 nm from MDR- <i>Mycobacterium tuberculosis</i> .	89
4.4	In vitro cytotoxicity of the <i>M. oleifera</i> extracted on REF cell line A: DES2:LGH and B: DES3:CHGl.	90
4.5	In vitro cytotoxicity of the DES3: CHGl extract of <i>P. odorata</i> A and the DES2:Tailor extract of <i>O. basilicum</i> B on REF cell line.	91
4.6	In vitro cytotoxicity of the <i>R. officinalis</i> extracted on REF cell line A: DES2:Tailor B: DES3:CHGl.	91
4.7	LC chromatograms of <b>A.</b> DES1: LGH extracts of <i>M. oleifera</i> , <b>B.</b> DES3: CHGl extracts of <i>M. oleifera</i> , <b>C.</b> DES3: CHGl extract of <i>P. odorata</i> , <b>D.</b> DES2:Tailor extract of <i>O. basilicum</i> , <b>E.</b> DES2:Tailor extracts of <i>R. officinalis</i> and <b>F.</b> DES3: CHGl extracts of <i>R. officinalis</i> .	93
4.8	Chemical representation of quercetin	100
4.9	Chemical representation of rutin and kaempferol	100
4.10	Chemical representation of apigenin and oleic acid	101

## LIST OF SYMBOLS AND ABBREVIATIONS

<	-	Less than
%	-	Percentage
°C	-	Degree celcius
>	-	Greater than
≤	-	Less than or equal to
≥	-	Greater than or equal to
$\mu g$	-	Microgram
$\mu l$	-	Microliter
$\mu m$	-	Micrometre
1X	-	1 fold of
2X	-	2 fold of
3X	-	3 fold of
ADC	-	Albumin Dextrose Catalase
AIDS	-	Acquired Immunodeficiency Syndrome
BCG	-	Bacille Calmette Gue'r'in
BL-3	-	biosafety level 3
CFU	-	Colony Forming Unit/mL cm – Centimetres
CNS	-	Central Nervous System
DES	-	Deep eutectic solvents
DES1	-	Deep eutectic solvents 1
DES2	-	Deep eutectic solvents 2
DES3	-	Deep eutectic solvents 3
DES4	-	Deep eutectic solvents 4
DIZ	-	Diameter of inhibition zone
DMSO	-	Dimethyl sulphoxide
GC-MS	-	Gas chromatography–mass spectrometry
ChGl	-	Choline Chloride: Glycerol
ChXl	-	Choline Chloride: Xylitol

<i>ChMa</i>	-	Choline Chloride: Malic acid
<i>ChLa</i>	-	Choline Chloride: laevulimic acid
<i>GI</i>	-	Gastrointestinal
<i>H</i>	-	Hour
<i>HIV</i>	-	Human Immunodeficiency Virus
<i>HPLC</i>	-	High Performance Liquid Chromatography
<i>IM</i>	-	Intramuscular
<i>INH</i>	-	Isoniazid
<i>IV</i>	-	Intravenous
<i>JNPC</i>	-	Johor National Park Corporation
<i>LGH</i>	-	Lactic acid, Glucose and Water
<i>MBC</i>	-	Minimum Bactericidal Concentration
<i>MDR</i>	-	Multidrug-Resistant
<i>Mg</i>	-	milligram
<i>MIC</i>	-	Minimum Inhibitory Concentration
<i>Min</i>	-	minute
<i>mL</i>	-	Millilitre
<i>mm</i>	-	millimetre
<i>NA</i>	-	Not active
<i>nm</i>	-	nanometres
<i>OADC</i>	-	Oleic acid, Albumin, Dextrose and Catalase
<i>OD</i>	-	Optical density
<i>PAS</i>	-	Para-Amino Salicylic acid
<i>PYR</i>	-	Pyrazinamide
<i>RIF</i>	-	Rifampin
<i>RNA</i>	-	Ribonucleic acid
<i>Rpm</i>	-	Revolution per minute
<i>RT</i>	-	Retention time
<i>SD</i>	-	Standard deviation
<i>Tailor</i>	-	Glycerol, Xylitol and D-(-)-Fructose
<i>TB</i>	-	Tuberculosis
<i>TDM</i>	-	Trehalose dimycolate
<i>TLC</i>	-	Thin-Layer Chromatography
<i>TLR</i>	-	Toll like receptors

<i>TMM</i>	-	Trehalose monomycolate
<i>TNJER</i>	-	Taman Negara Johor Endau-Rompin
<i>UTHM</i>	-	Universiti Tun Hussein Onn Malaysia
<i>WHO</i>	-	World Health Organization
<i>XDR-TB</i>	-	Extensive drug-resistant



PTTA UTHM  
PERPUSTAKAAN TUNKU TUN AMINAH

**LIST OF APPENDICES**

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
A	Research approval letter	157
B	Herbarium specimens	160
C	Images of biocompatibility results by agar disk diffusion assay	166
D	Images of diameter of inhibition zone (DIZ) by agar disk diffusion assay	168
E	Minimum Inhibitory Concentration	170
F	Statistical analysis of diameter of inhibition zone	171



## LIST OF PUBLICATIONS

### Journals:

- i) **Dheyab, A. S.**, Bakar, M. F. A., AlOmar, M. K., & Sabran, S. F. (2020). A Review of Medicinal Plant of Middle East and North Africa (Mena) Region as Source in Tuberculosis Drug Discovery. *Saudi journal of biological sciences*, 27(9), 2457.
- ii) **Dheyab, A. S.**, Abu Bakar, M. F., AlOmar, M., Sabran, S. F., Hanafi, A. F. M., & Mohamad, A. (2021). Deep Eutectic Solvents (DESs) as Green Extraction Media of Beneficial Bioactive Phytochemicals. *Separations*, 8(10), 176.
- iii) **Dheyab, A. S.**, Ibrahim, A. J. K., Aljumily, E. K., AlOmar, M. K., Bakar, M. F. A., & Sabran, S. F. (2022). Antimycobacterial activity and phytochemical properties of *Eucalyptus camaldulensis* (eucalyptus) extracted by deep eutectic solvents. *Materials Today: Proceedings*, 65, 2738-2742.
- iv) **Dheyab, A. S.**, Kanaan, M. Q., Hussein, N. A., AlOmar, M. K., Sabran, S. F., & Abu Bakar, M. F. (2022). Antimycobacterial Activity of *Rosmarinus officinalis* (Rosemary) Extracted by Deep Eutectic Solvents. *Separations*, 9(10), 271.

# **CHAPTER 1**

## **INTRODUCTION**

The anti-tuberculosis potential of 5 selected medicinal herbs was described in this study. The components of the selected medicinal plants were extracted using highly standardized deep eutectic solvents in this study. The extracts were bioassayed against a clinical *Mycobacterium tuberculosis* strain with multi-drug resistance utilizing a conventional antimycobacterial test. The effect of the selected plant extracts on mycobacterial cells at the cellular level and their cytotoxicity were also studied. The phytochemical profiling of the selected plant extracts revealed many chemicals that may be responsible for antimycobacterial activity. The claim has been scientifically verified that the extraction of medicinal plants by deep eutectic solvents are much better than traditional solvents and their effect on *Mycobacterium tuberculosis*.

### **1.1 Research background**

Tuberculosis (TB) is a devastating and widespread infectious illness caused by a severe pathogenic bacteria known as *M. tuberculosis*, a member of the Mycobacterium family. This illness is prevalent in various countries throughout the globe because of the varying transmission levels (Abuzeid *et al.*, 2014; Akintola *et al.*, 2013). About two billion individuals, or about one-fifth of the world's population, are believed to be afflicted with tuberculosis (Kandasamy *et al.*, 2018). According to the World Health Organization's (WHO) 2016 estimate, around 10 million people worldwide are afflicted with tuberculosis (TB), and one million of those sick have died as a result of this infectious illness (Organization, 2016). Because of the increased mortality rates of this illness, TB was designated as one of the main causes of death in the African and South-East Asian regions, ranking above HIV/AIDS as one of the top causes of death

worldwide (Organization, 2016). In 2015, tuberculosis was considered one of the top 10 causes of death worldwide, and it caused more deaths than HIV/AIDS (AlMatar *et al.*, 2017). The tuberculosis bacteria belong to the Mycobacteriaceae family, and the specific characteristic of tuberculosis bacteria that distinguishes it from other bacteria, whether gram-positive or gram-negative, is the presence of long fatty acid chains in the cell envelope, which is referred to as the mycolic acid layer (Jackson, 2014).

Despite government policies aimed at preventing and controlling tuberculosis, the disease is still regarded as a significant public health problem in Malaysia, despite efforts to combat it (Rafiza, Rampal, and Tahir 2011). TB disease is considered the 5<sup>th</sup> most common infectious disease in Malaysia, and this conundrum can be attributed to the high amount of universal labourers from many countries, including Indonesia, Myanmar, and Bangladesh, who come to work in the country (Rafiza, Rampal, & Tahir, 2011). Since the 1950s, TB-infected patients have relied on the supply of anti-TB medications, which are split into three groups: Anti-TB medications in the first line, second line, and third line of defence against tuberculosis (Jetan *et al.*, 2010; Keshavjee and Farmer, 2012; Zumla, Nahid, & Cole, 2013). In order to eradicate tuberculosis, modern chemotherapeutic drugs such as isoniazid, rifampicin, ethambutol, streptomycin, and pyrazinamide are utilized. On the other hand, these medications have the undesirable consequence of generating dangerous side effects such as hepatitis, gastroenteritis, and hearing loss in certain people (Altaf *et al.*, 2010; Famewo, Clarke, & Afolayan, 2017).

There are two types of TB resistance: multidrug-resistant TB (MDR-TB), which does not respond to first-line standard treatment, and extensive drug-resistant TB (XDR-TB), which occurs when resistant to second-line TB drugs. MDR-TB is the more serious of the two types of TB resistance. According to 2012 WHO report on monitoring and response to MDR-TB and XDR-TB, 310,000 MDR-TB cases were identified among pulmonary TB patients in 2011, with 84 countries reporting at least one case of XDR-TB. 450,000 people worldwide developed MDR-TB in 2012; 170,000 people died from it. Even though more than half of these cases came from Brazil, China, Russia and India, the overall picture is still alarming (Abdallah & Ali, 2012; Rennie *et al.*, 2011). The difficulties present a unique opportunity to begin investigating novel techniques to treat multidrug-resistant tuberculosis (Robles-Zepeda *et al.*, 2013; Umesiri *et al.*, 2015). As a result, a new anti-TB drug discovery effort should be launched to address the issues mentioned above in previous subsection

and demonstrate support for sustainable development and the TB Strategy (Organization, 2016). In light of these adverse side effects of synthetic drugs, the potential efficacy of herbal medicines has piqued the interest of scientists and healthcare providers, who have begun to shift their attention away from synthetic drugs and toward conventional medicinal products for the treatment of certain chronic and infectious diseases (Adaikkappan, Kannapiran, & Anthonisamy, 2012).

Anthropological botany is the study of both people and plants. Plants are a valuable source of traditional medicines and are used for various reasons, including food, fodder, and medicine (Jalil *et al.*, 2012; Tariq *et al.*, 2015). Because of their chemical diversity and crucial role in drug discovery, medicinal plants hold tremendous promise for meeting these needs. These plants have mainly been employed as pure chemicals or as raw materials. Only a few plant species have been thoroughly researched for their therapeutic potential (Gemechu *et al.*, 2013; Kaur and Kaur, 2015). Due to the fact that they are readily available and easily harvested, plants are still being utilized for treatment reasons, namely to manage TB-related clinical signs, by indigenous people all over the world today (Akintola *et al.*, 2013; Bunalema *et al.*, 2014; Jiménez-Arellanes *et al.*, 2014) Plant-based medications have traditionally been used to treat a wide range of infectious diseases all over the world. Approximately 75% of the world's population relies on medicinal plants for their primary health care. TB treatment, in particular, is dependent on the presence of an extensive range of secondary metabolites within them (Jamal *et al.*, 2011; sharma, 2016).

Phytochemical studies of various therapeutic plants have revealed a large number of active natural chemicals; however, few plants have been tested against mycobacteria, and several plants have shown promising results against tuberculosis disease (Kaur and Kaur, 2015). Secondary metabolites of plants, also known as phytochemicals, are chemical compounds found in plants that occur naturally in their environment. This class of chemicals, in contrast to the primary metabolites, does not directly contribute to the growth, proliferation, and development of plants (Ahmed, Arshad, & Khan, 2017). Phytochemicals in plants are closely linked to biological activities, particularly antimycobacterial activity, as these chemicals play a crucial role in describing such activity (Compean and Ynalvez, 2014; Kumar, Banik, & Sharma, 2010).

Malaysia's tropical rainforest is densely planted with various species, including medicinal herbs. There are over 14,500 plant species in Malaysia, of which over 2000

have diverse therapeutic qualities and have a strong potential for commercialization (Ahmad and Othman, 2013; Jamal *et al.*, 2011).

## 1.2 Problem statements

Among all infectious diseases, tuberculosis is one of the most dangerous killers of adults in the world today. The treatment options offered have the drawback of generating significant side effects such as hepatitis, gastroenteritis, nausea, and hearing loss, amongst other unpleasant side effects. In addition, the pharmaceutical regimen is overly lengthy, lasting anywhere from 6 to 9 months. There are many people who do not stick to the time restriction for therapy because of this reason. As a result, the situation has become more complicated, with *Mycobacterium* modifying resistance to TB medications giving rise to multidrug-resistant TB (MDR-TB) and extended drug-resistant TB (XDR-TB), as well as a pressing need to find new treatments for TB disease strains. Furthermore, the existing anti-TB treatment regimen, which has produced significant side effects in patients for a long time, has necessitated the urgent development of a novel anti-TB drug of plant origin. On the other hand, there are claims for the traditional solvents that have many drawbacks such as long extraction period, toxicity, not ecofriendly, expensive, the requirement of large volume solvent. The selected medicinal plants that have been no actual scientific evaluation of the antimycobacterial activity in these plants extracted by deep eutectic solvents to justify the claim. Deep eutectic solvents (DESs) are expected to be used extensively for the extraction of bioactive compounds from medicinal plants. As a result, a study of these plants in the laboratory is required through extracted by deep eutectic solvents and anti-TB benefits medicinally. The anti-mycobacterial effect agents derived from medicinal plants have been studied extensively in various parts of the world; however, despite Malaysia's rich plant diversity, little attention has been paid to evaluating and detecting anti-mycobacterial activity from Malaysian medicinal plants in the laboratory setting.

