

SYNTHESIS AND CHARACTERIZATION OF CURCUMIN-CHITOSAN
LOADED GOLD NANOPARTICLES BY *Oryctes rhinoceros*' CHITIN FOR
COSMECEUTICAL APPLICATION

NURUL ALYANI BINTI ZAINOL ABIDIN

A thesis submitted in
fulfillment of the requirement for the award of the
Doctor of Philosophy in Science



Faculty of Applied Sciences and Technology
Universiti Tun Hussein Onn Malaysia

DECEMBER 2022


PROF MADYA TS DR FARIDAH BT KORMIN
Jabatan Teknologi dan Sumber Semulajadi
Fakulti Sains Gunaaan dan Teknologi
Universiti Tun Hussein Onn Malaysia (UTHM)

DEDICATION

"I am only one, but still, I am one. I cannot do everything, but still, I can do something and because I cannot do everything, I will not refuse to do the something that I can do."

I dedicate this thesis to my family for their endless support, affection, belief, and love. I would also like to dedicate this thesis to my supervisor for her guidance, support, and letting me do the things I am most comfortable with. To my friends, who have always brought sunshine during the darkest of times. They bring joy and comfort during those stressful moments. And the ultimate supporter, as a thousand times I have failed, but still your mercy remains. Thank you, Allah S.W.T

ACKNOWLEDGEMENT

My family will always be my top priority, as I am forever theirs. Without the moral, financial, and continuous support, and encouragement from them, it is not possible for me to be where I am today. I would like to express my gratitude to my biggest supporters, my mother, Nor Halizatun Binti Shamsuddin, and my father, Zainol Abidin Bin Ibrahim. There is nothing that I want more than to give them my best, and I will do my hardest to be better for my future.

I wish to extend my sincere appreciation to my supervisor, Assoc. Prof. Ts. Dr. Faridah Binti Kormin and co-supervisor, Assoc. Prof. Ts. Dr. Mohd Fadzelly Bin Abu Bakar, for the endless patience, guidance, and support. With such a strong support system, I am confident in every decision I make. We have gone through thick and thin together, and even though I have never expressed my thanks to you, I will always and will eternally be grateful for everything you have done for me. I will forever be indebted to your act of kindness.

Finally, I'd like to thank all of my friends and companions who, both directly and indirectly, made things easier simply by being present and listening to all of my problems. Even the distractions are gold since they give me breaks here and there to refresh my mind. I have benefited greatly from their sun and joy.

In addition, many thanks to all the lecturers and UTHM staff for their guidance and support, especially to the laboratory technicians, for always being kind, helpful, and supportive during my lab work.

ABSTRACT

A breakthrough in cosmeceuticals by utilizing insects as major ingredients in cosmetic products is gaining popularity. Therefore, the interest in rare sources of ingredients, for instance, from the *Oryctes rhinoceros* beetle, can bring huge benefit in turning pest to wealth. In this study, curcumin was chosen as the active ingredient loaded into chitosan-gold nanoparticles (CCG-NP). However, curcumin is unstable, and has high sensitivity to light thus curcumin has low bioavailability. Therefore, chitosan extracted from *O. rhinoceros* acts as a drug carrier to overcome these problems. In order to overcome the problems, several objectives were achieved, including optimizing the parameters of chitin extraction, deacetylation of chitosan, followed by characterization of *O. rhinoceros*' chitin and chitosan by their physical properties and spectrophotometer evaluation, to name a few. CCG-NPs are synthesized and characterized, and their efficacy is determined by their drug release profile, antimicrobial and anti-tyrosinase properties, and toxicity test. The best RSM chitin extraction parameters were 1 M NaOH, 99 °C, 20 ml/g of sample for deproteinization and 3.92 M HCl, 75 °C, 12 ml/g of sample for decalcification. Chitin and chitosan from *O. rhinoceros* are comparable to commercial chitin and chitosan. CCG-NPs are successfully synthesized at 70 °C for 60 minutes under optimal conditions of reactant ratio of 2:0.5 (0.5 mM HAuCl₄: 0.1 % curcumin). The shape of the CCG-NP is determined via FE-SEM, and it is round, and the size is 128.27 d.nm. The value of the zeta potential is 20.2 ± 3.81 mV showing a stable mixture. An *in vitro* drug release profile shows a cumulative drug release in 24 hours of 52.99 %. The zone diameter for inhibition for *E. coli* and *S. aureus* was 7.0 mm and 8.5 mm, respectively, with tyrosinase enzyme inhibition of 66.385 ± 3.0%. The cytotoxicity of cell viability IC₅₀ is 58% of the CCG-NP concentration indicating a mild toxicity trait. To conclude, CCG-NP is a stable, spherical, nano-sized, homogeneous solution with a good drug release profile, antimicrobial properties, enhanced anti-tyrosinase activity, and is non-toxic for the cosmeceutical applications.

ABSTRAK

Penemuan baharu dalam kosmeseutikal dengan menggunakan serangga sebagai bahan utama dalam produk kosmetik menjadi semakin popular. Oleh itu, perhatian terhadap sumber bahan yang jarang ditemui, contohnya, daripada kumbang tanduk, *Oryctes rhinoceros*, boleh membawa manfaat yang besar dalam mengubah serangga perosak kepada sesuatu yang bernilai. Dalam kajian ini, kurkumin dipilih sebagai bahan aktif yang digabungkan bersama kitosan-nanozarah emas (CCG-NP). Walau bagaimanapun, kurkumin tidak stabil, serta mudah terurai justeru menyumbang kepada bioavailabiliti yang rendah. Oleh itu, kitosan yang diekstrak daripada *O. rhinoceros* bertindak sebagai ‘*drug carrier*’ untuk mengatasi masalah tersebut. Bagi mengatasi masalah tersebut, beberapa objektif telah dicapai, termasuk mengoptimumkan parameter pengekstrakan kitin, penyahetilasi kitosan, diikuti dengan pencirian kitin dan kitosan *O. Rhinoceros*. CCG-NPs disintesis dan dicirikan, dan keberkesanannya ditentukan oleh profil pelepasan ubat, sifat antimikrob dan anti-tirosinase, serta ujian ketoksikan. Parameter pengekstrakan kitin RSM terbaik ialah 1 M NaOH, 99 °C, 20 ml/g sampel untuk penyahproteinan dan 3.92 M HCl, 75 °C, 12 ml/g sampel untuk penyahkalsifikasian. Kitin dan kitosan daripada *O. rhinoceros* adalah setanding dengan kitin dan kitosan komersial. CCG-NPs berjaya disintesis pada 70 °C selama 60 minit di bawah keadaan optimum nisbah 2:0.5 (0.5 mM HAuCl₄: 0.1% curcumin). Melalui FE-SEM, bentuk CCG-NP adalah sfera dengan saiz 128.27 d.nm . Nilai potensi zeta ialah 20.2 ± 3.81 mV menunjukkan sebatian yang stabil. Profil pelepasan ubat *in vitro* menunjukkan pelepasan ubat kumulatif dalam 24 jam sebanyak 52.99 %. Antimikrob untuk *E. coli* dan *S. aureus* ialah 7.0 mm dan 8.5 mm, masing-masing, dengan perencatan enzim tyrosinase sebanyak $66.385 \pm 3.0\%$. Sitotoksisiti IC₅₀ adalah sederhana dengan hanya 58 %. Sebagai kesimpulan, CCG-NP ialah larutan homogen yang stabil, berbentuk sfera, bersaiz nano, dengan profil pelepasan ubat yang baik, sifat antimikrob, aktiviti anti-tirosinase yang lebih efektif, dan tidak bertoksik untuk kegunaan kosmetik.

TABLE OF CONTENTS

TITLE	i
DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	xiii
LIST OF FIGURES	xvi
LIST OF SYMBOLS AND ABBREVIATIONS	xxi
LIST OF APPENDICES	xxiii
LIST OF PUBLICATIONS	xxiv
LIST OF AWARDS	xxv
CHAPTER 1 INTRODUCTION	1
1.1 Background of study	1
1.2 Problem statement	4
1.3 Research objectives	6
1.4 Scope of study	7
1.4.1 Sample collection and procedure of extraction	7
1.4.2 Characterization of chitin and chitosan	7
1.4.3 Optimization of CCG-NP synthesis	8
1.4.4 Characterization of CCG-NP	8
1.4.5 <i>In vitro</i> study on drug release, antimicrobial and anti-tyrosinase properties	8
1.4.6 Cytotoxicity study via MTT assay	8

1.5	Significance of study	9
CHAPTER 2	LITERATURE REVIEW	10
2.1	Introduction	10
2.2	Nanoparticles in cosmeceutical drug delivery system	10
2.2.1	Properties of nanoparticles	12
2.2.2	Applications of nanoparticles in cosmeceutical	14
2.2.3	Advantages of nanoparticle in cosmeceutical industry	16
2.3	Chitin and chitosan as biocarrier of active ingredients in nanotechnology and cosmeceutical applications	16
2.3.1	Chemical extraction of chitin	19
2.3.2	Chitin's chemical extraction from insects	23
2.3.3	Chitosan: a derivative of chitin	31
2.3.4	Optimisation of chitin extraction from insects via Response Surface Methodology (RSM), Design Expert	32
2.4	Curcumin as bioactive ingredients for CCG-NP	33
2.4.1	Therapeutics properties of curcumin	35
2.5	<i>Oryctes rhinoceros</i> as the source of chitin and chitosan	36
2.5.1	Insect classification	36
2.5.2	Biogeography of <i>O. rhinoceros</i>	37
2.5.3	Anatomy and physiology of <i>O. rhinoceros</i>	37
2.5.4	<i>O. rhinoceros'</i> behaviour	38
2.6	Characterisation of chitin and chitosan	40
2.6.1	Physicochemical characteristics and properties of chitin and chitosan	40
2.6.2	Degree of deacetylation (DD)	41
2.6.3	Solubility properties	42

2.6.4	Fourier-transform infrared (FTIR) spectroscopy	42
2.6.5	Thermal analysis	44
2.6.6	Crystallinity properties by using X-ray Diffraction (XRD)	44
2.7	Preparation of curcumin chitosan-loaded nanoparticle (CCG-NP)	46
2.7.1	Optimisation and characterisation of CCG-NP	47
2.7.2	<i>In vitro</i> tests in cosmeceutical application; drug release, antimicrobial and anti-tyrosinase properties	53
2.8	Toxicity test of CCG-NP via MTT assay of 3T3 cells	58
2.9	Data analysis	59
2.10	Conclusion	59
CHAPTER 3	METHODOLOGY	61
3.1	Introduction	61
3.1.1	General flow of the methodology	63
3.2	Reagents and solvents	63
3.3	Apparatus and instruments	63
3.4	Sample collection	64
3.4.1	Sample preparation	64
3.5	Optimisation of chitin extraction by RSM	66
3.5.1	Deproteinization by strong base	67
3.5.2	Protein quantification by biuret method	67
3.5.3	Decalcification by strong acid	67
3.5.4	Calcium quantification by spectrophometry method	68
3.5.5	Kinetics of reaction for deproteinization and decalcification	68
3.6	Deacetylation of chitosan	69
3.7	Characterisation of chitin and chitosan	69

3.7.1	Physical characteristics and properties of chitin and chitosan	69
3.7.2	Degree of deacetylation: Titration method	70
3.7.3	Solubility characteristics	71
3.7.4	Morphology evaluation by using FE-SEM	72
3.7.5	Fourier-transform infrared (FTIR) spectroscopy	72
3.7.6	Thermal analysis	73
3.7.7	X-ray diffraction (XRD)	74
3.8	Preparation of CCG-NP by UV-vis surface plasmon resonance (SPR) band method	75
3.8.1	Optimization of reactant ratio for CCG-NP synthesis	75
3.8.2	Optimization of reactant time for CCG-NP synthesis	77
3.9	Characterisation of CCG-NP	77
3.9.1	Measurement of zeta potential	78
3.9.2	Morphology evaluation of CCG-NP via FE-SEM	78
3.9.3	Particle size distribution analysis by zetasizer	78
3.9.4	Energy dispersive X-ray spectroscopy (EDX)	78
3.9.5	<i>In vitro</i> tests in cosmeceutical application; drug release, antimicrobial and anti-tyrosinase properties	79
3.9.6	Toxicity test by MTT assay	81
3.10	Data analysis	82
CHAPTER 4	DISCUSSION	83
4.1	Optimisation of chitin extraction by RSM	83
4.1.1	Optimising the deproteinization of insect chitin by RSM	84
4.1.2	Kinetics of deproteinization: Rate of	89

	reaction	
4.1.3	Optimising the decalcification of insect chitin by RSM	92
4.1.4	Kinetics of decalcification: Rate of reaction	97
4.1.5	Order of chitin extraction	100
4.1.6	Relationship between chitin and chitosan extraction with cosmeceutical applications	102
4.2	Characterisation of chitin and chitosan	103
4.2.1	Physical characteristics and properties of chitin and chitosan	103
4.2.2	Degree of deacetylation (DD) of chitin and chitosan	107
4.2.3	Solubility characteristic of chitin and chitosan	109
4.2.4	Morphological properties of chitin and chitosan	110
4.2.5	Fourier-transform infrared spectroscopy	113
4.2.6	Thermal methods of analysis	118
4.2.7	Determining crystallinity index via XRD	128
4.3	Optimisation of CCG-NP by UV-vis surface plasmon resonance (SPR) band method	132
4.3.1	Preparation of CCG-NP	132
4.4	Characterisation of CCG-NP	144
4.4.1	Fourier transform infrared (FTIR) spectroscopy	144
4.4.2	Measurement of zeta potential	146
4.4.3	Morphological properties of CCG-NP by using FE-SEM	150
4.4.4	Particle size distribution analysis of CCG-NP	150
4.4.5	Determining crystallinity nature of CCG-	153

NP via XRD	
4.4.6 Elemental analysis via EDX	154
4.4.7 <i>In vitro</i> tests in cosmeceutical applications; drug release, antimicrobial and anti-tyrosinase properties	156
4.4.8 Toxicity test by MTT assay	167
4.5 Significance or the applied value of this research in cosmeceutical application	170
CHAPTER 5 CONCLUSION	171
5.1 Conclusion	171
5.2 Recommendation	172
REFERENCES	174
APPENDIX	203
VITA	213



PTTA AUTHN
PERPUSTAKAAN TUNKU TUN AMINAH

LIST OF TABLES

2.1	List of marketed products of gold nanoparticles by market research	15
2.2	A summary of demineralization and deproteinization of chitin and chitosan from insects.	29
2.3	<i>O. rhinoceros</i> beetle's classification (Bedford, 1980)	36
3.1	Composition for CCG-NP reaction mixtures and conditions from <i>O. rhinoceros</i> chitosan	77
3.2	Composition for CCG-NP reaction mixtures and conditions from commercial crustacean chitosan	77
3.3	Composition of enzyme assay without the sample	81
3.4	Composition of CCG-NP reaction mixtures	81
4.1	Response surface methodology of deproteinization (the actual design of experiments and response)	85
4.2	ANOVA table of analysis of residual protein content after 6 hours and the deproteinization variables for quadratic model	87
4.3	Optimization solution for the deproteinization	89
4.4	One-way ANOVA for optimisation of deproteinization solution	89
4.5	One sample <i>T</i> -test for validation	89
4.6	Variation of the rate constant (<i>k</i>)	92
4.7	Response surface methodology of decalcification (the actual design of experiments and response)	93
4.8	ANOVA table for final calcium concentration after 6 hours and the decalcification variables for quadratic model	95
4.9	Optimisation solution for decalcification	96

4.10	One-way ANOVA for optimisation solution of decalcification	97
4.11	One sample <i>T</i> -test for validation	97
4.12	Variation of the rate constant (<i>k</i>) for decalcification of beetle chitin	99
4.13	Summary for one sample <i>t</i> -test for determining the most preferable order of deproteinization and decalcification	102
4.14	Yield percentage of chitin and chitosan from <i>O. rhinoceros</i> beetle	105
4.15	Moisture content of chitin and chitosan from <i>O. rhinoceros</i> beetle and commercial	106
4.16	Water activity of chitin and chitosan from <i>O. rhinoceros</i> beetle and commercial	107
4.17	Degree of deacetylation of chitin and chitosan from <i>O. rhinoceros</i> beetle and commercial	109
4.18	Solubility value of chitin and chitosan from <i>O. rhinoceros</i> beetle and commercial	110
4.19	The FTIR bands (cm^{-1}) of chitin from <i>O. rhinoceros</i> , commercial chitin (from this study) and commercial shrimp chitin (Ibitoye, <i>et al.</i> , 2018)	116
4.20	The FTIR bands (cm^{-1}) of chitosan from <i>O. rhinoceros</i> , commercial chitosan (from this study) and commercial shrimp chitosan (Ibitoye, <i>et al.</i> , 2018).	118
4.21	Thermal stability data of <i>O. rhinoceros</i> and commercial chitin and chitosan	126
4.22	Crystallinity index of <i>O. rhinoceros</i> and commercial chitin and chitosan from XRD	130
4.23	Effect of curcumin concentration on CCG-NP synthesis in 24 hours at room temperature for chitosan from <i>O. rhinoceros</i>	134
4.24	Effect of curcumin concentration on CCG-NP synthesis in 24 hours at room temperature for chitosan	137

	from commercial crustacean	
4.25	Reaction mixture for the optimization of reactant time	141
4.26	The wavelength λ , (nm) and absorbance for each peak of C2, C10 and C16 (control) at minutes 5 to 60	141
4.27	Zeta potential of as-synthesised CCG-NP; mean \pm sd (n=3)	148
4.28	Size distribution of CCG-NP according to particle size	152
4.29	EDX profile of elemental composition of CCG-NP	156
4.30	Images of zone diameter inhibition for sample C2, C10, C16 and 0.1 % curcumin in methanol and streptomycin as positive control against <i>E. coli</i> and <i>S.</i> <i>aureus</i>	163
4.31	<i>In vivo</i> antimicrobial study of CCG-NP by zone diameter inhibition (mm) through disc diffusion method	164
4.32	Anti-tyrosinase enzyme assay value where C16 acts as negative control and kojic acid as positive control	167
4.33	Optical density value and percentage of cell viability in various concentration of CCG-NP exposed to Caco2 cells	169

LIST OF FIGURES

2.1	Colours of various sized monodispersed gold nanoparticles (Canovi, <i>et al.</i> , 2012)	13
2.2	General process for chitin and chitosan extraction (Zainol Abidin, <i>et al.</i> , 2020)	22
2.3	Deacetylation of chitin to chitosan (Benjakul & Prodpan, 2019)	31
2.4	Chemical structure of curcumin (Aggarwal, Kumar & Bharti, 2003)	33
2.5	Anatomy and physiology of adult (left) and left (middle and right) of different stages in life for <i>O. rhinoceros</i> (Bedford, 1980)	38
2.6	The cycle of one life span for <i>O. rhinoceros</i> from eggs to adult (Bedford, 1980)	40
2.7	FTIR Spectra of (a) chitosan, (b) chitosan-gold nanoparticle and (c) curcumin loaded chitosan-gold nanoparticle (Amanlou, <i>et al.</i> , 2019)	43
2.8	XRD pattern of curcumin, curcumin nanoparticle and chitosan nanoparticle (Nair, <i>et al.</i> , 2019)	46
2.9	Gold nanoparticle size dependent surface plasmon resonance. The red-shift of the absorption maximises as the gold nanoparticle size increases (Shafiqa, Abdul Aziz, & Mehrdel, 2018).	48
2.10	SPR band of (a) pure gold nanoparticle and (b) nanoparticles conjugated with bioactive compounds (Wojnarowska-Nowak, <i>et al.</i> , 2015)	48
2.11	UV-vis spectrum of gold nanoparticles, (a) before and (b) after loaded with bioactive compounds (Khurana, <i>et al.</i> , 2015)	49