## REFERENCES

- Abbott, A. P., Boothby, D., Capper, G., Davies, D. L., & Rasheed, R. K. (2004). Deep Eutectic Solvents Formed between Choline Chloride and Carboxylic Acids: Versatile Alternatives to Ionic Liquids. *Journal of the American Chemical Society*, 126(29), 9142-9147.
- Abdel-Latif, H. M., Abdel-Daim, M. M., Shukry, M., Nowosad, J., & Kucharczyk, D. (2022). Benefits and Applications of Moringa Oleifera as a Plant Protein Source in Aquafeed: A Review. *Aquaculture*, 547, 737369.
- Abdullah, A. K., Dheyab, A. S., & Saleh, R. O. (2021). Evaluation of Anti-Fungal Activity Derivative from Premna Odorata Blanco Extract by Deep Eutectic Solvents. *Plant Science Today*, 8(4), 1032-1036.
- Abidin, Z., Khaeriah, U., Zuhrina, Z., Pratama, M., & Baits, M. (2019). Tyrosinase Inhibitor Activity Measurement of Crude and Purified Extract of Moringa Leaves (Moringa Oleifera L.). *Indonesian Journal of Pharmaceutical Science and Technology*, 1(1), 52-58.
- Abuzeid, N., Kalsum, S., Koshy, R. J., Larsson, M., Glader, M., Andersson, H., Raffetseder, J., Pienaar, E., Eklund, D., & Alhassan, M. S. (2014). Antimycobacterial Activity of Selected Medicinal Plants Traditionally Used in Sudan to Treat Infectious Diseases. *Journal of Ethnopharmacology*, 157, 134-139.
- Adaikkappan, P., Kannapiran, M., & Anthonisamy, A. (2012). Antimycobacterial Activity of Withania Somnifera and Pueraria Tuberosa against Mycobacterium Tuberculosis H37rv. *J. Acad. Indus. Res*, 1(4), 153-156.
- Additives, E. P. o., & Feed, P. o. S. u. i. A. (2013). Scientific Opinion on the Safety and Efficacy of Betaine Anhydrous as a Feed Additive for All Animal Species Based on a Dossier Submitted by Danisco Animal Nutrition. *EFSA Journal*, 11(5), 3209.
- Aderogba, M., Madikizela, B., & McGaw, L. J. (2019). Bioactive Constituents from Malvastrum Coromandelianum (L.) Garcke Leaf Extracts. *South African Journal of Botany*, 126, 371-376.

- Adibian, F., Ghaderi, R. S., Sabouri, Z., Davoodi, J., Kazemi, M., Ghazvini, K., Youssefi, M., Soleimanpour, S., & Darroudi, M. (2022). Green Synthesis of Selenium Nanoparticles Using Rosmarinus Officinalis and Investigated Their Antimicrobial Activity. *BioMetals*, 1-12.
- Ahmad, S. (2010). Pathogenesis, Immunology, and Diagnosis of Latent Mycobacterium Tuberculosis Infection. *Clinical and Developmental Immunology*, 2011.
- Ahmad, S., & Othman, N. (2013). Strategic Planning, Issues, Prospects and the Future of the Malaysian Herbal Industry. *International Journal of Academic Research in Accounting, Finance and Management Sciences*, 3(4), 91-102.
- Ahmed, A. F., Attia, F. A., Liu, Z., Li, C., Wei, J., & Kang, W. (2019). Antioxidant Activity and Total Phenolic Content of Essential Oils and Extracts of Sweet Basil (*Ocimum Basilicum L.*) Plants. *Food Science and Human Wellness*, 8(3), 299-305.
- Ahmed, E., Arshad, M., & Khan, M. Z. (2017). Secondary Metabolites and Their Multidimensional Prospective in Plant Life. *Journal of Pharmacognosy and Phytochemistry*, 6(2), 205-214.
- Ahmed, M. M., Velayati, A. A., & Mohammed, S. H. (2016). Epidemiology of Multidrug-Resistant, Extensively Drug Resistant, and Totally Drug Resistant Tuberculosis in Middle East Countries. *International Journal of Mycobacteriology*, 5(3), 249-256.
- Ahn, S. K., Tran, V., Leung, A., Ng, M., Li, M., & Liu, J. (2018). Recombinant Bcg Overexpressing Phop-Phor Confers Enhanced Protection against Tuberculosis. *Molecular Therapy*, 26(12), 2863-2874.
- Ajanaku, C., Echeme, J., Mordi, R., Bolade, O., Okoye, S., Jonathan, H., & Ejilude, O. (2021). In-Vitro Antibacterial, Phytochemical, Antimycobacterial Activities and Gc-Ms Analyses of Bidens Pilosa Leaf Extract. *Journal of Microbiology, Biotechnology and Food Sciences*, 2021, 721-725.
- Akintola, A., Kehinde, A., Adebiyi, O., & Ademowo, O. (2013). Anti-Tuberculosis Activities of the Crude Methanolic Extract and Purified Fractions of the Bulb of Crinum Jagus. *Nigerian Journal of Physiological Sciences*, 28(2), 135–140.
- Al Muqarrabun, L. R., Ahmat, N., Aris, S. R. S., Shamsulrijal, N., Baharum, S. N., Ahmad, R., Rosandy, A. R., Suratman, M. N., & Takayama, H. (2014). A New

- Sesquiterpenoid from Scaphium Macropodium (Miq.) Beumee. *Natural product research*, 28(9), 597-605.
- Al Zuhairi, J. J. M. J., Kashi, F. J., Rahimi-Moghaddam, A., & Yazdani, M. (2020). Antioxidant, Cytotoxic and Antibacterial Activity of Rosmarinus Officinalis L. Essential Oil against Bacteria Isolated from Urinary Tract Infection. *European Journal of Integrative Medicine*, 38, 101192.
- Aldarkazali, M., Rihan, H. Z., Carne, D., & Fuller, M. P. (2019). The Growth and Development of Sweet Basil (*Ocimum Basilicum*) and Bush Basil (*Ocimum Minimum*) Grown under Three Light Regimes in a Controlled Environment. *Agronomy*, 9(11), 743.
- Alderwick, L. J., Harrison, J., Lloyd, G. S., & Birch, H. L. (2015). The Mycobacterial Cell Wall—Peptidoglycan and Arabinogalactan. *Cold Spring Harbor perspectives in medicine*, 5(8), a021113.
- Ali, N., Ahmed, G., Ali Shah, S. W., Shah, I., Ghias, M., & Khan, I. (2011). Acute Toxicity, Brine Shrimp Cytotoxicity and Relaxant Activity of Fruits of Callistemon Citrinus Curtis. *BMC complementary and alternative medicine*, 11(1), 1-8.
- AlMatar, M., AlMandal, H., Var, I., Kayar, B., & Köksal, F. (2017). New Drugs for the Treatment of Mycobacterium Tuberculosis Infection. *Biomedicine & Pharmacotherapy*, 91, 546-558.
- AlOmar, M. K., Hayyan, M., Alsaadi, M. A., Akib, S., Hayyan, A., & Hashim, M. A. (2016). Glycerol-Based Deep Eutectic Solvents: Physical Properties. *Journal of Molecular Liquids*, 215, 98-103.
- Alsarhan, A., Sultana, N., Al-Khatib, A., & Kadir, M. R. A. (2014). Review on Some Malaysian Traditional Medicinal Plants with Therapeutic Properties. *Journal of Basic and Applied Sciences*, 10, 149-159.
- Altaf, M., Miller, C. H., Bellows, D. S., & O'Toole, R. (2010). Evaluation of the Mycobacterium Smegmatis and Bcg Models for the Discovery of Mycobacterium Tuberculosis Inhibitors. *Tuberculosis*, 90(6), 333-337.
- Altıok, E., Bayçın, D., Bayraktar, O., & Ülkü, S. (2008). Isolation of Polyphenols from the Extracts of Olive Leaves (*Olea Europaea L.*) by Adsorption on Silk Fibroin. *Separation and Purification Technology*, 62(2), 342-348.
- Altyar, A. E., Ashour, M. L., & Youssef, F. S. (2020). Premna Odorata: Seasonal Metabolic Variation in the Essential Oil Composition of Its Leaf and

- Verification of Its Anti-Ageing Potential Via in Vitro Assays and Molecular Modelling. *Biomolecules*, 10(6), 879.
- Alvin, A., Miller, K. I., & Neilan, B. A. (2014). Exploring the Potential of Endophytes from Medicinal Plants as Sources of Antimycobacterial Compounds. *Microbiological research*, 169(7-8), 483-495.
- Ameer, K., Shahbaz, H. M., & Kwon, J. H. (2017). Green Extraction Methods for Polyphenols from Plant Matrices and Their Byproducts: A Review. *Comprehensive Reviews in Food Science and Food Safety*, 16(2), 295-315.
- Aminah, N. S., Kristanti, A. N., & Tanjung, M. (2014). Antioxidant Activity of Flavonoid Compounds from the Leaves of Macaranga Gigantea. *Journal of Chemical and Pharmaceutical Research*, 6(6), 688-692.
- Antonescu, A.-I., Miere, F., Fritea, L., Ganea, M., Zdrinca, M., Dobjanschi, L., Antonescu, A., Vicas, S. I., Bodog, F., & Sindhu, R. K. (2021). Perspectives on the Combined Effects of Ocimum Basilicum and Trifolium Pratense Extracts in Terms of Phytochemical Profile and Pharmacological Effects. *Plants*, 10(7), 1390.
- Araujo, R. G., Rodríguez-Jasso, R. M., Ruíz, H. A., Govea-Salas, M., Pintado, M., & Aguilar, C. N. (2021). Recovery of Bioactive Components from Avocado Peels Using Microwave-Assisted Extraction. *Food and Bioproducts Processing*, 127, 152-161.
- Arya, V. (2011). A Review on Anti-Tubercular Plants. *Int J Phar Tech Res*, 3, 872-880.
- Ashokkumar, M. (2015). Applications of Ultrasound in Food and Bioprocessing. *Ultrasonics sonochemistry*, 25, 17-23.
- Askun, T., Tekwu, E. M., Satil, F., Modanlioglu, S., & Aydeniz, H. (2013). Preliminary Antimycobacterial Study on Selected Turkish Plants (Lamiaceae) against Mycobacterium Tuberculosis and Search for Some Phenolic Constituents. *BMC complementary and alternative medicine*, 13(1), 365.
- Askun, T., Tumen, G., Satil, F., Modanlioglu, S., & Yalcin, O. (2012). Antimycobacterial Activity Some Different Lamiaceae Plant Extracts Containing Flavonoids and Other Phenolic Compounds. *Understanding Tuberculosis—New Approaches to Fighting Against Drug Resistance*, 14, 309-336.

- Asres, K., Bucar, F., Edelsbrunner, S., Kartnig, T., Höger, G., & Thiel, W. (2001). Investigations on Antimycobacterial Activity of Some Ethiopian Medicinal Plants. *Phytotherapy Research*, 15(4), 323-326.
- Atanasov, A. G., Waltenberger, B., Pferschy-Wenzig, E.-M., Linder, T., Wawrosch, C., Uhrin, P., Temml, V., Wang, L., Schwaiger, S., & Heiss, E. H. (2015). Discovery and Resupply of Pharmacologically Active Plant-Derived Natural Products: A Review. *Biotechnology advances*, 33(8), 1582-1614.
- Avoi, R., & Liaw, Y. C. (2021). Tuberculosis Death Epidemiology and Its Associated Risk Factors in Sabah, Malaysia. *International journal of environmental research and public health*, 18(18), 9740.
- Azaizeh, H., Fulder, S., Khalil, K., & Said, O. (2003). Ethnobotanical Knowledge of Local Arab Practitioners in the Middle Eastern Region. *Fitoterapia*, 74(1-2), 98-108.
- Aziah, A. (2004). Tuberculosis in Malaysia: Combating the Old Nemesis. *Med J Malaysia*, 59(1), 1-4.
- Azmir, J., Zaidul, I., Rahman, M., Sharif, K., Mohamed, A., Sahena, F., Jahurul, M., Ghafour, K., Norulaini, N., & Omar, A. (2013). Techniques for Extraction of Bioactive Compounds from Plant Materials: A Review. *Journal of Food Engineering*, 117(4), 426-436.
- Azwanida, N. (2015). A Review on the Extraction Methods Use in Medicinal Plants, Principle, Strength and Limitation. *Med Aromat Plants*, 4(196), 2167-0412.1000196.
- Bakirtzi, C., Triantafyllidou, K., & Makris, D. P. (2016). Novel Lactic Acid-Based Natural Deep Eutectic Solvents: Efficiency in the Ultrasound-Assisted Extraction of Antioxidant Polyphenols from Common Native Greek Medicinal Plants. *Journal of Applied Research on Medicinal and Aromatic Plants*, 3(3), 120-127.
- Balasubramanian, V., Pavelka Jr, M. S., Bardarov, S. S., Martin, J., Weisbrod, T. R., McAdam, R. A., Bloom, B. R., & Jacobs Jr, W. R. (1996). Allelic Exchange in Mycobacterium Tuberculosis with Long Linear Recombination Substrates. *Journal of bacteriology*, 178(1), 273-279.
- Balunas, M. J., & Kinghorn, A. D. (2005). Drug Discovery from Medicinal Plants. *Life sciences*, 78(5), 431-441.

- Banu, K. S., & Cathrine, L. (2015). General Techniques Involved in Phytochemical Analysis. *International Journal of Advanced Research in Chemical Science*, 2(4), 25-32.
- Barbieri, J. B., Goltz, C., Cavalheiro, F. B., Toci, A. T., Igarashi-Mafra, L., & Mafra, M. R. (2020). Deep Eutectic Solvents Applied in the Extraction and Stabilization of Rosemary (*Rosmarinus Officinalis* L.) Phenolic Compounds. *Industrial Crops and Products*, 144, 112049.
- Barnes, J., & Ernst, E. (1998). Traditional Herbalists' Prescriptions for Common Clinical Conditions: A Survey of Members of the UK National Institute of Medical Herbalists. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 12(5), 369-371.
- Bastos Záchia, N. R., Piana, M., dos Santos Siqueira, F., da Silva Jesus, R., Athayde, M. L., Essi, L., & de Campos, M. M. A. (2021). Phytochemical Analysis and Evaluation of Antioxidant and Antimycobacterial Activity of *Colletia Paradoxa* from Brazil. *Natural product research*, 35(5), 802-806.
- Beatovic, D., Krstic-Milosevic, D., Trifunovic, S., Siljegovic, J., Glamoclija, J., Ristic, M., & Jelacic, S. (2015). Chemical Composition, Antioxidant and Antimicrobial Activities of the Essential Oils of Twelve *Ocimum Basilicum* L. Cultivars Grown in Serbia. *Records of Natural Products*, 9(1), 62.
- Belwal, T., Ezzat, S. M., Rastrelli, L., Bhatt, I. D., Duglia, M., Baldi, A., Devkota, H. P., Orhan, I. E., Patra, J. K., & Das, G. (2018). A Critical Analysis of Extraction Techniques Used for Botanicals: Trends, Priorities, Industrial Uses and Optimization Strategies. *TrAC Trends in Analytical Chemistry*, 100, 82-102.
- Bhat, Z. S., Rather, M. A., Maqbool, M., Lah, H. U., Yousuf, S. K., & Ahmad, Z. (2017). Cell Wall: A Versatile Fountain of Drug Targets in *Mycobacterium Tuberculosis*. *Biomedicine & Pharmacotherapy*, 95, 1520-1534.
- Bhattacharya, A., Tiwari, P., Sahu, P. K., & Kumar, S. (2018). A Review of the Phytochemical and Pharmacological Characteristics of *Moringa Oleifera*. *Journal of pharmacy & bioallied sciences*, 10(4), 181.
- Bian, G., Deng, Z., & Liu, T. (2017). Strategies for Terpenoid Overproduction and New Terpenoid Discovery. *Current opinion in biotechnology*, 48, 234-241.
- Birhanu, A. G., Yimer, S. A., Kalayou, S., Riaz, T., Zegeye, E. D., Holm-Hansen, C., Norheim, G., Aseffa, A., Abebe, M., & Tønjum, T. (2019). Ample

- Glycosylation in Membrane and Cell Envelope Proteins May Explain the Phenotypic Diversity and Virulence in the *Mycobacterium* Tuberculosis Complex. *Scientific reports*, 9(1), 2927.
- Bodnar, K. A., Serbina, N. V., & Flynn, J. L. (2001). Fate of *Mycobacterium* Tuberculosis within Murine Dendritic Cells. *Infection and immunity*, 69(2), 800-809.
- Bonah, C. (2005). The ‘Experimental Stable’of the Bcg Vaccine: Safety, Efficacy, Proof, and Standards, 1921–1933. *Studies in History and Philosophy of Science Part C: Studies in History and Philosophy of Biological and Biomedical Sciences*, 36(4), 696-721.
- Brown-Elliott, B. A., & Wallace Jr, R. J. (2002). Clinical and Taxonomic Status of Pathogenic Nonpigmented or Late-Pigmenting Rapidly Growing Mycobacteria. *Clinical microbiology reviews*, 15(4), 716-746.
- Brown, A. K., Papaemmanouil, A., Bhowruth, V., Bhatt, A., Dover, L. G., & Besra, G. S. (2007). Flavonoid Inhibitors as Novel Antimycobacterial Agents Targeting Rv0636, a Putative Dehydratase Enzyme Involved in *Mycobacterium* Tuberculosis Fatty Acid Synthase II. *Microbiology*, 153(10), 3314-3322.
- Bubalo, M. C., Ćurko, N., Tomašević, M., Ganić, K. K., & Redovniković, I. R. (2016). Green Extraction of Grape Skin Phenolics by Using Deep Eutectic Solvents. *Food chemistry*, 200, 159-166.
- Bunalema, L., Obakiro, S., Tabuti, J. R., & Waako, P. (2014). Knowledge on Plants Used Traditionally in the Treatment of Tuberculosis in Uganda. *Journal of Ethnopharmacology*, 151(2), 999-1004.
- Caldwell, C. G., Franzblau, S. G., Suarez, E., & Timmermann, B. N. (2000). Oleanane Triterpenes from *Junellia Tridens*. *Journal of natural products*, 63(12), 1611-1614.
- Cambau, E., & Drancourt, M. (2014). Steps Towards the Discovery of *Mycobacterium* Tuberculosis by Robert Koch, 1882. *Clinical Microbiology and Infection*, 20(3), 196-201.
- Cantrell, C. L., Rajab, M. S., Franzblau, S. G., Fronczek, F. R., & Fischer, N. H. (1999). Antimycobacterial Ergosterol-5, 8-Endoperoxide from *Ajuga Remota*. *Planta Medica*, 65(08), 732-734.

- Cao, H., Song, S., Zhang, H., Zhang, Y., Qu, R., Yang, B., Jing, Y., Hu, T., Yan, F., & Wang, B. (2013). Chemopreventive Effects of Berberine on Intestinal Tumor Development in Apc Min/+ Mice. *BMC gastroenterology*, 13(1), 1-9.
- Carrara, V., Garcia, V., Faiões, V., Cunha-Júnior, E., Torres-Santos, E., & Cortez, D. (2017). Supercritical Fluid Extraction of Pyrrolidine Alkaloid from Leaves of Piper Amalago L. *Evidence-Based Complementary and Alternative Medicine*, 2017.
- Casey, M., Adams, J., & Sibbritt, D. (2007). An Examination of the Prescription and Dispensing of Medicines by Western Herbal Therapists: A National Survey in Australia. *Complementary therapies in medicine*, 15(1), 13-20.
- Catalão, M., & Pimentel, M. (2018). Mycobacteriophage Lysis Enzymes: Targeting the Mycobacterial Cell Envelope. *Viruses*, 10(8), 428.
- Cernuschi, T., Malvolti, S., Nickels, E., & Friede, M. (2018). Bacillus Calmette-Guérin (Bcg) Vaccine: A Global Assessment of Demand and Supply Balance. *Vaccine*, 36(4), 498-506.
- Chadha, V. (1997). Global Trends of Tuberculosis—an Epidemiological Review. *NTI Bulletin*, 33(1), 11-18.
- Chakraborty, P., & Kumar, A. (2019). The Extracellular Matrix of Mycobacterial Biofilms: Could We Shorten the Treatment of Mycobacterial Infections? *Microbial Cell*, 6(2), 105-122.
- Chan, S. M., Khoo, K. S., & Sit, N. W. (2015). Interactions between Plant Extracts and Cell Viability Indicators During Cytotoxicity Testing: Implications for Ethnopharmacological Studies. *Tropical Journal of Pharmaceutical Research*, 14(11), 1991-1998.
- Chang, C.-H., Chiang, M.-L., & Chou, C.-C. (2009). The Effect of Temperature and Length of Heat Shock Treatment on the Thermal Tolerance and Cell Leakage of Cronobacter Sakazakii Bcrc 13988. *International journal of food microbiology*, 134(3), 184-189.
- Chaudhary, N., Mehra, V., Mago, P., & Khatri, M. (2017). Antimycobacterial Potential of Indian Spices: A Review. *International Journal of Pharmacognosy and Phytochemical Research*, 9(8), 1127-1134.
- Chen, C.-Y., Nace, G. W., & Irwin, P. L. (2003). A 6× 6 Drop Plate Method for Simultaneous Colony Counting and Mpn Enumeration of Campylobacter

- Jejuni, Listeria Monocytogenes, and Escherichia Coli. *Journal of microbiological methods*, 55(2), 475-479.
- Chen, L. W., Cheng, M. J., Peng, C. F., & Chen, I. S. (2010). Secondary Metabolites and Antimycobacterial Activities from the Roots of Ficus Nervosa. *Chemistry & Biodiversity*, 7(7), 1814-1821.
- Chen, W., Shu, W., Wang, M., Hou, Y., Xia, Y., Xu, W., Bai, L., Nie, S., Cheng, S., & Xu, Y. (2013). Pulmonary Tuberculosis Incidence and Risk Factors in Rural Areas of China: A Cohort Study. *PLoS One*, 8(3), e58171.
- Chiaradia, L., Lefebvre, C., Parra, J., Marcoux, J., Burlet-Schiltz, O., Etienne, G., Tropis, M., & Daffé, M. (2017). Dissecting the Mycobacterial Cell Envelope and Defining the Composition of the Native Mycomembrane. *Scientific reports*, 7(1), 12807.
- Chinsembu, K. C. (2016). Tuberculosis and Nature's Pharmacy of Putative Anti-Tuberculosis Agents. *Acta Tropica*, 153, 46-56.
- Choudhary, K., Singh, M., & Pillai, U. (2008). Ethnobotanical Survey of Rajasthan-an Update. *American-Eurasian Journal of Botany*, 1(2), 38-45.
- Chraibi, M., Farah, A., Elamin, O., Iraqui, H. M., & Fikri-Benbrahim, K. (2020). Characterization, Antioxidant, Antimycobacterial, Antimicrobial Effects of Moroccan Rosemary Essential Oil, and Its Synergistic Antimicrobial Potential with Carvacrol. *Journal of Advanced Pharmaceutical Technology & Research*, 11(1), 25.
- Chung, K.-T., Wong, T. Y., Wei, C.-I., Huang, Y.-W., & Lin, Y. (1998). Tannins and Human Health: A Review. *Critical reviews in food science and nutrition*, 38(6), 421-464.
- Cole, S., Brosch, R., Parkhill, J., Garnier, T., Churcher, C., Harris, D., Gordon, S., Eiglmeier, K., Gas, S., & Barry, C. r. (1998). Deciphering the Biology of Mycobacterium Tuberculosis from the Complete Genome Sequence. *Nature*, 396(6707), 190-190.
- Compean, K., & Ynalvez, R. (2014). Antimicrobial Activity of Plant Secondary Metabolites: A Review. *Res J Med Plant*, 8(5), 204-213.
- Copp, B. R. (2003). Antimycobacterial Natural Products. *Natural product reports*, 20(6), 535-557.
- Coronado-Aceves, E. W., Gigliarelli, G., Garibay-Escobar, A., Zepeda, R. E. R., Curini, M., Cervantes, J. L., Espitia-Pinzón, C. I. I., Superchi, S., Vergura, S.,

- & Marcotullio, M. C. (2017). New Isoflavonoids from the Extract of Rhynchosia Precatoria (Humb. & Bonpl. Ex Willd.) DC. And Their Antimycobacterial Activity. *Journal of Ethnopharmacology*, 206, 92-100.
- Cox, S. D., Mann, C. M., Markham, J. L., Gustafson, J. E., Warmington, J. R., & Wyllie, S. G. (2001). Determining the Antimicrobial Actions of Tea Tree Oil. *Molecules*, 6(2), 87-91.
- Crutchfield, J. W., Butera, S. T., & Folks, T. M. (1996). Inhibition of Hiv Activation in Latently Infected Cells by Flavonoid Compounds. *AIDS research and human retroviruses*, 12(1), 39-46.
- Cseri, L., & Szekely, G. (2019). Towards Cleaner Polarclean: Efficient Synthesis and Extended Applications of the Polar Aprotic Solvent Methyl 5-(Dimethylamino)-2-Methyl-5-Oxopentanoate. *Green Chemistry*, 21(15), 4178-4188.
- Cui, Q., Liu, J.-Z., Wang, L.-T., Kang, Y.-F., Meng, Y., Jiao, J., & Fu, Y.-J. (2018). Sustainable Deep Eutectic Solvents Preparation and Their Efficiency in Extraction and Enrichment of Main Bioactive Flavonoids from Sea Buckthorn Leaves. *Journal of cleaner production*, 184, 826-835.
- D'Agostino, C., Harris, R. C., Abbott, A. P., Gladden, L. F., & Mantle, M. D. (2011). Molecular Motion and Ion Diffusion in Choline Chloride Based Deep Eutectic Solvents Studied by <sup>1</sup>H Pulsed Field Gradient Nmr Spectroscopy. *Physical Chemistry Chemical Physics*, 13(48), 21383-21391.
- da Silva Bomfim, N., Kohiyama, C. Y., Nakasugi, L. P., Nerilo, S. B., Mossini, S. A. G., Romoli, J. C. Z., Graton Mikcha, J. M., Abreu Filho, B. A. d., & Machinski Jr, M. (2020). Antifungal and Antiaflatoxigenic Activity of Rosemary Essential Oil (*Rosmarinus Officinalis* L.) against *Aspergillus Flavus*. *Food Additives & Contaminants: Part A*, 37(1), 153-161.
- Dai et al. (2013). Natural Deep Eutectic Solvents as a New Extraction Media for Phenolic Metabolites in *Carthamus Tinctorius* L. *Analytical chemistry*, 85(13), 6272-6278.
- Dai, Y., van Spronsen, J., Witkamp, G.-J., Verpoorte, R., & Choi, Y. H. (2013). Natural Deep Eutectic Solvents as New Potential Media for Green Technology. *Analytica chimica acta*, 766, 61-68.

- Dai, Y., Witkamp, G.-J., Verpoorte, R., & Choi, Y. H. (2013). Natural Deep Eutectic Solvents as a New Extraction Media for Phenolic Metabolites in *Carthamus Tinctorius L.* *Analytical chemistry*, 85(13), 6272-6278.
- Daniel, T. M. (2006). The History of Tuberculosis. *Respiratory medicine*, 100(11), 1862-1870.
- Dehyab, A. S., Bakar, M. F. A., AlOmar, M. K., & Sabran, S. F. (2020). A Review of Medicinal Plant of Middle East and North Africa (Mena) Region as Source in Tuberculosis Drug Discovery. *Saudi journal of biological sciences*, 27(9), 2457.
- Delogu, G., Sali, M., & Fadda, G. (2013). The Biology of *Mycobacterium Tuberculosis* Infection. *Mediterranean journal of hematology and infectious diseases*, 5(1).
- Desalu, O. O., Adeoti, A. O., Fadeyi, A., Salami, A. K., Fawibe, A. E., & Oyedepo, O. O. (2013). Awareness of the Warning Signs, Risk Factors, and Treatment for Tuberculosis among Urban Nigerians. *Tuberculosis research and treatment*, 2013.
- Dheyab, A. S., Abu Bakar, M. F., AlOmar, M., Sabran, S. F., Muhamad Hanafi, A. F., & Mohamad, A. (2021). Deep Eutectic Solvents (Dess) as Green Extraction Media of Beneficial Bioactive Phytochemicals. *Separations*, 8(10), 176.
- Dianita, R., & Jantan, I. (2017). Ethnomedicinal Uses, Phytochemistry and Pharmacological Aspects of the Genus *Premna*: A Review. *Pharmaceutical biology*, 55(1), 1715-1739.
- Dias, D. A., Urban, S., & Roessner, U. (2012). A Historical Overview of Natural Products in Drug Discovery. *Metabolites*, 2(2), 303-336.
- Diloksumpun, S., Jeenho, P., Namkhot, S., Saleepochn, T., & Luangkamin, S. (2021). Potent Antioxidant Activities of Half-Sib Families of *Eucalyptus Camaldulensis* Dehnh. Leaf Essential Oils Planted in Thailand and Their Antioxidative Components.
- Dodor, E. A. (2009). *An Exploration of the Causes, Manifestations and Consequences of Tuberculosis Stigma in an Urban District in Ghana*. University of Nottingham,
- Doughari, J. H. (2012). *Phytochemicals: Extraction Methods, Basic Structures and Mode of Action as Potential Chemotherapeutic Agents*: INTECH Open Access Publisher Rijeka, Croatia.

- Duan, L., Dou, L.-L., Guo, L., Li, P., & Liu, E.-H. (2016). Comprehensive Evaluation of Deep Eutectic Solvents in Extraction of Bioactive Natural Products. *ACS Sustainable Chemistry & Engineering*, 4(4), 2405-2411.
- Durand, G. A., Raoult, D., & Dubourg, G. (2018). Antibiotic Discovery: History, Methods and Perspectives. *International journal of antimicrobial agents*.
- Dzoyem, J. P., Aro, A. O., McGaw, L. J., & Eloff, J. N. (2016). Antimycobacterial Activity against Different Pathogens and Selectivity Index of Fourteen Medicinal Plants Used in Southern Africa to Treat Tuberculosis and Respiratory Ailments. *South African Journal of Botany*, 102, 70-74.
- Egelund, E. F., Dupree, L., Huesgen, E., & Peloquin, C. A. (2017). The Pharmacological Challenges of Treating Tuberculosis and Hiv Coinfections. *Expert review of clinical pharmacology*, 10(2), 213-223.
- Elmaidomy, A. H., Alhadrami, H. A., Amin, E., Aly, H. F., Othman, A. M., Rateb, M. E., Hetta, M. H., Abdelmohsen, U. R., & M Hassan, H. (2020). Anti-Inflammatory and Antioxidant Activities of Terpene-and Polyphenol-Rich *Premna Odorata* Leaves on Alcohol-Inflamed Female Wistar Albino Rat Liver. *Molecules*, 25(14), 3116.
- Elmaidomy, A. H., Hassan, H. M., Amin, E., Mohamed, W., & Hetta, M. H. (2017a). *Premna Odorata* Volatile Oil as a New Mycobacterium Tuberculosis Growth Inhibitor for the Control of Tuberculosis Disease. *European Journal of Medicinal Plants*, 1-11.
- Elmaidomy, A. H., Hassan, H. M., Amin, E., Mohamed, W., & Hetta, M. H. (2017b). *Premna Odorata* Volatile Oil as a New Mycobacterium Tuberculosis Growth Inhibitor for the Control of Tuberculosis Disease. *Eur. J. Med. Plants*, 21, 1-11.
- Elmaidomy, A. H., Mohammed, R., M Hassan, H., I Owis, A., E Rateb, M., A Khanfar, M., Krischke, M., J Mueller, M., & Ramadan Abdelmohsen, U. (2019). Metabolomic Profiling and Cytotoxic Tetrahydrofurofuran Lignans Investigations from *Premna Odorata* Blanco. *Metabolites*, 9(10), 223.
- Elmaidomy, A. H., Mohammed, R., Owis, A. I., Hetta, M. H., AboulMagd, A. M., Siddique, A. B., Abdelmohsen, U. R., Rateb, M. E., El Sayed, K. A., & Hassan, H. M. (2020). Triple-Negative Breast Cancer Suppressive Activities, Antioxidants and Pharmacophore Model of New Acylated Rhamnopyranoses from *Premna Odorata*. *RSC Advances*, 10(18), 10584-10598.