2012)	
2.12	FE-SEM image of turmeric crude extract gold nanoparticle at 100, 000 magnification (Latif, <i>et al.</i> , 2020) 51
2.13	EDX and elemental mapping of <i>D. morbifera</i> gold nanoparticles (Wang, <i>et al.</i> , 2016) 53
2.14	Illustration of dialysis bag technique (Lee & Koros, 2003) 55
2.15	Antimicrobial activity in disc diffusion technique for antibiotics' inhibition zone (Wheat, <i>et al.</i> , 2001) 56
2.16	Mechanism of reaction for tyrosinase inhibition; E = enzyme, S = substrate, I = inhibitor, P = product, ES = enzyme-substrate, EI = enzyme-inhibitor, and ESI = enzyme-substrate-inhibitor (Chang, 2009) 59
3.1	General flow chart of research activities 62
3.2	Images of the (a) sample, (b) pheromone trap used for pest control and (c) sample collection in Felda Maokil, Johor, Malaysia. 65
3.3	Images of sample preparation of <i>O. rhinoceros</i> from dried sample to powdered sample 65
4.1	Summary of results from RSM 83
4.2	Surface plot diagram for the parameters. 87
4.3	Kinetics of deproteinization in 1 M NaOH, 99 °C, 20 ml/g of sample (solution to solute ratio) for 360 minutes 91
4.4	Logarithmic variation of the protein content in chitin as a function of the deproteinization time in 1 M NaOH, 99 °C, 20 ml/g of sample (solution to solute ratio) for 360 minutes 91
4.5	Surface plot diagram for the parameters 96
4.6	Kinetics of decalcification in 3.92 M HCl, 75 °C and 12 ml/g of sample (solution to solute ratio) for 360 minutes 99
4.7	Logarithmic variation of the protein content in chitin as a function of the decalcification time in 3.92 M HCl, 75 °C and 12 ml/g of sample (solution to solute ratio) for 360 99

	minutes	
4.8	The flow charts showed the flow of experiments for when (a) deproteinization was done prior to decalcification and (b) decalcification was done prior to deproteinization	100
4.9	The images shown powdered <i>O. rhinoceros</i> (a) without chemical treatment, (b) after first treatment with HCl, (c) with second treatment using NaOH after being treated by HCl, (d) after first treatment with NaOH, and (e) with second treatment using HCl after being treated by NaOH	101
4.10	FE-SEM images showing the surface morphology of (a) chitin isolated from <i>O. rhinoceros</i> , (b) commercial chitin, (c) chitosan isolated from <i>O. rhinoceros</i> and (d) commercial chitosan at 35, 000 magnification	111
4.11	Images showing the powdered form of (a) commercial chitin, (b) <i>O. rhinoceros</i> chitin, (c) commercial chitosan, and (d) <i>O. rhinoceros</i> chitosan taken using standard digital camera	112
4.12	The IR Spectra of the samples from (a) commercial chitin and (b) <i>O. rhinoceros</i> chitin	115
4.13	Chemical structure of chitin	115
4.14	The IR Spectra of the samples from (a) commercial chitosan and (b) <i>O. rhinoceros</i> chitosan	117
4.15	Chemical structure of chitosan	117
4.16	DSC diffractogram for (a) <i>O. rhinoceros</i> chitin, (b) commercial chitin, (c) <i>O. rhinoceros</i> chitosan and (d) commercial chitosan	121
4.17	DSC diffractogram glass transition for (a) <i>O. rhinoceros</i> chitin, (b) commercial chitin, (c) <i>O. rhinoceros</i> chitosan and (d) commercial chitosan	123
4.18	Thermogravimetric analysis (TGA) of (a) <i>O. rhinoceros</i> chitin, (b) commercial chitin, (c) <i>O. rhinoceros</i> chitosan and (d) commercial chitosan	127
4.19	The image shown is an XRD spectra of (a) chitin isolated	131

from <i>O. rhinoceros</i> and (b) commercial chitin where the is 00-036-1523, N-acetyl-d-glucosamine ($C_8H_{15}NO_6$)	
4.20 The image shown is an XRD spectra of chitosan isolated from (a) <i>O. rhinoceros</i> chitosan and (b) commercial where the pattern is 00-054-1953, chitosan d-ascorbate ($C_{12}H_{21}NO_{11}$)n	131
4.21 The images show the changes in colour after 24 hours of reaction for CCG-NP using <i>O. rhinoceros</i> chitosan in room temperature	134
4.22 UV-vis spectra SPR band for C1-C7 for the formulation of CCG-NP with <i>O. rhinoceros</i> chitosan	135
4.23 The images show the changes in colour after 24 hours of reaction for CCG-NP using commercial chitosan in room temperature	137
4.24 UV-vis spectra SPR band for C8-C14 for the formulation of CCG-NP with commercial chitosan	138
4.25 The images show the changes in colour after their respective optimised reaction time with C16 as control	140
4.26 UV-vis spectra SPR band for (a) C2 (b) C10 and (c) C16 at minute 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55 and 60	142
4.27 UV-vis spectra SPR band for C2, C10 and C16 at 60 minutes of reaction time	143
4.28 UV–vis absorption spectra and absorption shift of (a) gold nanoparticles (Lopez Munoz, et al., 2012) and (b) silver nanoparticle (Belteky, et al., 2019)	143
4.29 FTIR spectra of C2, C10, curcumin, gold (III) chloride, TPP, <i>O. rhinoceros</i> chitosan and commercial chitosan	145
4.30 Zeta potential for (a) C2, (b) C10 and (c) C16	148
4.31 FE-SEM images showing the spherical morphology of CCG-NP (a) C2, (b) C10 and (c) C16 at 80 000 x magnification	150
4.32 Size distribution by intensity (a) C2, (b) C10, (c) C16 and (d) 0.1 % curcumin in methanol	152

4.33	The images shown is an XRD spectra of C2, C10 and C16 in their amorphous state	154
4.34	EDX spectra of CCG-NP (a) C2, (b) C10, (c) C16	156
4.35	The cumulative drug release kinetics of curcumin for (a) pH 5.3 and (b) pH 7.4 in a period of 1440 minutes	159
4.36	Standard curve of curcumin	160
4.37	The graph showed an absorbance value of anti-tyrosinase enzyme assay for 60 mins for 2500, 1000, 750, 500 and 250 units/ml of tyrosinase enzyme	166
4.38	The graph showed the inhibition % of anti-tyrosinase enzyme assay where kojic acid as positive control and C16 as negative control	167
4.39	Effect of CCG-NP on 3T3 cells after 72 hours of exposure	170



PTT AUTHN
PERPUSTAKAAN TUNKU TUN AMINAH

LIST OF SYMBOLS AND ABBREVIATIONS

$\%$	-	Percentage
$^{\circ}\text{C}$	-	Degree celsius
<i>13CP/MAS-NMR</i>	-	^{13}C cross-polarization magic-angle-spinning nuclear magnetic resonance
<i>1H-NMR</i>	-	Proton nuclear magnetic resonance
<i>ANOVA</i>	-	Analysis of variance
<i>Ca(OH)₂</i>	-	Calcium hydroxide
<i>CCG-NP</i>	-	Curcumin-chitosan loaded gold nanoparticle
<i>CH₃COOH</i>	-	Acetic acid
<i>DD</i>	-	Degree of deacetylation
<i>DMSO</i>	-	Dimethyl sulfoxide
<i>DSC</i>	-	Differential scanning calorimetry
<i>EDX</i>	-	Energy dispersive x-ray
<i>FE-SEM</i>	-	Field emission scanning electron microscope
<i>FTIR</i>	-	Fourier-transform infrared spectroscopy
<i>GlcNAc</i>	-	1,4 linked N-acetylglucosamine
<i>h</i>	-	Hour
<i>H₂O₂</i>	-	Hydrogen peroxide
<i>H₂SO₄</i>	-	Sulphuric acid
<i>HCl</i>	-	Hydrochloric acid
<i>HCOOH</i>	-	Formic acid
<i>HNO₃</i>	-	Nitric acid
<i>IC₅₀</i>	-	The half-maximal inhibitory concentration
<i>IR</i>	-	Infrared
<i>K₂CO₃</i>	-	Potassium carbonate
<i>KMnO₄</i>	-	Potassium permanganate
<i>KOH</i>	-	Potassium hydroxide

<i>M</i>	-	Molarity
<i>min</i>	-	Minute
<i>ml</i>	-	Milliliter
<i>mm</i>	-	Millimeter
<i>MTT</i>	-	[3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
<i>N</i>	-	Nomality
<i>Na₂CO₃</i>	-	Sodium carbonate
<i>Na₂SO₄</i>	-	Sodium sulphate
<i>NaClO</i>	-	Sodium hypochlorite
<i>NaOH</i>	-	Sodium hydroxide
<i>nm</i>	-	Nanometer
<i>OD</i>	-	Optical density
<i>P₂O₅</i>	-	Phosphorus pentoxide
<i>RSM</i>	-	Response surface methodology
<i>SEM</i>	-	Scanning electron microscopy
<i>SO₂</i>	-	Sulphur dioxide
<i>SPR</i>	-	Surface plasmon resonance
<i>TGA</i>	-	Thermogravimetric analysis
<i>TPP</i>	-	Tripolyphosphate
<i>Tween-20</i>	-	Polysorbate 20
<i>Tween-80</i>	-	Polysorbate 80
<i>UTM</i>	-	Universiti Teknologi Malaysia
<i>UV-vis</i>	-	Ultra-Violet visible spectrophotometer
<i>w/v</i>	-	Weight/volume
<i>w/w</i>	-	Weight/weight
<i>WBC</i>	-	Water binding capacity
<i>WHO</i>	-	World health organization
<i>XRD</i>	-	X-ray diffraction
α	-	Alpha
β	-	Beta
γ	-	Gamma
μl	-	Microliter