- Eloff, J. (1998). Which Extractant Should Be Used for the Screening and Isolation of Antimicrobial Components from Plants? *Journal of Ethnopharmacology*, 60(1), 1-8.
- Espino, M., Solari, M., de los Ángeles Fernández, M., Boiteux, J., Gómez, M. R., & Silva, M. F. (2019). Nades-Mediated Folk Plant Extracts as Novel Antifungal Agents against *Candida Albicans*. *Journal of pharmaceutical and biomedical analysis*, 167, 15-20.
- Fabry, W., Okemo, P. O., & Ansorg, R. (1998). Antibacterial Activity of East African Medicinal Plants. *Journal of Ethnopharmacology*, 60(1), 79-84.
- Fachriyah, E., Wibawa, P., & Awaliyah, A. (2020). *Antibacterial Activity of Basil Oil (Ocimum Basilicum L) and Basil Oil Nanoemulsion*. Paper presented at the Journal of Physics: Conference Series.
- Fallis, A. (2013). Chromatographic Techniques. *Journal of Chemical Information and Modeling*, 53, 1689–1699.
- Famewo, E. B., Clarke, A. M., & Afolayan, A. J. (2017). Ethno-Medicinal Documentation of Polyherbal Medicines Used for the Treatment of Tuberculosis in Amathole District Municipality of the Eastern Cape Province, South Africa. *Pharmaceutical biology*, 55(1), 696-700.
- Favela-Hernández, J., García, A., Garza-González, E., Rivas-Galindo, V., & Camacho-Corona, M. d. R. (2012). Antibacterial and Antimycobacterial Lignans and Flavonoids from *Larrea Tridentata*. *Phytotherapy Research*, 26(12), 1957-1960.
- Ferreira, I. C., Martins, N., & Barros, L. (2017). Phenolic Compounds and Its Bioavailability: In Vitro Bioactive Compounds or Health Promoters? In *Advances in Food and Nutrition Research* (Vol. 82, pp. 1-44): Elsevier.
- Fontana, R., Macchi, G., Caproni, A., Sicurella, M., Buratto, M., Salvatori, F., Pappadà, M., Manfredini, S., Baldisserotto, A., & Marconi, P. (2022). Control of *Erwinia Amylovora* Growth by *Moringa Oleifera* Leaf Extracts: In Vitro and in Planta Effects. *Plants*, 11(7), 957.
- Foongladda, S., Roengsanthia, D., Arjrattanakool, W., Chuchottaworn, C., Chaiprasert, A., & Franzblau, S. (2002). Rapid and Simple Mtt Method for Rifampicin and Isoniazid Susceptibility Testing of *Mycobacterium Tuberculosis*. *The International Journal of Tuberculosis and Lung Disease*, 6(12), 1118-1122.

- Forrellad, M. A., Klepp, L. I., Gioffré, A., Sabio y Garcia, J., Morbidoni, H. R., Santangelo, M. D. L. P., Cataldi, A. A., & Bigi, F. (2013). Virulence Factors of the Mycobacterium Tuberculosis Complex. *Virulence*, 4(1), 3-66.
- Fu, Y., Zu, Y., Chen, L., Shi, X., Wang, Z., Sun, S., & Efferth, T. (2007). Antimicrobial Activity of Clove and Rosemary Essential Oils Alone and in Combination. *Phytotherapy Research*, 21(10), 989-994.
- Fyhrquist, P., Laakso, I., Marco, S. G., Julkunen-Tiitto, R., & Hiltunen, R. (2014). Antimycobacterial Activity of Ellagitannin and Ellagic Acid Derivate Rich Crude Extracts and Fractions of Five Selected Species of Terminalia Used for Treatment of Infectious Diseases in African Traditional Medicine. *South African Journal of Botany*, 90, 1-16.
- Gandji, K., Chadare, F., Idohou, R., Salako, V., Assogbadjo, A., & Kakaï, R. G. (2018). Status and Utilisation of Moringa Oleifera Lam: A Review. *African Crop Science Journal*, 26(1), 137-156.
- Garzoli, S., Laghezza Masci, V., Franceschi, S., Tiezzi, A., Giacomello, P., & Ovidi, E. (2021). Headspace/Gc–Ms Analysis and Investigation of Antibacterial, Antioxidant and Cytotoxic Activity of Essential Oils and Hydrolates from Rosmarinus Officinalis L. And Lavandula Angustifolia Miller. *Foods*, 10(8), 1768.
- Gatfield, J., & Pieters, J. (2000). Essential Role for Cholesterol in Entry of Mycobacteria into Macrophages. *Science*, 288(5471), 1647-1651.
- Gautam, R., Saklani, A., & Jachak, S. M. (2007). Indian Medicinal Plants as a Source of Antimycobacterial Agents. *Journal of Ethnopharmacology*, 110(2), 200-234.
- Gemechu, A., Giday, M., Worku, A., & Ameni, G. (2013). In Vitro Anti-Mycobacterial Activity of Selected Medicinal Plants against Mycobacterium Tuberculosis and Mycobacterium Bovis Strains. *BMC complementary and alternative medicine*, 13(1), 291.
- González-Minero, F. J., Bravo-Díaz, L., & Ayala-Gómez, A. (2020). Rosmarinus Officinalis L.(Rosemary): An Ancient Plant with Uses in Personal Healthcare and Cosmetics. *Cosmetics*, 7(4), 77.
- Gorke, J., Srienc, F., & Kazlauskas, R. (2010). Toward Advanced Ionic Liquids. Polar, Enzyme-Friendly Solvents for Biocatalysis. *Biotechnology and Bioprocess Engineering*, 15(1), 40-53.

- Goroh, M. M. D., Rajahram, G. S., Avoi, R., Den Boogaard, V., Christel, H., William, T., Ralph, A. P., & Lowbridge, C. (2020). Epidemiology of Tuberculosis in Sabah, Malaysia, 2012–2018. *Infectious diseases of poverty*, 9(1), 1-11.
- Green, E., Samie, A., Obi, C. L., Bessong, P. O., & Ndip, R. N. (2010). Inhibitory Properties of Selected South African Medicinal Plants against Mycobacterium Tuberculosis. *Journal of Ethnopharmacology*, 130(1), 151-157.
- Groenewald, W., Parra Cruz, R., Jaeger, C., & Croft, A. (2018). Revealing Solvent-Dependent Folding Behavior of Mycolic Acids from Mycobacterium Tuberculosis by Advanced Simulation Analysis.
- Grzegorzewicz, A. E., De Sousa-D'Auria, C., McNeil, M. R., Huc-Claustre, E., Jones, V., Petit, C., kumar Angala, S., Zemanová, J., Wang, Q., & Belardinelli, J. M. (2016). Assembling of the Mycobacterium Tuberculosis Cell Wall Core. *Journal of Biological Chemistry*, 291(36), 18867-18879.
- Gunaherath, G. K. B., & Gunatilaka, A. L. (2006). Plant Steroids: Occurrence, Biological Significance, and Their Analysis. *Encyclopedia of Analytical Chemistry: Applications, Theory and Instrumentation*, 1-31.
- Gunjan, M., Karna, L., Dayalan, K., & Sasigaran, P. (2012). A Review and Search of Phytomedicine Used by Traditional People of Malaysia (Ipoh, Perak). *International Journal of Phytotherapy Research*, 2(3), 26-41.
- Gupta, A., Naraniwal, M., & Kothari, V. (2012). Modern Extraction Methods for Preparation of Bioactive Plant Extracts. *International journal of applied and natural sciences*, 1(1), 8-26.
- Gupta, R., Thakur, B., Singh, P., Singh, H., Sharma, V., Katoch, V., & Chauhan, S. (2010). Anti-Tuberculosis Activity of Selected Medicinal Plants against Multi-Drug Resistant Mycobacterium Tuberculosis Isolates. *Indian Journal of Medical Research*, 131(6), 809.
- Gupta, R. S. (2019). Commentary: Genome-Based Taxonomic Classification of the Phylum Actinobacteria. *Frontiers in Microbiology*, 10, 206.
- Gürgan, M., & Adiloglu, S. (2021). Increasing Concentrations of Iron Fertilizer Affect Antibacterial Activity of Basil (*Ocimum Basilicum L.*). *Industrial Crops and Products*, 170, 113768.
- Gurnani, N., Mehta, D., Gupta, M., & Mehta, B. (2014). Natural Products: Source of Potential Drugs. *Afr J Basic Appl Sci*, 6(6), 171-186.
- H Herzog, B. (1998). History of Tuberculosis. *Respiration*, 65(1), 5-15.

- Hajmohammadi, A., Pirouzifard, M., Shahedi, M., & Alizadeh, M. (2016). Enrichment of a Fruit-Based Beverage in Dietary Fiber Using Basil Seed: Effect of Carboxymethyl Cellulose and Gum Tragacanth on Stability. *LWT*, 74, 84-91.
- Hamed, Y. S., Abdin, M., Rayan, A. M., Akhtar, H. M. S., & Zeng, X. (2021). Synergistic Inhibition of Isolated Flavonoids from *Moringa Oleifera* Leaf on A-Glucosidase Activity. *LWT*, 141, 111081.
- Han, X., Shen, T., & Lou, H. (2007). Dietary Polyphenols and Their Biological Significance. *International journal of molecular sciences*, 8(9), 950-988.
- Handa, S. (2008). An Overview of Extraction Techniques for Medicinal and Aromatic Plants. *Extraction technologies for medicinal and aromatic plants*, 1, 21-40.
- Hao, C., Chen, L., Dong, H., Xing, W., Xue, F., & Cheng, Y. (2020). Extraction of Flavonoids from *Scutellariae Radix* Using Ultrasound-Assisted Deep Eutectic Solvents and Evaluation of Their Anti-Inflammatory Activities. *ACS omega*, 5(36), 23140-23147.
- Harland, C. W., Rabuka, D., Bertozzi, C. R., & Parthasarathy, R. (2008). The *Mycobacterium Tuberculosis* Virulence Factor Trehalose Dimycolate Imparts Desiccation Resistance to Model Mycobacterial Membranes. *Biophysical journal*, 94(12), 4718-4724.
- Hayyan, A., Hashim, M. A., Hayyan, M., Mjalli, F. S., & AlNashef, I. M. (2014). A New Processing Route for Cleaner Production of Biodiesel Fuel Using a Choline Chloride Based Deep Eutectic Solvent. *Journal of cleaner production*, 65, 246-251.
- Hayyan, M., Hashim, M. A., Hayyan, A., Al-Saadi, M. A., AlNashef, I. M., Mirghani, M. E., & Saheed, O. K. (2013). Are Deep Eutectic Solvents Benign or Toxic? *Chemosphere*, 90(7), 2193-2195.
- He, X. (2000). On-Line Identification of Phytochemical Constituents in Botanical Extracts by Combined High-Performance Liquid Chromatographic – Diode Array Detection – Mass Spectrometric Techniques. *Journal of Chromatography A*, 880, 203–232.
- Hedhammar, M., Karlstrom, A., & Hober, S. (2006). Chromatographic Methods for Protein Purification. *Stockholm: Royal Institute of Technology*, 1-31.
- Heinrich, M. (2013). Ethnopharmacology and Drug Discovery. *Comprehensive Natural Products II: Chemistry and Biology, Development & Modification of Bioactivity*, 3, 351-381.

- Heinrich, M., Kufer, J., Leonti, M., & Pardo-de-Santayana, M. (2006). Ethnobotany and Ethnopharmacology—Interdisciplinary Links with the Historical Sciences. *Journal of Ethnopharmacology*, 107(2), 157-160.
- Hershkovitz, I., Donoghue, H. D., Minnikin, D. E., Besra, G. S., Lee, O. Y., Gernaey, A. M., Galili, E., Eshed, V., Greenblatt, C. L., & Lemma, E. (2008). Detection and Molecular Characterization of 9000-Year-Old Mycobacterium Tuberculosis from a Neolithic Settlement in the Eastern Mediterranean. *PLoS One*, 3(10), e3426.
- Hertz, C. J., Kiertscher, S. M., Godowski, P. J., Bouis, D. A., Norgard, M. V., & Roth, M. D. (2015). Microbial Lipopeptides Stimulate Dendritic Cell Maturation Via Toll-Like Receptor 2. *The Journal of Immunology*, 166, pp. 2444–2450.
- Hollman, P. C. H., & Arts, I. C. W. (2000). Flavonols, Flavones and Flavanols—Nature, Occurrence and Dietary Burden. *Journal of the Science of Food and Agriculture*, 80(7), 1081-1093.
- Hsieh, Y.-H., Li, Y., Pan, Z., Chen, Z., Lu, J., Yuan, J., Zhu, Z., & Zhang, J. (2020). Ultrasonication-Assisted Synthesis of Alcohol-Based Deep Eutectic Solvents for Extraction of Active Compounds from Ginger. *Ultrasonics sonochemistry*, 63, 104915.
- Hudzicki, J. (2009). Kirby-Bauer Disk Diffusion Susceptibility Test Protocol.
- Ibekwe, N. N., Nvau, J. B., Oladosu, P. O., Usman, A. M., Ibrahim, K., Boshoff, H. I., Dowd, C. S., Orisadipe, A. T., Aiyelaagbe, O., & Adesomoju, A. A. (2014). Some Nigerian Anti-Tuberculosis Ethnomedicines: A Preliminary Efficacy Assessment. *Journal of Ethnopharmacology*, 155(1), 524-532.
- Igbokwe, C., Lawal, T., & Adeniyi, B. (2014). In Vitro Anti-Mycobacteria Sensitivity and Kill-Kinetics of Allium Ascalonicum L.(Whole Plant) on Nontuberculous Mycobacteria Species. *African Journal of Biomedical Research*, 17(2), 93-99.
- Irchhaiya, R., Kumar, A., Yadav, A., Gupta, N., Kumar, S., Gupta, N., Kumar, S., Yadav, V., Prakash, A., & Gurjar, H. (2015). Metabolites in Plants and Its Classification. *World journal of pharmacy and pharmaceutical sciences*, 4(1), 287-305.
- Islam, M. M., Hameed, H. A., Mugweru, J., Chhotaray, C., Wang, C., Tan, Y., Liu, J., Li, X., Tan, S., & Ojima, I. (2017). Drug Resistance Mechanisms and Novel Drug Targets for Tuberculosis Therapy. *Journal of Genetics and Genomics*, 44(1), 21-37.