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Certificate of analysis for antimicrobial testing screening results by Research Centre, Universiti Teknologi Malaysia	203
B	XRD spectra of <i>O. rhinoceros</i> chitosan isolated from <i>O. rhinoceros</i> and commercial chitosan purchased from isolated from shrimp shells where the standard is chitosan d-ascorbate ($C_{12}H_{21}NO_{11}$) _n	206
C	XRD spectra of <i>O. RHINOCEROS</i> chitin isolated from <i>O. rhinoceros</i> and commercial chitin (Com chitin) purchased from Sigma Aldrich isolated from shrimp shells where the standard is N-acetyl-d-glucosamine ($C_8H_{15}NO_6$)	208
D	Cytotoxicity report from UPM_MAKNA Cancer Research Laboratory (CanRes), Institute of Bioscience, Universiti Putra Malaysia	210

LIST OF PUBLICATIONS

Journals:

- (i) **N. Alyani Zainol Abidin**, F. Kormin, N. Akhma Zainol Abidin, N.A.F Mohamed Anuar, M.F. Abu Bakar (2020). “The potential of insects as alternative sources of chitin: An overview on the chemical method of extraction from various sources”, International Journal of Molecular Sciences. Vol. 21, No. 14, pp. 4978.
- (ii) **N.A. Zainol Abidin**, F. Kormin, N. Akhma Zainol Abidin, M.F. Abu Bakar, A.C. Iwansyah (2023). “Anti-tyrosinase activities of curcumin-chitosan gold nanoparticles synthesized from beetle (*Oryctes rhinoceros*)”, Asian Journal of Chemistry. Vol. 35, No. 1, pp. 79-82.

LIST OF AWARDS

- (i) **Silver medal in International Research and Innovation Symposium and Exposition 2022 (RISE2022):**

N. Alyani Zainol Abidin, F. Kormin, N. Akhma Zainol Abidin, "Synthesis and characterization of curcumin-chitosan loaded gold nanoparticles by *Oryctes rhinoceros*' chitin for cosmeceutical application."



PTT AUTHM
PERPUSTAKAAN TUNKU TUN AMINAH

REFERENCES

- Abdel-Raouf, N., Al-Enazi, N. M., & Ibraheem, I. B. (2017). Green biosynthesis of gold nanoparticles using Galaxaura elongata and characterization of their antibacterial activity. *Arabian Journal of Chemistry*, 10, S3029-S3039.
- Aggarwal, B. B., Kumar, A., & Bharti, A. C. (2003). *Anticancer Research*, 23, 363-398.
- Aggarwal, B. B., Sundaram, C., Malani, N., & Ichikawa, H. (2007). Curcumin: The Indian solid gold. *Advances in Botanical Research*, 595, 1-75.
- Ahmed, J. (2017). Thermal properties of gelatin and chitosan. In *Glass Transition and Phase Transitions in Food and Biological Materials*, 281 - 304.
- Ai, H., Wang, F., Yang, Q., Zhu, F., & Lei, C. (2008). Preparation and biological activities of chitosan from the larvae of housefly, *Musca domestica*. *Carbohydrate Polymers*, 72, 419-423.
- Akhtar, F., Alam Rizki, M. M., & Kumar Kar, S. (2012). Oral delivery of curcumin bound to chitosan nanoparticles cured *Plasmodium yoelii* infected mice. *Biotechnology Advances*, 30, 310-320.
- Ali, M. E., Mustafa, S., Hashim, U., Che Man, Y. B., & Foo, K. L. (2012). Nanoprobe for the determination of pork adulteration in burger formulations. *Journal of Nanomaterials*, 2012, 1-7.
- Allen, T. M., & Cullins, P. R. (2013). Liposomal drug delivery systems: From concept to clinical applications. *Advance Drug Delivery*, 65, 36-48.
- Amanlou, N., Parsa, M., Rostamizadeh, K., Sadighian, S., & Moghaddam, F. (2019). Enhanced cytotoxic activity of curcumin on cancer cell lines by incorporating into gold/chitosan nanogels. *Materials Chemistry and Physics*, 226, 151-157.
- Amendola, V., Pilot, R., Frasconi, M., Marago, O. M., & Antonio lati, M. (2017). Surface plasmon resonance in gold nanoparticles: a review. *Journal of Physics: Condensed Matter*, 29, 203002.
- Anand, P., Kunnumakkara, A. B., Newman, R. A., & Aggarwal, B. (2007). Bioavailability of curcumin: problems and promises. *Molecular Pharmaceutics*, 4(6), 807-818.

- Andrew, C. A., Wan, A. C., & Tai, B. C. (2013). Chitin- A promising biomaterial for tissue engineering and stem cell technologies. *Biotechnology Advances*, 31(8), 1776-1785.
- Anitha, A., Deepagan, V. G., Divya Rani, V. V., Menon, D., Nair, S. V., & Jayakumar, R. (2011). Preparation, characterization, *in vitro* drug release and biological studies of curcumin loaded dextran sulphate-chitosan nanoparticles. *Carbohydrates Polymers*, 84(3), 1158-1164.
- Aranaz, I., Acosta, N., Civera, C., Elorza, B., Mingo, J., Castro, C., Gandia, M. L., Caballero, A. H. (2018). Cosmetics and Cosmeceutical applications of chitin, chitosan and their derivatives. *Polymers*, 10(213), 1-25.
- Arbia, W., Arbia, L., Adour, L., & Amrane, A. (2013). Chitin extraction from crustacean shells using biological methods- a review. *Food Technology and Biotechnology*, 51, 12-25.
- Asasutjarit, R., Sorrachaitawatwong, C., Tipchuwong, N., & Pouthai, S. (2013). Effect of formulation compositions on particle size and zeta potential of Diclofenac sodium-loaded chitosan nanoparticles. *World Academy of Sciences, Engineering and Technology*, 7(9), 568-570.
- Aydar, A. Y. (2018). Utilization of response surface methodology in optimization of extraction of plant materials. In *Statistical Approaches With Emphasis on Design of Experiments Applied to Chemical Processes* (pp. 157-170).
- Azuma, K., Ifuku, S., Osaki, T., Okamoto, Y., & Minami, S. (2014). Preparation and biomedical applications of chitin and chitosan nanofibers. *Journal of Biomedical Nanotechnology*, 10, 2891-2920.
- Bailey, A. J. (2001). Molecular mechanisms of ageing in connective tissues. *Mechanisms of ageing and Development*, 122, 765-755.
- Bedford, G. O. (1980). Biology, ecology and control of palm rhinoceros beetles. *Annual Review of Entomology*, 25, 309-339.
- Belteky, P., Ronavari, A., Igaz, N., Szerencses, B., Toth, I. Y., Pfeiffer, I., Konya, Z. (2019). Silver nanoparticles: aggregation behavior in biorelevant conditions and its impact on biological activity. *International Journal of Nanomedicin*, 14, 667-687.
- Bianchi-Bosisio, A. (2005). Proteins: Physiological samples. In *Encyclopedia of analytical science (second edition)* (pp. 357-375). Elsevier.

- Bisht, S., Feldmann, G., Soni, S., Ravi, R., Karikar, C., Maitra, A., & Maitra , A. (2007). Polymeric nanoparticle-encapsulated curcumin ("nanocurcumin"): a novel strategy for human cancer therapy. *Journal of Nanobiotechnology.*, 5, 3.
- Black, M., & Schwartz, H. (1950). The estimation of chitin and chitin nitrogen in crawfish waste and derived products. *Analyst.*, 889(1950), 185.
- Bohn, R. E., & Terwiesch, C., (1999). The econominc of yield-driven processes. *Journal of Operations Management.*, 18(1), 41-59.
- Borchert, H., Shevchenko, E. V., Robert, A., Mekis, I., Kornowski, A., Grubel, G., & Weller, H. (2005). Determination od nanocrystal sizes: a comparison of TEM, SAXS, and XRD studies of highly monodisperse CoPt₃ particles. *Langmuir*, 21(5), 1931-1936.
- Bough, W. A., Salter, W. L., Wu, A. C., & Perkins, B. E. (1978). Influence of manufacturing variables on the characteristics and effectiveness of chitosan porducts. Chemical composition, viscosity, and molecular weight distribution of chitosan products. *Biotechnology and Bioengineering*, 20, 19-31.
- Braconnot, H. (1811). Sur la nature des champignons. *Annales de Chimie et de Physique*, 79, 265-304.
- Braydich-Stolle, L., Hussain, S., Schlager, J. J., & Hofmann, M.-C. (2005). *In vitro* cytotoxicity of nanoparticles in mammalian germline stem cells. *Toxicological Sciences*, 88(2), 412-419.
- Brine, C. J., & Austin, P. R. (1981). Chitin variablity with species and method of preparation. *Comparative Biochemistry and Physiology*, 69, 283-286.
- Brown, S. D., Nativo, P., Smith, J.-A., Stirling, D., Edwards, P. R., Venugopal, B., Wheate, N. J. (2010). Gold nanoparticles for the improved anticancer drug delivery of the active component of oxaliplatin. *Journal of the American Chemical Society*, 132(13), 4678-4684.
- Budishevka, O., Popadyuk, N., Musyanovych, A., Kohut, A., Donchak, V., Voronov, A., Voronov, S. Formation of three-dimensional polymer stuctures through radical and ionic reactions of peroxychitosan. In *Studies in Natural Products Chemistry; Bioactive Natural Products Volume 64*, (pp. 365 - 390)
- Canovi, M., Lucchetti, J., Stravalaci, M., Re, F., Moscatelli, D., Bigini, P., Gobbi, M. (2012). Application of surface plasmon resonance (SPR) for the characterization of nanoparticles developed for biomedical purposes. *Sensors*, 12, 16420-16432.