- Jackson, M. (2014). The Mycobacterial Cell Envelope—Lipids. *Cold Spring Harbor perspectives in medicine*, a021105.
- Jahanian, H., Kahkeshani, N., Sanei-Dehkordi, A., Isman, M. B., Saeedi, M., & Khanavi, M. (2022). Rosmarinus Officinalis as a Natural Insecticide: A Review. *International Journal of Pest Management*, 1-46.
- Jalil, A., Azri, M., Shuid, A. N., & Muhammad, N. (2012). Role of Medicinal Plants and Natural Products on Osteoporotic Fracture Healing. *Evidence-Based Complementary and Alternative Medicine*, 2012.
- Jamal, J. A. (2006). Malay Traditional Medicine. *Tech Monitor (Special Feature: traditional Medicine: S & T Advancement)*, 37-49.
- Jamal, P., Karim, I. A., Abdullah, E., Raus, R. A., & Hashim, Y. Z. (2011). Phytochemical Screening for Antibacterial Activity of Potential Malaysian Medicinal Plants. *African Journal of Biotechnology*, 10(81), 18795-18799.
- Jeong, K. M., Han, S. Y., Kim, E. M., Jin, Y., & Lee, J. (2018). Deep Eutectic Solvent-Based Valorization of Spent Coffee Grounds. *Food chemistry*, 255, 357-364.
- Jeong, K. M., Lee, M. S., Nam, M. W., Zhao, J., Jin, Y., Lee, D.-K., Kwon, S. W., Jeong, J. H., & Lee, J. (2015). Tailoring and Recycling of Deep Eutectic Solvents as Sustainable and Efficient Extraction Media. *Journal of Chromatography A*, 1424, 10-17.
- Jetan, C., Jamaiah, I., Rohela, M., & Nissapatorn, V. (2010). Tuberculosis: An Eight Year (2000–2007) Retrospective Study at the University of Malaya Medical Centre (Ummc), Kuala Lumpur, Malaysia. *Southeast Asian J Trop Med Public Health*, 41(2), 378-385.
- Jiménez-Arellanes, M. A., Gutiérrez-Rebolledo, G., Rojas-Tomé, S., & Meckes-Fischer, M. (2014). Medicinal Plants, an Important Reserve of Antimycobacterial and Antitubercular Drugs: An Update. *Journal of Infectious Diseases and Therapy*.
- Jonsson, P., Johansson, A. I., Gullberg, J., Trygg, J., Grung, B., Marklund, S., Sjöström, M., Antti, H., & Moritz, T. (2005). High-Throughput Data Analysis for Detecting and Identifying Differences between Samples in Gc/Ms-Based Metabolomic Analyses. *Analytical chemistry*, 77(17), 5635-5642.
- Jordao, L., & Vieira, O. V. (2011). Tuberculosis: New Aspects of an Old Disease. *International journal of cell biology*, 2011.

- Kabongo-Kayoka, P. N., Eloff, J. N., Obi, C. L., & McGaw, L. J. (2016). Antimycobacterial Activity and Low Cytotoxicity of Leaf Extracts of Some African Anacardiaceae Tree Species. *Phytotherapy Research*, 30(12), 2001-2011.
- Kahaliw, W., Aseffa, A., Abebe, M., Teferi, M., & Engidawork, E. (2017). Evaluation of the Antimycobacterial Activity of Crude Extracts and Solvent Fractions of Selected Ethiopian Medicinal Plants. *BMC complementary and alternative medicine*, 17(1), 1-9.
- Kamran, M., Khan, N., Jabeen, H., Iqbal, S., & Siddiqi, A. M. (2022). Comparison of Antioxidant Effect of Crocus Sativus and Vitamin E on Amakinin Induced Nephrotoxicity in Albino Rats. *ANNALS OF ABBASI SHAHEED HOSPITAL AND KARACHI MEDICAL & DENTAL COLLEGE*, 27(1), 31-39.
- Kandasamy, S., Govindarajalou, R., Chakkalakkooombil, S. V., & Penumadu, P. (2018). Isolated Hepatobiliary Tuberculosis: A Diagnostic Challenge. *BMJ case reports*, 2018.
- Kanokmedhakul, S., Kanokmedhakul, K., & Lekphrom, R. (2007). Bioactive Constituents of the Roots of Polyalthia Cerasoides. *Journal of natural products*, 70(9), 1536-1538.
- Kashyap, P., Kumar, S., Riar, C. S., Jindal, N., Baniwal, P., Guiné, R. P., Correia, P. M., Mehra, R., & Kumar, H. (2022). Recent Advances in Drumstick (*Moringa Oleifera*) Leaves Bioactive Compounds: Composition, Health Benefits, Bioaccessibility, and Dietary Applications. *Antioxidants*, 11(2), 402.
- Kaur, N., Arora, D. S., Kalia, N., & Kaur, M. (2020). Antibiofilm, Antiproliferative, Antioxidant and Antimutagenic Activities of an Endophytic Fungus *Aspergillus Fumigatus* from *Moringa Oleifera*. *Molecular biology reports*, 47(4), 2901-2911.
- Kaur, R., & Kaur, H. (2015). Antitubercular Activity and Phytochemical Screening of Selected Medicinal Plants. *Orient J Chem*, 31(1), 597-600.
- Kaur, S., Samota, M. K., Choudhary, M., Choudhary, M., Pandey, A. K., Sharma, A., & Thakur, J. (2022). How Do Plants Defend Themselves against Pathogens-Biochemical Mechanisms and Genetic Interventions. *Physiology and Molecular Biology of Plants*, 1-20.
- Keshavjee, S., & Farmer, P. E. (2012). Tuberculosis, Drug Resistance, and the History of Modern Medicine. *New England Journal of Medicine*, 367(10), 931-936.

- Kessel, M., Rana, B., & Boehme, C. (2019). How to Fix Tuberculosis R&D—the Community Speaks. *Nature Biotechnology*, 37(4), 350-351.
- Khadka, P., Dummer, J., Hill, P. C., & Das, S. C. (2018). Considerations in Preparing for Clinical Studies of Inhaled Rifampicin to Enhance Tuberculosis Treatment. *International journal of pharmaceutics*, 548(1), 244-254.
- Khan, J., Tousif, M. I., Saleem, M., Nazir, M., Touseef, S., Saleem, K., Asim, S., Khan, A., Asghar, M. A., & Zengin, G. (2021). Insight into the Phytochemical Composition, Biological Activities and Docking Studies of Moringa Oleifera Lam. To Authenticate Its Use in Biopharmaceutical Industries. *Industrial Crops and Products*, 172, 114042.
- Khan, N., & Das, A. (2022). Time for Isoniazid Pharmacogenomic-Guided Therapy of Tuberculosis Based on Nat2 Acetylation Profiles in India. *European Journal of Drug Metabolism and Pharmacokinetics*, 1-5.
- Khara, J. S., Wang, Y., Ke, X.-Y., Liu, S., Newton, S. M., Langford, P. R., Yang, Y. Y., & Ee, P. L. R. (2014). Anti-Mycobacterial Activities of Synthetic Cationic A-Helical Peptides and Their Synergism with Rifampicin. *Biomaterials*, 35(6), 2032-2038.
- Kidd, T., Mitchell, S., Dehays, J., & Wibberley, E. (2022). Fluoroquinolones: With Great Power Comes Great Risk. *Nursing2022*, 52(1), 24-27.
- Kim, S., Lee, H., Lee, S., Yoon, Y., & Choi, K.-H. (2015). Antimicrobial Action of Oleanolic Acid on Listeria Monocytogenes, Enterococcus Faecium, and Enterococcus Faecalis. *PLoS One*, 10(3), e0118800.
- Kishore, N., Mishra, B. B., Tripathi, V., & Tiwari, V. K. (2009). Alkaloids as Potential Anti-Tubercular Agents. *Fitoterapia*, 80(3), 149-163.
- Kontogianni, V. G., Tomic, G., Nikolic, I., Nerantzaki, A. A., Sayyad, N., Stosic-Grujicic, S., Stojanovic, I., Gerohanassis, I. P., & Tzakos, A. G. (2013). Phytochemical Profile of Rosmarinus Officinalis and Salvia Officinalis Extracts and Correlation to Their Antioxidant and Anti-Proliferative Activity. *Food chemistry*, 136(1), 120-129.
- Korfmacher, W. A. (2005). Foundation Review: Principles and Applications of LC-MS in New Drug Discovery. *Drug discovery today*, 10(20), 1357-1367.
- Koul, A., Arnoult, E., Lounis, N., Guillemont, J., & Andries, K. (2011). The Challenge of New Drug Discovery for Tuberculosis. *Nature*, 469(7331), 483-490.

- Krieger, M. S., & Hites, R. A. (1992). Diffusion Denuder for the Collection of Semivolatile Organic Compounds. *Environmental science & technology*, 26(8), 1551-1555.
- Kumar, N., Banik, A., & Sharma, P. (2010). Use of Secondary Metabolite in Tuberculosis: A Review. *Der Pharma Chemica*, 2(6), 311-319.
- Kuula, L. S., Backman, J. T., & Blom, M. L. (2022). Healthcare Costs and Mortality Associated with Serious Fluoroquinolone-Related Adverse Reactions. *Pharmacology Research & Perspectives*, 10(2), e00931.
- Lapierre, D. (2011). *Mycobacterium Tuberculosis Cases*.
- Laurenzi, M., Ginsberg, A., & Spigelman, M. (2007). Challenges Associated with Current and Future Tb Treatment. *Infectious Disorders-Drug Targets (Formerly Current Drug Targets-Infectious Disorders)*, 7(2), 105-119.
- Lawal, T. O., Adeniyi, B. A., Adegoke, A. O., Franzblau, S. G., & Mahady, G. B. (2012). In Vitro Susceptibility of Mycobacterium Tuberculosis to Extracts of Eucalyptus Camaldulensis and Eucalyptus Torelliana and Isolated Compounds. *Pharmaceutical biology*, 50(1), 92-98.
- Leitão, F., Leitão, S. G., de Almeida, M. Z., Cantos, J., Coelho, T., & da Silva, P. E. A. (2013). Medicinal Plants from Open-Air Markets in the State of Rio De Janeiro, Brazil as a Potential Source of New Antimycobacterial Agents. *Journal of Ethnopharmacology*, 149(2), 513-521.
- Li, L., Liu, J.-Z., Luo, M., Wang, W., Huang, Y.-Y., Efferth, T., Wang, H.-M., & Fu, Y.-J. (2016). Efficient Extraction and Preparative Separation of Four Main Isoflavonoids from Dalbergia Odorifera T. Chen Leaves by Deep Eutectic Solvents-Based Negative Pressure Cavitation Extraction Followed by Macroporous Resin Column Chromatography. *Journal of Chromatography B*, 1033, 40-48.
- Liang, J., Zeng, Y., Wang, H., & Lou, W. (2019). Extraction, Purification and Antioxidant Activity of Novel Polysaccharides from Dendrobium Officinale by Deep Eutectic Solvents. *Natural product research*, 33(22), 3248-3253.
- Liao, H., Banbury, L. K., & Leach, D. N. (2008). Antioxidant Activity of 45 Chinese Herbs and the Relationship with Their Tcm Characteristics. *Evidence-Based Complementary and Alternative Medicine*, 5(4), 429-434.
- Lirio, S. B., Macabeo, A. P. G., Paragas, E. M., Knorn, M., Kohls, P., Franzblau, S. G., Wang, Y., & Aguinaldo, M. A. M. (2014). Antitubercular Constituents

- from Premna Odorata Blanco. *Journal of Ethnopharmacology*, 154(2), 471-474.
- Liu, Y., Friesen, J. B., McAlpine, J. B., Lankin, D. C., Chen, S.-N., & Pauli, G. F. (2018). Natural Deep Eutectic Solvents: Properties, Applications, and Perspectives. *Journal of natural products*, 81(3), 679-690.
- López-Torres, I. I., Vaquero-Martín, J., Torres-Suárez, A.-I., Navarro-García, F., Fraguas-Sánchez, A.-I., León-Román, V. E., & Sanz-Ruiz, P. (2022). The Tale of Microencapsulated Rifampicin: Is It Useful for the Treatment of Periprosthetic Joint Infection? *International Orthopaedics*, 1-9.
- Luetragoon, T., Pankla Sranujit, R., Noysang, C., Thongsri, Y., Potup, P., Suphrom, N., Nuengchamnong, N., & Usuwanthim, K. (2020a). Anti-Cancer Effect of 3-Hydroxy-B-Ionone Identified from Moringa Oleifera Lam. Leaf on Human Squamous Cell Carcinoma 15 Cell Line. *Molecules*, 25(16), 3563.
- Luetragoon, T., Pankla Sranujit, R., Noysang, C., Thongsri, Y., Potup, P., Suphrom, N., Nuengchamnong, N., & Usuwanthim, K. (2020b). Bioactive Compounds in Moringa Oleifera Lam. Leaves Inhibit the Pro-Inflammatory Mediators in Lipopolysaccharide-Induced Human Monocyte-Derived Macrophages. *Molecules*, 25(1), 191.
- Ma, Z., Lienhardt, C., McIllemon, H., Nunn, A. J., & Wang, X. (2010). Global Tuberculosis Drug Development Pipeline: The Need and the Reality. *The Lancet*, 375(9731), 2100-2109.
- Machmudah, S., Lestari, S. D., Kanda, H., Winardi, S., & Goto, M. (2018). Subcritical Water Extraction Enhancement by Adding Deep Eutectic Solvent for Extracting Xanthone from Mangosteen Pericarps. *The Journal of Supercritical Fluids*, 133, 615-624.
- Mahdi, H. J., Khan, N. A. K., Asmawi, M. Z. B., Mahmud, R., Vikneswaran, A., & Murugaiyah, L. (2018). In Vivo Anti-Arthritic and Anti-Nociceptive Effects of Ethanol Extract of Moringa Oleifera Leaves on Complete Freund's Adjuvant (Cfa)-Induced Arthritis in Rats. *Integrative medicine research*, 7(1), 85-94.
- Maher, T., Ahmad Raus, R., Daddiouaissa, D., Ahmad, F., Adzhar, N. S., Latif, E. S., Abdulhafiz, F., & Mohammed, A. (2021). Medicinal Plants with Anti-Leukemic Effects: A Review. *Molecules*, 26(9), 2741.
- Maisetta, G., Batoni, G., Esin, S., Florio, W., Bottai, D., Favilli, F., & Campa, M. (2006). In Vitro Bactericidal Activity of Human B-Defensin 3 against

- Multidrug-Resistant Nosocomial Strains. *Antimicrobial agents and chemotherapy*, 50(2), 806-809.
- Majekodunmi, S. O. (2015). Review of Extraction of Medicinal Plants for Pharmaceutical Research. *Merit research journal of medicine and medical sciences*, 11, 521-527.
- Mali, J. K., Sutar, Y. B., Pahelkar, A. R., Verma, P. M., & Telvekar, V. N. (2020). Novel Fatty Acid-Thiadiazole Derivatives as Potential Antimycobacterial Agents. *Chemical Biology & Drug Design*, 95(1), 174-181.
- Manilal, A., Sabu, K. R., Woldemariam, M., Aklilu, A., Biresaw, G., Yohanes, T., Seid, M., & Merdekios, B. (2021). Antibacterial Activity of Rosmarinus Officinalis against Multidrug-Resistant Clinical Isolates and Meat-Borne Pathogens. *Evidence-Based Complementary and Alternative Medicine*, 2021.
- Mann, A., Ibrahim, K., Oyewale, A. O., Amupitan, J. O., Fatope, M. O., & Okogun, J. I. (2011). Antimycobacterial Friedelane-Terpenoid from the Root Bark of Terminalia Avicennioides. *American Journal of Chemistry*, 1(2), 52-55.
- Mariita, R. M., Orodho, J. A., Okemo, P. O., Kirimuhuza, C., Otieno, J. N., & Magadula, J. J. (2011). Methanolic Extracts of Aloe Secundiflora Engl. Inhibits in Vitro Growth of Tuberculosis and Diarrhea-Causing Bacteria. *Pharmacognosy research*, 3(2), 95.
- Mathew, T., Ovsyanikova, T., Shin, S., Gelmanova, I., Balbuena, D., Atwood, S., Peremitin, G., Strelis, A., & Murray, M. (2006). Causes of Death During Tuberculosis Treatment in Tomsk Oblast, Russia. *The International Journal of Tuberculosis and Lung Disease*, 10(8), 857-863.
- McGaw, L., Lall, N., Meyer, J., & Eloff, J. (2008). The Potential of South African Plants against Mycobacterium Infections. *Journal of Ethnopharmacology*, 119(3), 482-500.
- Medjahed, H., Gaillard, J.-L., & Reyrat, J.-M. (2010). Mycobacterium Abscessus: A New Player in the Mycobacterial Field. *Trends in microbiology*, 18(3), 117-123.
- Mehwish, H. M., Liu, G., Rajoka, M. S. R., Cai, H., Zhong, J., Song, X., Xia, L., Wang, M., Aadil, R. M., & Inam-Ur-Raheem, M. (2021). Therapeutic Potential of Moringa Oleifera Seed Polysaccharide Embedded Silver Nanoparticles in Wound Healing. *International journal of biological macromolecules*, 184, 144-158.