- Cardenas, G., Cabrera, G., Taboada, E., & Miranda, P. S. (2004). Chitin characterization by SEM, FTIR, XRD and ^{13}C cross polarization/mass angle spinning NMR. *Journal of Applied Polymer Science*, 93, 1876-1885.
- Cardillo, A., & Morganti, P. (1994). Fast and noninvasive method for assessing skin hydration. *Journal of Applied Cosmetology*, 12, 11-16.
- Casadidio, C., Peregrina, D. V., Gigliobianco, M. R., Deng, S., Censi, R., & Di Martino, P. (2019). Chitin and chitosans: Characteristics, eco-friendly processes, and applications in cosmetic science. *Marine drugs* 17(6), 369.
- Catley, A. (1969). The coconut rhinoceros beetle *Oryctes rhinoceros* (L.). *PANS*, 15, 18-30.
- Cerqueira-Coutinho, C., Santos-Oleivera, R., dos Santos, & E., Mansur, C. R. (2015). Development of a photoprotective and antioxidant nanoemulsion containing chitosan as an agent for improving skin retention. *Engineering in Life Sciences*, 15, 593-604.
- Chandrkrachang, S. (2002). The applications of chitin in agriculture in Thailand. *Advance Chitin Science*, 5, 458-462.
- Chang, K. L., & Tsai, G. (1997). Response surface optimization and kinetics of isolating chitin from pink shrimp (*Solenocera melancho*) shell waste. *Journal of Agricultural and Food Chemistry*, 45, 1900-1904.
- Chang, T. S. (2009). An update review of tyrosinase inhibitors. *Internationl Journal of Molecular Sciences*, 10, 2440-2475.
- Chen, C. S., Liau, W. Y., & Tsai, G. J. (1998). Antibacterial effects of N-sulfonated and N-sulfobenzoyl chitosan and application to oyster preservation. *Journal of Food Protection*, 64, 1124-1128.
- Chen, J., Huang, Q., Du, Q. G., Zhao, D., Xu, F., Pan, J., & Nathan, A. (2014). Localized surface plasmon resonance enhanced quantum dot light-emitting diodes via quantum dot-capped gold nanoparticles. *RSC Advances*, 4(101), 57574-57579.
- Chen, W., Yu, H., Li, Q., Liu, Y., & Li, J. (2011). Ultralight and highly flexible aerogels with long cellulose I nanofibers. *Soft Matter*, 7(21), 10360-10368.
- Cheng, A. L., Hsu, C. H., Lin, J. K., Hsu, M. M., Ho, Y. F., Shen, T. S., Hsieh, & C. Y. (2001). Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Research*, 21, 2895-2900.

- Chereddy, K. K., Coco, R., Memvanga, P. B., Ucakar, B., des Rieux, A., Vandermeulen, G., & Preat, V. (2013). Combined effect of PLGA and curcumin on wound healing activity. *Journal of Controlled Release*, 171(2), 208-215.
- Cho, Y.-W., Cho, Y.-N., Chung, S.-H., Yoo, G., & Ko, S. W. (1999). Water-soluble chitin as a wound healing accelerator. *Biomaterials*, 20(22), 2139-2145.
- Chuah, L. H., Billa, N., Roberts, C. J., Burley, J. C., & Manickam, S. (2011). Curcumin-containing chitosan nanoparticles as a potential mucoadhesive delivery system to the colon. *Pharmaceutical Development and Technology*, 1-9.
- Chung, G. F., Sim, S. C., & Tan, M. W. (September 1991). Chemical control of rhinoceros beetles in the nursery and immature oil palms. *PORIM International Palm Oil Development Conference- Progress, Prospect and Challenges Towards the 21st century*. Kuala Lumpur.
- Coelho, J. F., Ferreira, P. C., Alves, P., Cordeiro, R., Fonseca, A. C., Gois, J. R., & Gil, M. H. (2010). Drug delivery systems: Advanced technologies potentially applicable in personalized treatments. *The Official Journal of the European Association for Predictive, Preventive and Personalised Medicine*, 1(1), 164-209.
- Cohen, E. (2009). Chitin. In *Encyclopedia of insects (second edition)* (pp. 156-157). Academic Press.
- Corazzari, I., Nistico, R., Turci, F., Faga, M. G., Franzoso, F., Tabasso, S., & Magnacca, G. (2015). Advanced physico-chemical characterization of chitosan by means of TGA-coupled on-line with FTIR and GCMS: Thermal degradation and water adsorption capacity. *Polymer Degradation and Stability*, 112, 1-9.
- Dai, M., Zheng, X., Xu, X., Kong, X., Li, X., Guo, G., Qian, Z. (2009). Chitosan-alginate sponge: Preparation and application in curcumin delivery for dermal wound healing in rat. *BioMed Research International*, 2009, 8.
- Daraghmeh, N. H., Chowdhry, B. Z., Leharne, S. A., Al Omari, M. M., & Badwan, A. A. (2011). Chitin. *Profiles on Drug Substances, Excipients and Related Methodology*, 36.

- Das, R. K., Kasoju, N., & Bora, U. (2010). Encapsulation of curcuminin alginat-chitosan-pluronic composition nanoparticles for delivery to cancer cells. *Nanomedicine*, 6(1), 153-160.
- De Smedt, S. C., Demeester, J., & Hennink, W. E. (2000). Cationic polymer based gene delivery systems. *Pharmacology Research*, 17, 113-126.
- Di Guglielmo, C., Lopez, D. R., De Lapuente, J., Mallafre, J. M., & Suarez, M. B. (2010). Embryotoxicity of cobalt ferrite and gold nanoparticles: a first *in vitro* approach. *Reproductive Toxicology*, 30(2), 271-276.
- Domard, A., & Rinaudo, M. (1983). Preparation and characterization of fully deacetylated chitosan. *International Journal of Biological Macromolecules*, 5, 49-52.
- Dracelos, Z. D. (2007). Skin lightening preparations and the hydroquinone controversy. *Dermatologic Theraphy*, 20, 308-313.
- Dreaden, E. C., Austin, L. A., Mackey, M. A., & El-Sayed, M. A. (2012). Size matters: gold nanoparticles in targeted cancer drug delivery. *Therapeutic Delivery*, 3(4), 457-478.
- Du, J., Zhou, Z., Zhang, X., Wu, S., Xiong, J., Wang, W., & Luo, Q. (2017). Biosynthesis of gold nanoparticles by flavonoids from *Lilium casa blanca*. *Journal of Cluster Science*, 28(6), 3149-3158.
- Dudhani, A. R., & Kosaraju, S. L. (2010). Bioadhesive chitosan nanoparticles: preparation and characterization. *Carbohydrate Polymers*, 81, 243-251.
- Duong, N. T., & Nghia, N. D. (2014). Kinetics and optimization of the deproteinization by pepsin in chitin extraction from white shrimp shell. *Journal of Chitin and Chitosan Science*, 2, 1-8.
- El-Ashram, S., El-Samad, L. M., Basha, A. A., & El Wakil, A. (2021). Naturally-derived targeted therapy for wound healing: Beyond classical strategies. *Pharmacological Research*, 170, 105749.
- Enumo, A. J., Argenta, D. F., Bazzo, G. C., Caon, T., Stulzer, H. K., & Parize, A. L. (2020). Development of curcumin-loaded chitosan/pluronic membranes for wound healing applications. *International Journal of Biological Macromolecules*, 163, 167-179.
- Fadlaoui, S., El Asri, O., Mohammed, L., Sihame, A., Omar, A., & Melhaoui, M. (2019). Isolation and characterization of chitin from shells of the freshwater

- crab *Potamon algeriense*. *Progress on Chemistry and Application of Chitin and its Derivatives*, 24, 23-35.
- Fan, Y., Saito, T., & Isogai, A. (2010). Individual chitin nano-whiskers prepared from partially deacetylated alpha-chitin by fibril surface cationization. *Carbohydrate Polymers*, 79, 1046-1051.
- Feng, Y., Li, J., Wang, B., Tian, X., Chen, K., Zeng, J., Gao, W. (2018). Novel nanofibrillated cellulose/chitin whisker hybrid nanocomposites and their use for mechanical performance enhancements. *Bioresources*, 13(2), 3030-3044.
- Fernandez-kim, S. O. (2004). Physicochemical and functional properties of crawfish chitosan as effected by different processing protocols. *LSU Master's Thesis*. 1338.
- Fernando, L. T., Poblete, M. S., Ongkiko, A. M., & Diaz, L. L. (2016). Chitin extraction and synthesis of chitin-based polymer films from Philiphine blue swimming crab (*Portumus pelagicus*) shells. *Procedia Chemistry*, 19, 462-468.
- Focher, B., Naggi, A., Torri, G., Cosani, A., & Terbojevich, M. (1992). Chitosans from *Euphausia superba*. 2: Characterization of solid state structure. *Carbohydrate Polymers*, 18, 43-49.
- Franconetti, A., Carnerero, J. M., Prado-Gotor, R., Cabrera-Escribano, F., & Jaime, C. (2019). Chitosan as a capping agent: Insights on the stabilization of gold nanoparticles. *Carbohydrate Polymers*, 1(207), 806-814.
- Jimtaisong, A. & Saewan, N. (2014). Utilization of carboxymethyl chitosan in cosmetics. *International Journal of Cosmetic Science*, 36, 12-21.
- Jung, H. Lee, Y. J., Yoon, Q. B. (2018). Effect of moisture content on the grinding process and powder properties in food: A review. *Processes*, 6(6), 69.
- Gao, W., Xu, K., Ji, L., & Tang, B. (2011). Effect of gold nanoparticles on glutathione depletion-induced hydrogen peroxide generation and apoptosis in HL7702 cells. *Toxicology Letters*, 205(1), 86-95.
- Garcea, G., Berry, D. P., Jones, D. J., Singh, R., Dennison, A. R., Farmer, P. B., Gescher, A. J. (2005). Consumption of the putative chemopreventive agent curcumin by cancer patients; assessment of curcumin levels in the colorectum and their pharmacodynamic consequences. *Cancer Epidemiology Biomarkers & Prevention*, 14(1), 120-125.

- Garcea, G., Jones, D. J., Singh, R., Dennison, A. R., Farmer, P. B., Sharma, R. A., Berry, D. P. (2004). Detection of curcumin and its metabolites in hepatic tissue and portal blood of patients following oral administration. *British Journal of Cancer*, 90(5), 1011-1015.
- Gilchrest, B. A., & Krutman, J. (2006). *Skin aging*. Verlag Berlin, Germany: Springer.
- Goel, A., Kunnumakkara, A. B., & Aggarwal , B. B. (2007). Curcumin as "Curecumin": From kitchen to clinic. *Biochemical Pharmacology*, in press.
- Gonçalves, .G. M. S., da Silva, G. H., Barros, P. P., Srebernich, S. M., Shiraishi, C. T. C., de Camargos, V. R., LascaGornall, T. B. (2014). Use of *Curcuma longa* in cosmetics: extraction of curcuminoid pigments, development of formulations, and in vitro skin permeation studies. *Brazillian Journal of Pharmaceutical Sciences*, 50(4), 885-893.
- A. G., Bardawill, C. J., & David, M. M. (1949). Determination of serum proteins by means of the biuret reaction. *Journal of Biology Chemistry*, 177, 751-766.
- Grenha, A., Seijo, B., & Remunan-Lopez, C. (2005). Microencapsulated chitosan nanoparticles for lung protein delivery. *European Journal Pharmacology Science*, 25, 427-437.
- Guo, N., Sun, J., Zhang, Z., & Mao, X. (2019). Recovery of chitin and protein from shrimp head waste by endogenous enzyme autolysis and fermentation. *Journal of Ocean University of China*, 18(3), 719-726.
- Hackman, R. H. (1954). Studies on chitin I. Enzymatic Degradation of chitin and chitin esters. *Australian Journal of Biological Sciences*, 7(2), 168-178.
- Haiss, W., Thanh, N. K., Aveyard, J., & Fernig, D. G. (2007). Determination of size and concentration of gold nanoparticles from Uv-vis spectra . *Analytical Chemistry*, 79, 4215-4221.
- Hajji, S., Younes, I., Ghorbel-Bellaj, O., Hajji, R., Rinaudo, M., Nasri, M., & Jellouli, K. (2014). Structural differences between chitin and chitosan extracted from three different marine sources. *International Journal of Biological Macromolecules*, 65, 298-306.
- Hamidi, M., Azadi, A., & Rafiei, P. (2008). Hydrogel nanoparticles in drug delivery. *Advanced Drug Delivery Reviews.*, 60, 1638-1649.
- Hariharan, S., Bhardwaj, V., Bala, I., Sitterberg, J., Bakowsky, U., & Ravi Kumar, M. (2006). Design of estradiol loaded PLGA nanoparticulate formulations: a

- potential oral delivery system for hormone therapy. *Pharmaceutical Research*, 23, 184-195.
- Harris, R., Lecumberri, E., Mateos-Aparicio, I., Mengíbar, M., Heras, A. (2011). Chitosan nanoparticles and microspheres for the encapsulation of natural antioxidants extracted from *Ilex paraguariensis*. *Carbohydrates Polymer*, 84, 803–806.
- Hatcher, H., Planalp, R., Cho, J., Torti, F. M., & Torti, S. V. (2008). Curcumin: from ancient medicine to current clinical trials. *Cellular and Molecular Life Sciences*, 65, 1631-1652.
- Henry Garcia, Y., Troncoso-Rojas, R., Tiznado-Hernandez, M. E., Baez-Flores, M. E., Carvajal-Mill, E., Rason-Chu, A., Martinez-Robinson, K. G. (2019). Enzymatic treatments as alternative to produce chitin fragments of low molecular weight from alternaria alternata. *Journal of Applied Polymer Science*, 136(15), 47339.
- Honary, S., & Zahir, F. (2013). Effect of zeta potential on the properties of nano-drug delivery systems-a review (Part 1). *Tropical Journal of Pharmaceutical Research*, 12(2), 225-264.
- Hosoya, T., Nakata, A., Yamasaki, F., Abas, F., Shaari, K., Lajis, N. H., & Morita, H. (2012). Curcumin-like diarylpentanoid analogues as melanogenesis inhibitors. *J. Nat. Med.*, 66(1), 166-176.
- Hossain, M. S., & Iqbal, A. (2014). Production and characterization of chitosan from shrimp waste. *Journal of the Bangladesh Agricultural University*, 12(1), 153-160.
- Huang, H., Yuan, Q., & Yang, X. (2005). Morphology study of gold-chitosan nanocomposites. *Journal of Colloid and Interface Science*, 282(1), 26-31.
- Huang, T., & Xu, X. N. (2010). Synthesis and characterization of tunable rainbow colored colloidal silver nanoparticles using single-nanoparticle plasmonic. *Journal of Materials Chemistry*, 20(44), 9867-9876.
- Huang, W. C., Zhao, D., Guo, N., Xue, C., & Mao, X. (2018). Green and facile production of chitin from crustacean shells using a natural deep eutectic solvent. *Journal of Agricultural and Food Chemistry*, 66(45), 11897-11901.
- Huang, Y., Fang, Y., Chen, L., & Zhang, L. (2017). One-step synthesis of size-tunable gold nanoparticles immobilized on chitin nanofibrils via green

- pathway and their potential applications. *Chemical Engineering Journal*, 315, 573-582.
- Huet, G., Hadad, C., Husson, E., Laclef, S., Lambertyn, V., Farias, M. A., Van Nhien, A. N. (2020). Straightforward extraction and selective bioconversion of high purity chitin from *Bombyx eri* larva: Toward an integrated insect biorefinery. *Carbohydrate Polymers*, 228.
- Ibitoye, E. B., Lokman, I. H., Hezmee, M. N., Goh, Y. M., Zuki, A. B., & Jimoh, A. A. (2018). Extraction and physicochemical characterization of chitin and chitosan isolated from house cricket. *Biomedical Materials*, 13.
- Idris, A., Kormin, F., Noordin, M. Y. (2006). Application of response surface methodology in describing the performance of thin film composite membrane. *Separation and Purification Technology*, 49(3), 271-280.
- Ifuku, S., Nogi, M., Abe, K., Yoshioka, M., Morimoto, M., Saimoto, H., & Yano, H. (2009). Preparation of chitin nanofibers with a uniform width as alpha-chitin from crab shells. *Biomacromolecules*, 10, 1584-1588.
- Illum, L. (1998). Chitosan and its use as a pharmaceutical excipient. *Pharmaceutical research*, 15(9), 1326-1331.
- Ilyas, H. N., Mahmood Zia, K., R, Rehman, S., Ilyas, R., & Sultana, S. (2021). Utilization of shellfish industrial waste for isolation, purification, and characterizations of chitin from crustacean's sources in Pakistan. *Journal of Polymers and the Environment*, 29, 2337-2348.
- Ito, I., Yoneda, T., Omura, Y., Osaki, T., Ifuku, S., Saimoto, H., Minami, S. (2015). Protective effect of chitin urocanate nanofibres against ultraviolet radiation. *Marine Drugs*, 13(12), 7463-7475.
- Jahanizadeh, S., & Chegeny, M. (2019). Curcumin-encapsulated polysaccharide nanocomposite: Formulation design, optimization and characterization. *Journal of Bioengineering Research*, 1(1), 54-62.
- Jana, J., Ganguly, M., & Pal, T. (2016). Resonance effect of metal nanoparticles for practical spectroscopic application. *The Royal Society of Chemistry*, 6(89), 86174.
- Jayakumar, R., Prabaharan, M., Nair, S. V., & Tamura, H. (2010). Novel chitin and chitosan nanofibers in biomedical applications. *Biotechnology Advances*, 28(1), 142-150.

- Jayaprakasha, G. K., Jagan, L., Rao, M., & Sakariah, K. K. (2006). Antioxidant activities of curcumin, demethoxycurcumin and bisdemethoxycurcumin. *Food Chemistry*, 98, 720-724.
- Je, J.-Y., & Kim, S.-K. (2012). Chitooligosaccharides as potential nutraceuticals: Production and bioactivities. *Advances in Food and Nutrition Research*, 65, 321-336.
- Jennifer, C., Stephie, C. M., Abhishri, S. B., & Shalini, B. U. (2012). A review on skin whitening property of plant extracts. *International Journal of Pharma and Bio Sciences*, 3(4), 332-347.
- Jeon, Y.-J., Shahidi, F., & Kim, S.-K. (2012). Preparation of chitin and chitosan oligomers and their applications in physiological functional foods. *Food Reviews International*, 16(2), 159-176.
- Jeong, J. H., Park, T. G., & Kim, S. H. (2011). Self-assembled and nanostructured siRNA delivery systems. *Pharmacology Research*, 28, 2072-2085.
- Kanthraj, G. R. (2010). Skin-lightening agents: new chemical and plant extracts-ongoing search for the Holy Grail! *Indian Journal of Dermatology, Venereology and Leprology*, 76, 3-6.
- Kar, S. K., Akhtar, F., Ray, G., & Pandey, A. K. (A1 Aug, 2011). *United States Patent No. patent US 20110290399*.
- Karami, Z., Sadighian, S., Rostamizadeh, K., Parsa, M., & Rezaee, S. (2016). Naproxen conjugated mPEG-PCL micelles for dual triggered drug delivery. *Materials Science and Engineering: C*, 61, 665-673.
- Kardas, I., Struszczyk, M. H., Kucharska, M., van den Broek, L. A., van Dam, J. E., & Ciechanska, D. (2012). Chitin and chitosan as functional biopolymers for industrial applications. *The European Polysaccharide Network of Excellence*, 329-373.
- Kasaai, M. R. (2009). Various methods for determination of the degree of N-acetylation of chitin and chitosan: a review. *Journal of Agricultural and Food Chemistry*, 57, 1667-1676.
- Kast, C. E., & Schnurch, A. B. (2002). Influence of the molecular mass on the permeation enhancing effect of different poly(acrylates). *STP Pharmaceutical Science*, 6, 351-356.