- Meireles, D., Gomes, J., Lopes, L., Hinzmann, M., & Machado, J. (2020). A Review of Properties, Nutritional and Pharmaceutical Applications of Moringa Oleifera: Integrative Approach on Conventional and Traditional Asian Medicine. *Advances in Traditional Medicine*, 20(4), 495-515.
- Mena-García, A., Ruiz-Matute, A. I., Soria, A. C., & Sanz, M. L. (2019). Green Techniques for Extraction of Bioactive Carbohydrates. *TrAC Trends in Analytical Chemistry*, 119, 115612.
- Mendoza, R. A., Shen, C.-C., Tsai, P.-W., & De Castro-Cruz, K. A. (2021). Bioassay-Guided Isolation and Structure Elucidation of Bioactive Phytoconstituents with Inhibitory Activity against Carbohydrate-Hydrolyzing Enzymes from the Aerial Parts of Premna Odorata Blanco. *INDIAN JOURNAL OF PHARMACEUTICAL EDUCATION AND RESEARCH*, 55(3), 846-856.
- Minh, T. N., Minh, B. Q., Duc, T. H. M., Thinh, P. V., Anh, L. V., Dat, N. T., Nhan, L. V., & Trung, N. Q. (2022). Potential Use of Moringa Oleifera Twigs Extracts as an Anti-Hyperuricemic and Anti-Microbial Source. *Processes*, 10(3), 563.
- Mohamad, S., Ismail, N. N., Parumasivam, T., Ibrahim, P., Osman, H., & A Wahab, H. (2018). Antituberculosis Activity, Phytochemical Identification of Costus Speciosus (J. Koenig) Sm., Cymbopogon Citratus (Dc. Ex Nees) Stapf., and Tabernaemontana Coronaria (L.) Willd. And Their Effects on the Growth Kinetics and Cellular Integrity of Mycobacterium Tuberculosis H37rv. *BMC complementary and alternative medicine*, 18(1), 1-14.
- Mohamad, S., Zin, N. M., Wahab, H. A., Ibrahim, P., Sulaiman, S. F., Zahariluddin, A. S. M., & Noor, S. S. M. (2011). Antituberculosis Potential of Some Ethnobotanically Selected Malaysian Plants. *Journal of Ethnopharmacology*, 133(3), 1021-1026.
- Mohapatra, G. C., Khan, M. J., & Nayak, S. (2021). Incidence of Hyperuricemia and Gouty Arthritis in Patients Taking Pyrazinamide for the Treatment of Tuberculosis. *Annals of the Romanian Society for Cell Biology*, 25(6), 324–328-324–328.
- Mokhtar, K. S., Rahman, N., Shariff, N., & Nor, W. (2012). Tuberculosis in Malaysia: A Study on the Level of Societal Awareness and Stigma. *IOSR Journal of Humanities and Social Science*, 1(4), 59-64.

- Mollejon, B. C., & Mollejon, C. V. (2019). Anti-Microbial Activities and Phytochemical Screening of the Premna Odorata Blanco (Alagaw) Leaf Extract.
- MONRE. (2014). Fifth Report to Convention on Biological Diversity. <https://www.cbd.int/doc/world/my/my-nr-05-en.pdf>.
- Morais, T., Cotas, J., Pacheco, D., & Pereira, L. (2021). Seaweeds Compounds: An Ecosustainable Source of Cosmetic Ingredients? *Cosmetics*, 8(1), 8.
- Mostafavi, S., Asadi-Gharneh, H. A., & Miransari, M. (2019). The Phytochemical Variability of Fatty Acids in Basil Seeds (*Ocimum Basilicum* L.) Affected by Genotype and Geographical Differences. *Food chemistry*, 276, 700-706.
- Mtewa, A. G., Deyno, S., Kasali, F. M., Annu, A., & Sesaazi, D. C. (2018). General Extraction, Isolation and Characterization Techniques in Drug Discovery: A Review. *Int. J. Sci. Basic Appl. Res*, 38(1), 10-24.
- Murray, J. F. (2004). A Century of Tuberculosis. *American journal of respiratory and critical care medicine*, 169(11), 1181-1186.
- Mushtaq, M. U., Shahid, U., Abdullah, H. M., Saeed, A., Omer, F., Shad, M. A., Siddiqui, A. M., & Akram, J. (2011). Urban-Rural Inequities in Knowledge, Attitudes and Practices Regarding Tuberculosis in Two Districts of Pakistan's Punjab Province. *International journal for equity in health*, 10(1), 1-9.
- Nadar, S. S., Rao, P., & Rathod, V. K. (2018). Enzyme Assisted Extraction of Biomolecules as an Approach to Novel Extraction Technology: A Review. *Food Research International*, 108, 309-330.
- Newton, S. M., Lau, C., Gurcha, S. S., Besra, G. S., & Wright, C. W. (2002). The Evaluation of Forty-Three Plant Species for in Vitro Antimycobacterial Activities; Isolation of Active Constituents from *Psoralea Corylifolia* and *Sanguinaria Canadensis*. *Journal of Ethnopharmacology*, 79(1), 57-67.
- Nguta, J. M., Appiah-Opong, R., Nyarko, A. K., Yeboah-Manu, D., & Addo, P. G. (2015). Current Perspectives in Drug Discovery against Tuberculosis from Natural Products. *International Journal of Mycobacteriology*, 4(3), 165-183.
- Nguta, J. M., Appiah-Opong, R., Nyarko, A. K., Yeboah-Manu, D., Addo, P. G., Otchere, I. D., & Kissi-Twum, A. (2016). In Vitro Antimycobacterial and Cytotoxic Data on Medicinal Plants Used to Treat Tuberculosis. *Data in brief*, 7, 1124-1130.

- Nguyen, H. C., Nguyen, H. N. T., Huang, M. Y., Lin, K. H., Pham, D. C., Tran, Y. B., & Su, C. H. (2021). Optimization of Aqueous Enzyme-Assisted Extraction of Rosmarinic Acid from Rosemary (*Rosmarinus Officinalis L.*) Leaves and the Antioxidant Activity of the Extract. *Journal of Food Processing and Preservation*, 45(3), e15221.
- Nguyen, T.-T., Ketha, A., Hieu, H. V., & Tatipamula, V. B. (2021). In Vitro Antimycobacterial Studies of Flavonols from *Bauhinia Vahlii* Wight and Arn. *3 Biotech*, 11(3), 1-5.
- Nielsen, E. I., Viberg, A., Lo, E., Cars, O., Karlsson, M. O., & Sandstro, M. (2007). Semimechanistic Pharmacokinetic/ Pharmacodynamic Model for Assessment of Activity of Antibacterial Agents from Time-Kill Curve Experiments. *Antimicrobial agents and chemotherapy*, 51, 128-136.
- Nieto, G., Ros, G., & Castillo, J. (2018). Antioxidant and Antimicrobial Properties of Rosemary (*Rosmarinus Officinalis*, L.): A Review. *Medicines*, 5(3), 98.
- NM, N. N. R., Mohd, N., Sharina, D., & NH, N. R. (2011). Factors Associated with Unsuccessful Treatment Outcome of Pulmonary Tuberculosis in Kota Bharu, Kelantan. *Malaysian Journal of Public Health Medicine*, 11(1), 6-15.
- Noumi, V. D., Nguimbou, R. M., Tsague, M. V., Deli, M., Rup-Jacques, S., Amadou, D., Sokeng, S., & Njintang, N. Y. (2021). Phytochemical Profile and in Vitro Antioxidant Properties of Essential Oils from Powder Fractions of *Eucalyptus Camaldulensis* Leaves. *American Journal of Plant Sciences*, 12(03), 329.
- Nwaogu, J., Argungu, A., & Adefarakan, A. J. (2021). Antiulcer and Antioxidant Potential of *Eucalyptus Camaldulensis* Leaves Methanol Extract in Albino Rats. *Asian Journal of Biochemistry, Genetics and Molecular Biology*, 39-46.
- Nyambuya, T., Mautsa, R., & Mukanganyama, S. (2017). Alkaloid Extracts from *Combretum Zeyheri* Inhibit the Growth of *Mycobacterium Smegmatis*. *BMC complementary and alternative medicine*, 17(1), 1-11.
- Ohiagu, F. O., Chikezie, P. C., Chikezie, C. M., & Enyoh, C. E. (2021). Anticancer Activity of Nigerian Medicinal Plants: A Review. *Future Journal of Pharmaceutical Sciences*, 7(1), 1-21.
- Ojo, S., Ejims-Enukwe, O., & Esumeh, F. (2013). In-Vitro Antibacterial Time-Kill Assay of *Phyllanthus Amarus* and *Diodia Scandens* Crude Extracts on *Staphylococci* Isolated from Wounds and Burns Patients. *Int J Pharm Sci Invent*, 2, 9-13.

- Oladosu, P., Isu, N., Ibrahim, K., Okolo, P., & Oladepo, D. (2013). Time Kill-Kinetics Antibacterial Study of Acacia Nilotica. *Afr. J. Microbiol. Res.*, 7(46), 5248-5252.
- Oladosu, P., Isu, N. R., Ibrahim, K., Okolo, P., & Oladepo, D. K. (2013). Time Kill Kinetics Antibacterial Study of Acacia Nilotica. *Afr. J. Microbiol. Res.*, 7, 5248-5252.
- Olajuyigbe, O. O., & Afolayan, A. J. . (2012). In Vitro Antibacterial and Time-Kill Evaluation of the Erythrina Caffra Thunb. Extract against Bacteria Associated with Diarrhea. *The ScientificWorld Journal*.
- Olivoto, T., Nardino, M., Carvalho, I. R., Follmann, D. N., Szareski, V., Jardel, i., Ferrari, M., de Pelegrin, A. J., de Souza, V. Q., & oz. (2017). Plant Secondary Metabolites and Its Dynamical Systems of Induction in Response to Environmental Factors: A Review. *African Journal of Agricultural Research*, 12(2), 71-84.
- Organisation, G. W. H. (2019). Global Tuberculosis Report. *Ann. ICRP*, 39.
- Organization, W. H. (2016). Global Tuberculosis Report 2016.
- Othman, M. S., Khaled, A. M., Al-Bagawi, A. H., Fareid, M. A., Ghany, R. A., Habotta, O. A., & Moneim, A. E. A. (2021). Hepatorenal Protective Efficacy of Flavonoids from Ocimum Basilicum Extract in Diabetic Albino Rats: A Focus on Hypoglycemic, Antioxidant, Anti-Inflammatory and Anti-Apoptotic Activities. *Biomedicine & Pharmacotherapy*, 144, 112287.
- Ou, B., Huang, D., Hampsch-Woodill, M., & Flanagan, J. A. (2003). When East Meets West: The Relationship between Yin-Yang and Antioxidation-Oxidation. *The FASEB journal*, 17(2), 127-129.
- Pal, R., Hameed, S., & Fatima, Z. (2015). Iron Deprivation Affects Drug Susceptibilities of Mycobacteria Targeting Membrane Integrity. *Journal of pathogens*, 2015.
- Pal, S. K., & Shukla, Y. (2003). Herbal Medicine: Current Status and the Future. *Asian pacific journal of cancer prevention*, 4(4), 281-288.
- Pandey, A., & Pandey, S. (2014). Solvatochromic Probe Behavior within Choline Chloride-Based Deep Eutectic Solvents: Effect of Temperature and Water. *The Journal of Physical Chemistry B*, 118(50), 14652-14661.

- Pandey, A., Rai, R., Pal, M., & Pandey, S. (2014). How Polar Are Choline Chloride-Based Deep Eutectic Solvents? *Physical Chemistry Chemical Physics*, 16(4), 1559-1568.
- Pandey, A., & Tripathi, S. (2014). Concept of Standardization, Extraction and Pre Phytochemical Screening Strategies for Herbal Drug. *Journal of Pharmacognosy and Phytochemistry*, 2(5).
- Pandey, A. K., & Tripathi, Y. (2017). Ethnobotany and Its Relevance in Contemporary Research. *Journal of Medicinal Plants Studies*, 5(3), 123-129.
- Panić, M., Stojković, M. R., Kraljić, K., Škevin, D., Redovniković, I. R., Srček, V. G., & Radošević, K. (2019). Ready-to-Use Green Polyphenolic Extracts from Food by-Products. *Food chemistry*, 283, 628-636.
- Pauli, G. F., Case, R. J., Inui, T., Wang, Y., Cho, S., Fischer, N. H., & Franzblau, S. G. (2005). New Perspectives on Natural Products in Tb Drug Research. *Life sciences*, 78(5), 485-494.
- Peng, X., Duan, M.-H., Yao, X.-H., Zhang, Y.-H., Zhao, C.-J., Zu, Y.-G., & Fu, Y.-J. (2016). Green Extraction of Five Target Phenolic Acids from Lonicerae Japonicae Flos with Deep Eutectic Solvent. *Separation and Purification Technology*, 157, 249-257.
- Podolak, I., Galanty, A., & Sobolewska, D. (2010). Saponins as Cytotoxic Agents: A Review. *Phytochemistry Reviews*, 9(3), 425-474.
- Popoola, J. O., & Obembe, O. O. (2013). Local Knowledge, Use Pattern and Geographical Distribution of Moringa Oleifera Lam.(Moringaceae) in Nigeria. *Journal of Ethnopharmacology*, 150(2), 682-691.
- Prasad, R., & Srivastava, D. K. (2013). Multi Drug and Extensively Drug-Resistant Tb (M/Xdr-Tb) Management: Current Issues. *Clinical epidemiology and global health*, 1(3), 124-128.
- Prasasty, V. D., Cindana, S., Ivan, F. X., Zahroh, H., & Sinaga, E. (2020). Structure-Based Discovery of Novel Inhibitors of Mycobacterium Tuberculosis Cyp121 from Indonesian Natural Products. *Computational Biology and Chemistry*, 85, 107205.
- Qureshi, K. A., Imtiaz, M., Parvez, A., Rai, P. K., Jaremko, M., Emwas, A.-H., Bholay, A. D., & Fatmi, M. Q. (2022). In Vitro and in Silico Approaches for the Evaluation of Antimicrobial Activity, Time-Kill Kinetics, and Anti-Biofilm