- Katas, H., Hussain, Z., & Ling, T. C. (2012). Chitosan nanoparticles as a percutaneous drug delivery system for hydrocortisone. *Journal of Nanomaterials*, 2012, 1-11.
- Kaul, S., Gulati, N., Verma, D., Mukherjee, S., Nagaich, U. (2018). Role of nanotechnology in cosmeceuticals: A review of recent advances. *Journal of Pharmaceutics*, 2018, 3420204.
- Kaya, M., & Baran, T. (2015). Description of a new surface methodology for chitin extracted from wings of cockroach (*Periplaneta americana*). *International Journal of Biology of Macromolecules*, 75, 7-12.
- Kaya, M., Baublys, V., Satkauskiene, I., Akyuz, B., Bulut, E., Tubelyte, V. (2015a). First chitin extraction from *Plumatella repens* (Bryozoa) with comparison to chitins of insects and fungal origin. *International Journal of Biological Macromolecules*, 79, 126-132.
- Kaya, M., Baran, T., & Karaarslan, M. (2015b). A new method for fast chitin extraction from shells of crab, crayfish and shrimp. *Natural Product Research*, 29, 1477-1480.
- Kaya, M., Baran, T., Erdogan, S., Mentes, A., Ozusaglam, M. A., & Cakmak, Y. S. (2014a). Physicochemical comparison of chitin and chitosan obtained from larvae and adult Colorado potato beetle (*Leptinotarsa decemlineata*). *Materials Science and Engineering C*, 45, 72-81.
- Kaya, M., Baublys, V., Can, E., Satkauskiene, I., Bitim, B., Tubelyte, V., & Baran, T. (2014b). Comparison of physicochemical properties of chitins isolated from an insect (*Melolontha melolontha*) and a crustacean species (*Oniscus asellus*). *Zoomorphology*, 133(3), 285–293.
- Kaya, M., Bitim, B., Mutjaba, M., & Koyuncu, T. (2015c). Surface morphology of chitin highly related with the isolated body part of butterfly. *International Journal of Biology Macromolecules*, 81, 443-449.
- Kaya, M., Cakmak, Y. S., Baran, T., Asan-Ozusaglam, M., Mentes, A., & Tozak, K. O. (2014c). New chitin, chitosan and o-carboxymethyl chitosan sources from resting eggs of *Daphnia longispina* (Crustacea): with physicochemical characterization, and antimicrobial and antioxidant activities. *Biotechnology and Bioprocess Engineering*, 19, 58-69.

- Kaya, M., Mujtaba, M., Bulut, E., Akyuz, B., Zelencova, L., & Sofi, K. (2015d). Fluctuation in physicochemical properties of chitins extracted from different body parts of honeybee. *Carbohydrate Polymers*, 132, 9-16.
- Kaya, M., Sargin, I., Tozak, K. O., Baran, T., Erdogan, S., & Sezen, G. (2013). Chitin extraction and characterization from *Daphnia magna* resting eggs. *International Journal of Biological Macromolecules*, 61, 459-464.
- Kaya, M., Seyyar, O., Baran, T., Erdogan, S., & Kar, M. (2014d). A physicochemical characterization of fully acylated chitin structure isolated from two spider species: with new surface morphology. *International Journal of Biology Macromolecules*, 65, 553-558.
- Kazemi, S., Pournmadadi, M., Yazdian, F., & Ghadami, A. (2021). The synthesis and characterization of targeted delivery curcumin using chitosan-magnetite-reduced graphene oxide as nano-carrier . *International Journal of Biological Macromolecules* , 186, 554-562.
- Khan, T. A., Peh, K. K., & Ch'ng, H. S. (2002). Reporting degree of acetylation values of chitosan: the influence of analytical methods. *Journal of Pharmacy & Pharmaceutical Sciences*, 5(3), 205-212.
- Khatib, S., Nerya, O., Musa, R., Shmuel, M., Tamir, S., & Vaya, J. (2005). Chalcones as potent tyrosinase inhibitors: the importance of a 2,4-substituted resorcinol moiety. *Bioorganic & Medicinal Chemistry*, 13, 433-441.
- Khoo, K. C., Ooi, P. A., & Ho, C. T. (1991). *Crop pests and their management in Malaysia*. Kuala Lumpur: Tropical Press Sdn. Bhd. .
- Khurana, P., Thatai, S., Wang, P., Lihitkar, P., Zhang, L., Fang, Y., & Kulkarni, S. K. (2012). Speckled SiO₂@Au core-shell paticles as surface enhanced raman scattering probes. *Plasmonics*, 8(2), 185-191.
- Kim, D.-G.; Jeong, Y.-I.; Choi, C.; Roh, S.-H.; Kang, S.-K.; Jang, M.-K.; Nah, J.-W. (2006). Retinol-encapsulated low molecular water-soluble chitosan nanoparticles. *International Journal of Pharmaceutics*, 319, 130–138.
- Kim, M.-W., Song, Y.-S., Han, Y.-S., Jo, Y.-H., Choi, M.-H., Park, Y.-K., Jung, W.-J. (2017). Production of chitin and chitosan from the exoskeleton of adult two-spotted field crickets (*Gryllus bimaculatus*). *Entomological Research*, 47, 279-285.
- Knorr, D. (1984). Use of chitinous polymers in food: A challenge for food research and development. *Food Technology*, 38(1), 92-97.

- Ko, J., Park, H. J., Hwang, S., Park, J., & Lee, J. (2002). Preparation and characterization of chitosan microparticles intended for controlled drug delivery. *International Journal of Pharmaceutics*, 249(165), 67-74.
- Kotze, A. F., de Leeuw, B. J., Lueben, H. L., de Boer, A. G., Verhoef, J. C., & Junginger, H. E. (1997). Chitosans for enhanced delivery of therapeutic peptides across intestinal epithelia: in vitro evaluation of Caco-2 cell monolayer. *International Journal of Pharmaceutics*, 159, 243-253.
- Kucukgulmez, A., Celik, M., Yanar, Y., Sen, D., Polat, H., & Kadak, A. E. (2011). Physicochemical characterization of chitosan extracted from Metapenaeis stebbingi shells. *Food Chemistry*, 126, 1144-1148.
- Kumar, S., & Koh, J. (2012). Physicochemical optical and biological activity of chitosan-chrome derivative for biomedical applications. *International Journal of Molecular Sciences*, 13, 6102-6116.
- Kumari, A., Yadav, S. K., & Yadav, S. C. (2010). Biodegradable polymeric nanoparticles based drug delivery systems. *Colloids and Surfaces B: Biointerfaces*, 75, 1-18.
- Kuria, K. (2001). Controlled fractionation of the polysaccharide chitin. *Progress in Polymer Science*, 26(9), 1921-1971.
- Labandeira, C. C., & Sepkoski, J. J. (1993). Jr. Insect diversity in the fossil record. *Science*, 261, 310-315.
- Langoth, N., Kahlbacher, H., Schoffmann, G., Schmerold, I., Schuh, M., Franz, S., Schnurch, A. B. (2006). Thiolated chitosans: design and *in vivo* evaluation of a mucoadhesive buccal peptide drug delivery system. *Pharmaceutical Research*, 23, 573-579.
- Latif, M. S., Abbas, S., Kormin, F., & Muhamad, I. I. (2020). Green synthesis and characterization of gold nanoparticles using ethanolic turmeric crude extract at neutral pH. *International Journal of Recent Technology and Engineering*, 8(6), 1535-1539.
- Latif, M. S., Kormin, F., Mustafa, M. K., Mohamad, I. I., Khan, M., Abbas, S., Mohamad Fuzi, S. F. (2018). Effect of temperature on the synthesis of *Centella asiatica* flavonoids extract-mediated gold nanoparticles: UV-visible spectra analyses. *AIP Conference Proceedings 2016*, 020071.