- Potential of Thymoquinone (2-Methyl-5-Propan-2-Ylcyclohexa-2, 5-Diene-1, 4-Dione) against Selected Human Pathogens. *Antibiotics*, 11(1), 79.
- Rabiee, N., Bagherzadeh, M., Kiani, M., & Ghadiri, A. M. (2020). Rosmarinus Officinalis Directed Palladium Nanoparticle Synthesis: Investigation of Potential Anti-Bacterial, Anti-Fungal and Mizoroki-Heck Catalytic Activities. *Advanced Powder Technology*, 31(4), 1402-1411.
- Racchi, M. L. (2013). Antioxidant Defenses in Plants with Attention to Prunus and Citrus Spp. *Antioxidants*, 2(4), 340-369.
- Radji, M., Kurniati, M., & Kiranasari, A. (2015). Comparative Antimycobacterial Activity of Some Indonesian Medicinal Plants against Multi-Drug Resistant Mycobacterium Tuberculosis. *Journal of Applied Pharmaceutical Science*, 5(1), 019-022.
- Radošević, K., Čurko, N., Srček, V. G., Bubalo, M. C., Tomašević, M., Ganić, K. K., & Redovniković, I. R. (2016). Natural Deep Eutectic Solvents as Beneficial Extractants for Enhancement of Plant Extracts Bioactivity. *LWT*, 73, 45-51.
- Rafiee, S., Besharat, S., Jabbari, A., Golalipour, F., & Nasermoaadeli, A. (2009). Epidemiology of Tuberculosis in Northeast of Iran: A Population-Based Study. *Iranian Journal of Medical Sciences*, 34(3), 193-197.
- Rafiza, S., Rampal, K. G., & Tahir, A. (2011). Prevalence and Risk Factors of Latent Tuberculosis Infection among Health Care Workers in Malaysia. *BMC infectious diseases*, 11(1), 19.
- Raghunandan, S., Jose, L., Gopinath, V., & Kumar, R. A. (2019). Comparative Label-Free Lipidomic Analysis of Mycobacterium Tuberculosis During Dormancy and Reactivation. *Scientific reports*, 9(1), 3660.
- Rahgozar, N., Khaniki, G. B., & Sardari, S. (2018). Evaluation of Antimycobacterial and Synergistic Activity of Plants Selected Based on Cheminformatic Parameters. *Iranian biomedical journal*, 22(6), 401.
- Rahimifard, M., Maqbool, F., Moeini-Nodeh, S., Niaz, K., Abdollahi, M., Braidy, N., Nabavi, S. M., & Nabavi, S. F. (2017). Targeting the Tlr4 Signaling Pathway by Polyphenols: A Novel Therapeutic Strategy for Neuroinflammation. *Ageing research reviews*, 36, 11-19.
- Rajha, H. N., Mhanna, T., El Kantar, S., El Khoury, A., Louka, N., & Maroun, R. G. (2019). Innovative Process of Polyphenol Recovery from Pomegranate Peels

- by Combining Green Deep Eutectic Solvents and a New Infrared Technology. *LWT*, 111, 138-146.
- Rathnasamy, S. K., sri Rajendran, D., Balaraman, H. B., & Viswanathan, G. (2019). Functional Deep Eutectic Solvent-Based Chaotic Extraction of Phycobiliprotein Using Microwave-Assisted Liquid-Liquid Micro-Extraction from Spirulina (Arthrospira Platensis) and Its Biological Activity Determination. *Algal Research*, 44, 101709.
- Raviglione, M. C., & Pio, A. (2002). Evolution of Who Policies for Tuberculosis Control, 1948–2001. *The Lancet*, 359(9308), 775-780.
- Reid, A., Oosthuizen, C. B., & Lall, N. (2020). In Vitro Antimycobacterial and Adjuvant Properties of Two Traditional South African Teas, Aspalathus Linearis (Burm. F.) R. Dahlgren and Lippia Scaberrima Sond. *South African Journal of Botany*, 128, 257-263.
- Reyes-García, V. (2010). The Relevance of Traditional Knowledge Systems for Ethnopharmacological Research: Theoretical and Methodological Contributions. *Journal of ethnobiology and ethnomedicine*, 6(1), 32.
- Robles-Zepeda, R. E., Coronado-Aceves, E. W., Velázquez-Contreras, C. A., Ruiz-Bustos, E., Navarro-Navarro, M., & Garibay-Escobar, A. (2013). In Vitro Anti-Mycobacterial Activity of Nine Medicinal Plants Used by Ethnic Groups in Sonora, Mexico. *BMC complementary and alternative medicine*, 13(1), 329.
- Rodriguez-Fragoso, L., Reyes-Esparza, J., Burchiel, S. W., Herrera-Ruiz, D., & Torres, E. (2008). Risks and Benefits of Commonly Used Herbal Medicines in Mexico. *Toxicology and applied pharmacology*, 227(1), 125-135.
- Rodriguez-Rivera, F. P., Zhou, X., Theriot, J. A., & Bertozzi, C. R. (2017). Visualization of Mycobacterial Membrane Dynamics in Live Cells. *Journal of the American Chemical Society*, 139(9), 3488-3495.
- Rubab, S., Bahadur, S., Hanif, U., Durrani, A. I., Sadiqa, A., Shafique, S., Zafar, U., Shuaib, M., Urooj, Z., & Nizamani, M. M. (2021). Phytochemical and Antimicrobial Investigation of Methanolic Extract/Fraction of Ocimum Basilicum L. *Biocatalysis and Agricultural Biotechnology*, 31, 101894.
- Rugutt, J. K., & Rugutt, K. J. (2012). Antimycobacterial Activity of Steroids, Long-Chain Alcohols and Lytic Peptides. *Natural product research*, 26(11), 1004-1011.

- Ryter, S. W., & Choi, A. M. (2015). Autophagy in Lung Disease Pathogenesis and Therapeutics. *Redox biology*, 4, 215-225.
- Sabo, V. A., & Knezevic, P. (2019). Antimicrobial Activity of Eucalyptus Camaldulensis Dehn. Plant Extracts and Essential Oils: A Review. *Industrial Crops and Products*, 132, 413-429.
- Sabran, S. F. (2016). *Documentation, Antimycobacterial Activity, and Phytochemical Profiling of Selected Medicinal Plants Used by the Jakun Community in Johor*. Universiti Tun Hussein Onn Malaysia,
- Sabran, S. F., Mohamed, M., Bakar, A., & Fadzelly, M. (2016). Ethnomedical Knowledge of Plants Used for the Treatment of Tuberculosis in Johor, Malaysia. *Evidence-Based Complementary and Alternative Medicine*, 2016.
- Sajid, A., Arora, G., Virmani, R., & Singhal, A. (2017). Antimycobacterial Agents: To Target or Not to Target. In *Microbial Applications Vol. 2* (pp. 83-104): Springer.
- Sakamoto, K. (2012). The Pathology of Mycobacterium Tuberculosis Infection. *Veterinary pathology*, 49(3), 423-439.
- Salim, A. A., Chin, Y.-W., & Kinghorn, A. D. (2008). Drug Discovery from Plants. In *Bioactive Molecules and Medicinal Plants* (pp. 1-24): Springer.
- Santos, L., Silva, P., Moura, M., Junior, A. C., Amorim, P., Procópio, T., Coelho, L., Silva, L., Paiva, P., & Santos, N. (2021). Anti-Candida Activity of the Water-Soluble Lectin from Moringa Oleifera Seeds (Wsmol). *Journal of Medical Mycology*, 31(2), 101074.
- Sanusi, S. B., Abu Bakar, M. F., Mohamed, M., Sabran, S. F., & Mainasara, M. M. (2017). Southeast Asian Medicinal Plants as a Potential Source of Antituberculosis Agent. *Evidence-Based Complementary and Alternative Medicine*, 2017.
- Sasikumar, K., Ghosh, A. R., & Dushthackeer, A. (2018). Antimycobacterial Potentials of Quercetin and Rutin against Mycobacterium Tuberculosis H37rv. *3 Biotech*, 8(10), 1-6.
- Selmi, S., Rtibi, K., Grami, D., Sebai, H., & Marzouki, L. (2017). Rosemary (*Rosmarinus Officinalis*) Essential Oil Components Exhibit Anti-Hyperglycemic, Anti-Hyperlipidemic and Antioxidant Effects in Experimental Diabetes. *Pathophysiology*, 24(4), 297-303.

- Severino, V. G. P., Felix, M. A., Silva, M. F. d. G. F. d., Lucarini, R., & Martins, C. H. G. (2015). Chemical Study of Hortia Superba (Rutaceae) and Investigation of the Antimycobacterial Activity of Crude Extracts and Constituents Isolated from Hortia Species. *Química Nova*, 38, 42-45.
- Shafie, M. H., Yusof, R., & Gan, C.-Y. (2019). Deep Eutectic Solvents (Des) Mediated Extraction of Pectin from Averrhoa Bilimbi: Optimization and Characterization Studies. *Carbohydrate polymers*, 216, 303-311.
- Shahrajabian, M. H., Sun, W., & Cheng, Q. (2020). Chemical Components and Pharmacological Benefits of Basil (Ocimum Basilicum): A Review. *International Journal of Food Properties*, 23(1), 1961-1970.
- Shang, X., Dou, Y., Zhang, Y., Tan, J.-N., Liu, X., & Zhang, Z. (2019). Tailor-Made Natural Deep Eutectic Solvents for Green Extraction of Isoflavones from Chickpea (Cicer Arietinum L.) Sprouts. *Industrial Crops and Products*, 140, 111724.
- sharma, D., & Yadav,J. (2016). An Overview Phytotherapeutic Approaches for the Treatment of Tuberculosis Mini-Review *medicinal chemistry* 17(2), 167-183.
- Sharma, S., Hameed, S., & Fatima, Z. (2020). Lipidomic Insights to Understand Membrane Dynamics in Response to Vanillin in Mycobacterium Smegmatis. *International Microbiology*, 23(2), 263-276.
- Shen, D., Kou, X., Wu, C., Fan, G., Li, T., Dou, J., Wang, H., & Zhu, J. (2021). Cocktail Enzyme-Assisted Alkaline Extraction and Identification of Jujube Peel Pigments. *Food chemistry*, 357, 129747.
- Shi, Y.-P., Rodríguez, A. D., Barnes, C. L., Sánchez, J. A., Raptis, R. G., & Baran, P. (2002). New Terpenoid Constituents from Eunicea P Inta. *Journal of natural products*, 65(9), 1232-1241.
- Silva, F., Lourenço, O., Queiroz, J. A., & Domingues, F. C. (2011). Bacteriostatic Versus Bactericidal Activity of Ciprofloxacin in Escherichia Coli Assessed by Flow Cytometry Using a Novel Far-Red Dye. *The Journal of antibiotics*, 64(4), 321.
- Singh, A. K., Rana, H. K., Tshabalala, T., Kumar, R., Gupta, A., Ndhlala, A. R., & Pandey, A. K. (2020). Phytochemical, Nutraceutical and Pharmacological Attributes of a Functional Crop Moringa Oleifera Lam: An Overview. *South African Journal of Botany*, 129, 209-220.

- Singh, R., Hussain, S., Verma, R., & Sharma, P. (2013). Anti-Mycobacterial Screening of Five Indian Medicinal Plants and Partial Purification of Active Extracts of Cassia Sophera and Urtica Dioica. *Asian Pacific journal of tropical medicine*, 6(5), 366-371.
- Singh, R., Kakkar, A., & Mishra, V. K. (2015). Anti Tuberculosis Activity and Gc-Ms Analysis of Ethyl Acetate Extracts of Wrightia Tinctoria Bark. *World Journal of Pharmaceutical Research*, 4(08), 1938-1948.
- Skandamis, P., Tsigarida, E., & Nychas, G.-J. (2000). Ecophysiological Attributes of *Salmonella Typhimurium* in Liquid Culture and within a Gelatin Gel with or without the Addition of Oregano Essential Oil. *World Journal of Microbiology and Biotechnology*, 16(1), 31-35.
- Śliwka, L., Wiktorska, K., Suchocki, P., Milczarek, M., Mielczarek, S., Lubelska, K., Cierpiał, T., Łyżwa, P., Kiełbasiński, P., & Jaromin, A. (2016). The Comparison of Mtt and Cvs Assays for the Assessment of Anticancer Agent Interactions. *PLoS One*, 11(5), e0155772.
- Smith, E. L., Abbott, A. P., & Ryder, K. S. (2014). Deep Eutectic Solvents (Dess) and Their Applications. *Chemical reviews*, 114(21), 11060-11082.
- Smith, I. (2003). Mycobacterium Tuberculosis Pathogenesis and Molecular Determinants of Virulence. *Clinical microbiology reviews*, 16(3), 463-496.
- Solliman, M. A., Hassali, M. A., Al-Haddad, M., Hadida, M. M., Saleem, F., Atif, M., & Aljadhey, H. (2012). Assessment of Knowledge Towards Tuberculosis among General Population in North East Libya. *Journal of Applied Pharmaceutical Science*, 2(4), 24.
- Stefanović, O., Radojević, I., Vasić, S., & Čomić, L. (2012). Antibacterial Activity of Naturally Occurring Compounds from Selected Plants. *Antimicrobial Agents*, 1-25.
- Stohs, S. J., & Hartman, M. J. (2015). Review of the Safety and Efficacy of Moringa Oleifera. *Phytotherapy Research*, 29(6), 796-804.
- Street, R., Stirk, W., & Van Staden, J. (2008). South African Traditional Medicinal Plant Trade—Challenges in Regulating Quality, Safety and Efficacy. *Journal of Ethnopharmacology*, 119(3), 705-710.
- Stuckler, D., Basu, S., McKee, M., & Lurie, M. (2011). Mining and Risk of Tuberculosis in Sub-Saharan Africa. *American journal of public health*, 101(3), 524-530.

- Sun, T., Qin, B., Gao, M., Yin, Y., Wang, C., Zang, S., Li, X., Zhang, C., Xin, Y., & Jiang, T. (2015). Effects of Epigallocatechin Gallate on the Cell-Wall Structure of Mycobacterial Smegmatis Mc2155. *Natural product research*, 29(22), 2122-2124.
- Sunthitikawinsakul, A., Kongkathip, N., Kongkathip, B., Phonnakhu, S., Daly, J. W., Spande, T. F., Nimit, Y., & Rochanaruangrai, S. (2003). Coumarins and Carbazoles from Clausena Excavata Exhibited Antimycobacterial and Antifungal Activities. *Planta Medica*, 69(02), 155-157.
- Svendsen, B. A., & Verpoorte, R. (1983). *Chromatography of Alkaloids* (Vol. 23): New York: Elsevier Scientific Publishing Company.
- Tabuti, J. R., Kukunda, C. B., & Waako, P. J. (2010). Medicinal Plants Used by Traditional Medicine Practitioners in the Treatment of Tuberculosis and Related Ailments in Uganda. *Journal of Ethnopharmacology*, 127(1), 130-136.
- Tachfouti, N., Slama, K., Berraho, M., & Nejjari, C. (2012). The Impact of Knowledge and Attitudes on Adherence to Tuberculosis Treatment: A Case-Control Study in a Moroccan Region. *Pan African Medical Journal*, 12(1).
- Takla, S. S., Shawky, E., Hammoda, H. M., & Darwish, F. A. (2018). Green Techniques in Comparison to Conventional Ones in the Extraction of Amaryllidaceae Alkaloids: Best Solvents Selection and Parameters Optimization. *Journal of Chromatography A*, 1567, 99-110.
- Talaro, K. P., & Chess, B. (2015). *Mycobacteria: Acid-Fast Bacilli Foundations in Microbiology* New York:: MacGraw-Hill.
- Talip, B. A., Sleator, R. D., Lowery, C. J., Dooley, J. S., & Snelling, W. J. (2013). An Update on Global Tuberculosis (Tb). *Infectious Diseases: Research and Treatment*, 6, IDRT. S11263.
- Tam, V. H., Schilling, A. N., & Nikolaou, M. (2005). Modelling Time–Kill Studies to Discern the Pharmacodynamics of Meropenem. *Journal of antimicrobial chemotherapy*, 55(5), 699-706.
- Tamaru, A., Nakajima, C., Wada, T., Wang, Y., Inoue, M., Kawahara, R., Maekura, R., Ozeki, Y., Ogura, H., & Kobayashi, K. (2012). Dominant Incidence of Multidrug and Extensively Drug-Resistant Specific Mycobacterium Tuberculosis Clones in Osaka Prefecture, Japan.
- Tandon, S., & Rane, S. (2008). Decoction and Hot Continuous Extraction Techniques. *Extraction technologies for medicinal and aromatic plants*, 93.

- Tang, B., & Row, K. H. (2013). Recent Developments in Deep Eutectic Solvents in Chemical Sciences. *Monatshefte für Chemie-Chemical Monthly*, 144(10), 1427-1454.
- Tangjitjaroenkun, J., Chavasiri, W., Thunyaharn, S., & Yompakdee, C. (2012). Bactericidal Effects and Time-Kill Studies of the Essential Oil from the Fruits of *Zanthoxylum Limonella* on Multi-Drug Resistant Bacteria. *Journal of Essential Oil Research*, 24(4), 363-370.
- Tariq, A., Mussarat, S., Adnan, M., Abd\_Allah, E., Hashem, A., Alqarawi, A. A., & Ullah, R. (2015). Ethnomedicinal Evaluation of Medicinal Plants Used against Gastrointestinal Complaints. *BioMed research international*, 2015.
- Teofilović, B., Tomas, A., Martić, N., Stilinović, N., Popović, M., Čapo, I., Grujić, N., Ilinčić, B., & Rašković, A. (2021). Antioxidant and Hepatoprotective Potential of Sweet Basil (*Ocimum Basilicum L.*) Extract in Acetaminophen-Induced Hepatotoxicity in Rats. *Journal of Functional Foods*, 87, 104783.
- Tepsorn, R. (2009). Antimicrobial Activity of Thai Traditional Medicinal Plants Extract Incorporated Alginate-Tapioca Starch Based Edible Films against Food Related Bacteria Including Foodborne Pathogens.
- Terry K. Means, S. W., \* Egil Lien, † Atsutoshi Yoshimura, † Douglas T. Golenbock, † and Matthew J. Fenton2\*. (1999). Human Toll-Like Receptors Mediate Cellular Activation by *Mycobacterium Tuberculosis*1. *The Journal of Immunology*, 162, 3920-3927.
- Thomas, C., Newell, J. N., Baral, S. C., & Byanjankar, L. (2007). The Contribution of Volunteers to a Successful Community-Orientated Tuberculosis Treatment Centre in an Urban Setting in Nepal: A Qualitative Assessment of Volunteers' Roles and Motivations. *Journal of health organization and management*.
- Tiam, E. R., Ngono Bikobo, D. S., Abouem A Zintchem, A., Mbabi Nyemeck, N., Moni Ndidi, E. D. F., Betote Diboué, P. H., Nyegue, M. A., Atchadé, A. d. T., Emmanuel Pegnyemb, D., & Bochet, C. G. (2019). Secondary Metabolites from *Triclisia Gilletii* (De Wild) Staner (Menispermaceae) with Antimycobacterial Activity against *Mycobacterium Tuberculosis*. *Natural product research*, 33(5), 642-650.
- Tiwari, R., & Rana, C. (2015). Plant Secondary Metabolites: A Review. *International Journal of Engineering Research and General Science*, 3(5), 661-670.

- Tosun, F., Akyüz Kızılay, Ç., Şener, B., & Vural, M. (2005). The Evaluation of Plants from Turkey for in Vitro Antimycobacterial Activity. *Pharmaceutical biology*, 43(1), 58-63.
- Trumpower, B. L., & Gennis, R. B. (1994). Energy Transduction by Cytochrome Complexes in Mitochondrial and Bacterial Respiration: The Enzymology of Coupling Electron Transfer Reactions to Transmembrane Proton Translocation. *Annual review of biochemistry*, 63(1), 675-716.
- Udofia, P. G., Ojimelukwe, P. C., Olaoye, O. A., Ukom, A. N., Ekanem, M. L., & Okparauka, I. I. (2022). Evaluation of Antioxidant Activity of Ethanol Extract of Root and Stem Bark of Moringa Oleifera (Mo) Obtained from Utu Ikpe, Ikot Ekpene Local Government Area, Nigeria. *Communication in Physical Sciences*, 8(1).
- Umesiri, F. E., Lick, A., Fricke, C., Funk, J., & Nathaniel, T. (2015). Anti-Tubercular Activity of Edta and Household Chemicals against Mycobacterium Smegmatis, a Surrogate for Multi-Drug Resistant Tuberculosis. *European Scientific Journal, ESJ*, 11(21).
- Urdahl, K., Shafiani, S., & Ernst, J. (2011). Initiation and Regulation of T-Cell Responses in Tuberculosis. *Mucosal immunology*, 4(3), 288.
- Usman, H., Majiya, H., Aremu, K. H., Legbo, M., & Musa, A. (2022). Evaluation of Phytochemical Analysis and Antibacterial Potential of Different Solvent Extracts of Leaves and Stem Bark of Eucalyptus Camaldulensis against Selected Pathogenic Bacteria. *Biokemistri*, 33(4).
- Ustinova, V. V., Smirnova, T. G., Sochivko, D. G., Varlamov, D. A., Larionova, E. E., Andreevskaya, S. N., Andrievskaya, I. Y., Kiseleva, E. A., Chernoussova, L. N., & Ergeshov, A. (2019). New Assay to Diagnose and Differentiate between Mycobacterium Tuberculosis Complex and Nontuberculous Mycobacteria. *Tuberculosis*, 114, 17-23.
- Van Puyvelde, L., Ntawukiliyayo, J., Portaels, F., & Hakizamungu, E. (1994). In Vitro Inhibition of Mycobacteria by Rwandese Medicinal Plants. *Phytotherapy Research*, 8(2), 65-69.
- Veggi, P. C., Martinez, J., & Meireles, M. A. A. (2012). Fundamentals of Microwave Extraction. In *Microwave-Assisted Extraction for Bioactive Compounds* (pp. 15-52): Springer.

- Vinatoru, M., Mason, T., & Calinescu, I. (2017). Ultrasonically Assisted Extraction (Uae) and Microwave Assisted Extraction (Mae) of Functional Compounds from Plant Materials. *TrAC Trends in Analytical Chemistry*, 97, 159-178.
- Vriet, C., Russinova, E., & Reuzeau, C. (2012). Boosting Crop Yields with Plant Steroids. *The Plant Cell*, 24(3), 842-857.
- Vukovic, D. S., & Nagorni-Obradovic, L. M. (2011). Knowledge and Awareness of Tuberculosis among Roma Population in Belgrade: A Qualitative Study. *BMC infectious diseases*, 11(1), 1-6.
- Waleed A, M., Samah S, A., Mona F, S., Abeer H, E., Hossam M, H., Elham, A., & Mona H, H. (2019). Immunomodulatory Effect of Premna Odorata Volatile Oils in Mycobacterium Tuberculosis by Inhibiting Tlr4/Nf-Kb Pathway. *Journal of Herbmed Pharmacology*, 8(1).
- Wang, G., Tang, W., & Bidigare, R. R. (2005). Terpenoids as Therapeutic Drugs and Pharmaceutical Agents. In *Natural Products* (pp. 197-227): Springer.
- Wang, T., Jiao, J., Gai, Q.-Y., Wang, P., Guo, N., Niu, L.-L., & Fu, Y.-J. (2017). Enhanced and Green Extraction Polyphenols and Furanocoumarins from Fig (*Ficus Carica L.*) Leaves Using Deep Eutectic Solvents. *Journal of pharmaceutical and biomedical analysis*, 145, 339-345.
- Wang, Y.-Y., Peng, C., Zhang, Y., Wang, Z.-R., Chen, Y.-M., Dong, J.-F., Xiao, M.-L., Li, D.-L., Li, W., & Zou, Q.-J. (2022). Optimization, Identification and Bioactivity of Flavonoids Extracted from *Moringa Oleifera* Leaves by Deep Eutectic Solvent. *Food Bioscience*, 101687.
- Warner, R., Wu, B.-S., MacPherson, S., & Lefsrud, M. (2021). A Review of Strawberry Photobiology and Fruit Flavonoids in Controlled Environments. *Frontiers in Plant Science*, 12.
- Wink, M. (2012). Medicinal Plants: A Source of Anti-Parasitic Secondary Metabolites. *Molecules*, 17(11), 12771-12791.
- Wolf, A. J., Desvignes, L., Linas, B., Banaiee, N., Tamura, T., Takatsu, K., & Ernst, J. D. (2008). Initiation of the Adaptive Immune Response to Mycobacterium Tuberculosis Depends on Antigen Production in the Local Lymph Node, Not the Lungs. *Journal of Experimental Medicine*, 205(1), 105-115.
- Wu, H., Guo, J., Chen, S., Liu, X., Zhou, Y., Zhang, X., & Xu, X. (2013). Recent Developments in Qualitative and Quantitative Analysis of Phytochemical Constituents and Their Metabolites Using Liquid Chromatography–Mass

- Spectrometry. *Journal of pharmaceutical and biomedical analysis*, 72, 267-291.
- Wu, L., Li, L., Chen, S., Wang, L., & Lin, X. (2020). Deep Eutectic Solvent-Based Ultrasonic-Assisted Extraction of Phenolic Compounds from Moringa Oleifera L. Leaves: Optimization, Comparison and Antioxidant Activity. *Separation and Purification Technology*, 247, 117014.
- Xia, G.-H., Li, X.-H., & Jiang, Y.-h. (2021). Deep Eutectic Solvents as Green Media for Flavonoids Extraction from the Rhizomes of Polygonatum Odoratum. *Alexandria Engineering Journal*, 60(2), 1991-2000.
- Xu, W., DeJesus, M. A., Rücker, N., Engelhart, C. A., Wright, M. G., Healy, C., Lin, K., Wang, R., Park, S. W., & Ioerger, T. R. (2017). Chemical Genetic Interaction Profiling Reveals Determinants of Intrinsic Antibiotic Resistance in Mycobacterium Tuberculosis. *Antimicrobial agents and chemotherapy*, 61(12), e01334-01317.
- Yadav, S., Mathur, M., & Dixit, A. (2006). Knowledge and Attitude Towards Tuberculosis Amongsandstone Quarry Workers in Desert Parts of Rajasthan.
- Yagi, A., Uchida, R., Hamamoto, H., Sekimizu, K., Kimura, K.-i., & Tomoda, H. (2017). Anti-Mycobacterium Activity of Microbial Peptides in a Silkworm Infection Model with Mycobacterium Smegmatis. *The Journal of antibiotics*, 70(5), 685-690.
- Yalavarthi, C., & Thiruvengadarajan, V. (2013). A Review on Identification Strategy of Phyto Constituents Present in Herbal Plants. *Int. J. Res. Pharma. Sci*, 4(2), 123-140.
- Yao, C., Li, X., Bi, W., & Jiang, C. (2014). Relationship between Membrane Damage, Leakage of Intracellular Compounds, and Inactivation of Escherichia Coli Treated by Pressurized Co<sub>2</sub>. *Journal of basic microbiology*, 54(8), 858-865.
- Youssef, F. S., Ovidi, E., Musayeib, N. M. A., & Ashour, M. L. (2021). Morphology, Anatomy and Secondary Metabolites Investigations of Premna Odorata Blanco and Evaluation of Its Anti-Tuberculosis Activity Using in Vitro and in Silico Studies. *Plants*, 10(9), 1953.
- Yu, Y., Yao, C., & Guo, D.-a. (2021). Insight into Chemical Basis of Traditional Chinese Medicine Based on the State-of-the-Art Techniques of Liquid Chromatography– Mass Spectrometry. *Acta Pharmaceutica Sinica B*, 11(6), 1469-1492.

- Zakaria, F., Ibrahim, W. N. W., Ismail, I. S., Ahmad, H., Manshoor, N., Ismail, N., Zainal, Z., & Shaari, K. (2019). Lcms/Ms Metabolite Profiling and Analysis of Acute Toxicity Effect of the Ethanolic Extract of Centella Asiatica on Zebrafish Model. *Pertanika Journal of Science & Technology*, 27(2).
- Zakawa, N., Timon, D., Yusuf, C., Oyebanji, E., Batta, K., & Jalani, R. (2020). Ethno-Botanical Survey and Phytochemical Analysis of Moringa Oleifera in Mubi Local Government of Adamawa State. *Journal of Medicinal Plants Studies*, 8(2), 107-111.
- Zhang, D.-Y., Zu, Y.-G., Fu, Y.-J., Luo, M., Wang, W., Gu, C.-B., Zhao, C.-J., Jiao, J., & Efferth, T. (2012). Enzyme Pretreatment and Negative Pressure Cavitation Extraction of Genistein and Apigenin from the Roots of Pigeon Pea [Cajanus Cajan (L.) Millsp.] and the Evaluation of Antioxidant Activity. *Industrial Crops and Products*, 37(1), 311-320.
- Zhang, L., & Wang, M. (2017). Optimization of Deep Eutectic Solvent-Based Ultrasound-Assisted Extraction of Polysaccharides from Dioscorea Opposita Thunb. *International journal of biological macromolecules*, 95, 675-681.
- Zhang, Q., Vigier, K. D. O., Royer, S., & Jerome, F. (2012). Deep Eutectic Solvents: Syntheses, Properties and Applications. *Chemical Society Reviews*, 41(21), 7108-7146.
- Zhao, B.-Y., Xu, P., Yang, F.-X., Wu, H., Zong, M.-H., & Lou, W.-Y. (2015). Biocompatible Deep Eutectic Solvents Based on Choline Chloride: Characterization and Application to the Extraction of Rutin from Sophora Japonica. *ACS Sustainable Chemistry & Engineering*, 3(11), 2746-2755.
- Zhao, J., Evangelopoulos, D., Bhakta, S., Gray, A. I., & Seidel, V. (2014). Antitubercular Activity of Arctium Lappa and Tussilago Farfara Extracts and Constituents. *Journal of Ethnopharmacology*, 155(1), 796-800.
- Zhou, K., Zhou, W., Li, P., Liu, G., Zhang, J., & Dai, Y. (2008). Mode of Action of Pentocin 31-1: An Antilisteria Bacteriocin Produced by Lactobacillus Pentosus from Chinese Traditional Ham. *Food Control*, 19(8), 817-822.
- Zhuang, B., Dou, L.-L., Li, P., & Liu, E.-H. (2017). Deep Eutectic Solvents as Green Media for Extraction of Flavonoid Glycosides and Aglycones from Platycladi Cacumen. *Journal of pharmaceutical and biomedical analysis*, 134, 214-219.
- Ziegler, J., & Facchini, P. J. (2008). Alkaloid Biosynthesis: Metabolism and Trafficking. *Annu. Rev. Plant Biol.*, 59, 735-769.

- Zumla, A., Nahid, P., & Cole, S. T. (2013). Advances in the Development of New Tuberculosis Drugs and Treatment Regimens. *Nature reviews Drug discovery*, 12(5), 388.
- Zumla, A. I., Gillespie, S. H., Hoelscher, M., Philips, P. P., Cole, S. T., Abubakar, I., McHugh, T. D., Schito, M., Maeurer, M., & Nunn, A. J. (2014). New Antituberculosis Drugs, Regimens, and Adjunct Therapies: Needs, Advances, and Future Prospects. *The Lancet infectious diseases*, 14(4), 327-340.
- Zwenger, S., & Basu, C. (2008). Plant Terpenoids: Applications and Future Potentials. *Biotechnology and Molecular Biology Reviews*, 3(1), 1.



PTTA UTHM  
PERPUSTAKAAN TUNKU TUN AMINAH

## **VITA**

The author was born in July 31, 1986, in Alanbar, Iraq. He attended Primary school at Al-muhag, Baghdad capital city, from 1992 to 1998. He later attended Government Secondary School Al-adamea, Baghdad capital city from 1998 to 2004. He pursued his degree at the Al-mousal University, Mousal, Iraq , and graduated with the B.V.S.M (Hons) in veterinary medicine science in 2009. The author was attended in 2010/2011 session to obatain Masters of Science Degree in 2013. The author enrolled in a PhD in Science program in 2018/2019 session. The author has been married to Sanna Qasim since 2013, and is blessed with one son (Qaeath).



PTTA  
PERPUSTAKAAN TUNKU TUN AMINAH