- Lee, E. K., & Koros, W. J. (2003). Membranes, Synthetic, Applications . In *Encyclopedia of Physical Science and Technology (Third Edition)* (pp. 79-344). Academic Press.
- Lee, J. H., Jang, J. Y., Park, C., Kim, B. W., Choi, Y. H., & Choi, B. T. (2010). Curcumin suppresses alpha-melanocyte stimulating hormone-stimulated melanogenesis in B16F10 cells. *International Journal of Molecular Medicine*, 26, 101-106.
- Lee, K.-H., Ab. Aziz, F. H., Syahida, A., Abas, F., Shaari, K., Ahmad Israf, D., & Haji Lajis, N. (2009). Synthesis and biological evaluation of curcumin-like diarylpentoid analogues for anti-inflammatory, antioxidant and anti-tyrosinase activities. *European Journal of Medicinal Chemistry*, 44, 3195-3200.
- Leiva, A., Bonardd, S., Pino, M., Saldias, C., Kortaberria, G., & Radic, D. (2015). Improving the performance of chitosan in the synthesis and stabilization of gold nanoparticles. *European Polymer Journal*, 68, 419-431.
- Le-Vinh, B., Nguyen Le, N.-M., Nazir, I., Matusczak, B., & Bernkop-Schnurch, A. (2019). Chitosan based micelle with zeta potential changing property for effective mucosal drug delivery. *International Journal of Biological Macromolecules*, 133, 647-655.
- Li, Q., Dunn, E. T., Grandmaison, E. W., & Goosen, M. F. (1992). Applications and properties of chitosan. *Journal of Bioactive and Compatible Polymers*, 7, 370-397.
- Liau, S. S., & Ahmad, A. (1993). The control of *Oryctes rhinoceros* by clean clearing and its effect on early yield in palm replants. *1991 PORIM International Palm Oil Development Conference MPOB*, (pp. 393-403). Bangi, Malaysia.
- Lim, S. H., & Hudson, S. M. (2004). Synthesis and antimicrobial activity of water-soluble chitosan derivative with a fiber-reactive group. *Carbohydrate Research*, 339, 313-319.
- Lin, J. K. (2007). Molecular targets of curcumin. *Advances in Experimental Medicine and Biology*, 595, 227-243.
- Liu, S., Sun, J., Yu, L., Zhang, C., Bi, J., Zhu, F., Yang, Q. (2012). Extraction and characterization of chitin from the beetle *Holotrichia parallela* motschulsky. *Molecules*, 17(4), 4604-4611.

- Lohman, B., Powell, J. A., Cingarapu, S., Aekeroy, C. B., Chakrabarti, A., Klabunde, K. J., Sorensen, C. M. (2012). Solubility of gold nanoparticles as a function of ligand shell and alkane solvent. *Physical Chemistry Chemical Physics*, 14(18), 6509-6513.
- Lopez Munoz, G. A., Pescador-Rojas, J. A., Ortega-Lopez, J., & Balderas-Lopez, J. A. (2012). Thermal diffusivity measurement of spherical gold nanofluids of different sizes/concentrations. *Nanoscale Research Letters*, 7(1), 423.
- Lorke, D. (1983). A new approach to practical acute toxicity testing. *Archives of Toxicology*, 54, 275-287.
- Lu, P.J., Fu, W.-E., Huang, S.-C., Lin, C.-Y., Ho, M.-L., Chen, Y.-P., & Cheng, H.-F. (2018). Methodology for sample preparation and size measurement of commercial ZnO nanoparticles. *Journal of Food and Drug Analysis*, 26, 628-636.
- Luer, S., Troller, R., & Aebi, C. (2012). Antibacterial and antiinflammatory kinetics of curcumin as a potential antimucositis agent in cancer patients. *Nutrition and cancer*, 64(7), 975-981.
- Machalowski, T., Wysokowski, M., Tsurkan, M. V., Galli, R., Schimpf, C., Rafaja, D., Ehrlich, H. (2019). Spider chitin: An ultrafast microwave-assisted method for chitin isolation from *Caribena versicolor* spider molt cuticle. *Molecules*, 24, 3736.
- Madhumathi, K., Sudheesh Kumar, P. T., Abhilash, S., Sreeja, V., Tamura, H., Manzoor, K., Jayakumar, R. (2010). Development of novel chitin/nanosilver composite scaffolds for wound dressing applications. *Journal of Material Science; Material Medicine*, 21, 807-813.
- Mahmood Zia, K., Zuber, M., Ahmad Bhatti, I., Barikani, M., & Ahmad Sheikh, M. (2009). Evaluation of biocompatibility and mechanical behavior of polyurethane elastomers based on chitin/1,4-butane diol blends. *International Journal of Biological Macromolecules*, 44(1), 18-22.
- Mai, Z., Chen, J., He, T., Hu, Y., Dong, X., Zhang, H., Zhou, W. (2017). Electrospray biodegradable microcapsules loaded with curcumin for drug delivery systems with high bioactivity. *Royal Society of Chemistry Advances*, 7(3), 1724-1734.

- Majtan, J., Bilikova, K., Markovic, O., Grof, J., Kogan, G., & Simuth, J. (2007). Isolation and characterization of chitin from bumblebee (*Bombus terrestris*). *International Journal of Biological Macromolecules.*, 40, 237-241.
- Manjeri, G., Muhamad, R., Faridah, Q. Z., & Tan, S. G. (2013). Morphometric analysis of *Oryctes rhinoceros* (L.) (Coleoptera: Scarabaeidae) from oil palm plantations. *The Coleopterists Bulletin*, 67(2), 194-200.
- Marei, N. H., Samiee, E. E., Salah, T., Saad, G. R., & Elwahy, A. M. (2015). Isolation and characterization of chitosan from different local insects in Egypt. *International Journal of Biological Macromolecules.*
- Martins, A. F., de Oliveira, D. M., Pereira, A. G., Rubira, A. F., & Muniz, E. C. (2012). Chitosan/TPP microparticles obtained by microemulsion method applied in controlled release of heparin. *International Journal of Biological Macromolecules*, 51, 1127-1133.
- McGavin, G. C. (2000). *Insects; Spider and other terrestrial arthropods*. London: Dorling Kindersley Limited.
- Melo-Silveria, R. F., Fidelis, G. P., Costa, M. P., Telles, C. S., Dantas-Santos, N., Elias, S. O., Oliveira Rocha, H. A. (2012). *In vitro* antioxidant, anticoagulant and antimicrobial activity and in inhibition of cancer proliferation by xylan extracted form corn cobs. *International Journal of Molecular Sciences*, 1, 409-426.
- Mendonca, L. M., Dos Santos, G. C., Antonucci, G. A., Dos Santos, A. C., Bianchi Mde, L., & Antunes, L. M. (2009). Evaluation of the cytotoxicity and genotoxicity of curcumin in pc12 cells. *Mutation Research*, 675, 29-34.
- Mesaik, M. A., Zaheer, U. H., Murad, S., Ismail, Z., Abdullah, N. R., Gill, H. K., Choudhary, M. I. (2006). Biological and molecular docking studies on coagulin-H: human IL-2 novel natural inhibitor. *Molecular Immunology*, 43, 1855-1863.
- Mezzana, P. (2009). Clinical efficacy of a new chitin nanofibrils based gel in wound healing. *Acta Chirurgiae Plasticae*, 50(3), 81-84.
- Mislovicova, D., Masarova, J., Bendzalova, K., Soltes, L., & Machova, E. (2000). Sonication of chitin-glucan, preparation of water-soluble fractions and characterization by HPLC. *Ultrasonic Sonochemistry*, 7, 63-68.
- Mofazzal Jahromi, M. A., Al-Musawi, S., Pirestani, M., Ramandi, M. F., Ahmadi, K., Rajayi, H., Mirnejad, R. (2014). Curcumin-loaded Chitosan

- Tripolyphosphate Nanoparticles as a safe, natural and effective antibiotic inhibits the infection of *Staphylococcus aureus* and *Pseudomonas* in vivo. *Iranian Journal of Biotechnology*, 12(3), 1-8.
- Mohammed, M. H., Williams, P. A., & Tverezovskaya, O. (2013). Extraction of chitin from prawn shells and conversion to low molecular mass chitosan. *Food Hydrocolloids*, 31(2), 166-171.
- Morganti, P. (2005). Patent No. Int patent PCT/IB2005/053576.
- Morganti, P. (2009). Chemical-physical activity of chitin nanofibrils to stabilize cosmetic emulsion and increase skin penetration. *Journal of Applied Cosmetology*, 27(4), 251-270.
- Morganti, P. (2010). Chitin nanofibrils for cosmetic delivery. *Cosmetics, Dermatological Sciences and Applications*, 125(4), 36-39.
- Morganti, P., & del Ciotto, P. (2011). Chemical physical properties and biocompatibility of chitin nanofibrils complexes. *Journal of Applied Cosmetology*, (in press).
- Morganti, P., Fabrizi, G., Palombo, M., Ruocco, E., Cardillo, A., & Morganti, G. (2008). Chitin-nanofibrils: A new active cosmetic carrier. *Journal of Applied Cosmetology*, 26, 113-128.
- Morganti, P., Morganti, G., & Morganti, A. (2011). Transforming nanostructured chitin from crustacean waste into beneficial health products: a must for our society. *Nanotechnology, Science and Applications*, 4, 123-129.
- Morganti, P., Morganti, G., Fabrizi, G., & Cardillo, A. (2008). A new sun to rejuvenate the skin. *Journal of Applied Cosmetology*, 26, 159-168.
- Morganti, P., Palombo, P., Palombo, M., Fabrizi, G., Cardillo, A., Fabiano, S., Mezzana, P. (2012). A phosphatidylcholine hyaluronic acid chitin–nanofibrils complex for a fast skin remodeling and a rejuvenating look. *Clinical, Cosmetic and Investigational Dermatology*, 5, 213-220.
- Morsy, R., Ali, S.S., El-Shetehy, M. (2017) Development of hydroxyapatite-chitosan gel sunscreen combating clinical multidrug-resistant bacteria. *Journal of Molecular Structure*, 1143, 251–258.
- Muhammad Mailafiya, M., Abubakar, K., Danmaigoro, A., Chiroma, S. M., Abdul Rahim, E., Mohd Moklas, M. A., & Zakaria, Z. A. (2019). Evaluation of in vitro release kinetics and mechanisms of curcumin-loaded cockle shell-derived calcium carbonate nanoparticles. *Biomedpress*, 6(12), 3518-3540.

- Muzzarelli, R. A. (2011). Chitin nanostructure in living organisms. In *Chitin: Formation and Diagenesis*, edited by N. Gupta (pp. 1-34). Dordrecht: Springer.
- Muzzarelli, R., Cucchiara, M., & Muzzarelli, C. (2002). N-Carboxymethyl chitosan in innovative cosmeceutical products. *Journal of Applied Cosmetology*, 20, 201-208.
- Myers, R. H., Douglas, M. C., & Christine, A.-C. M. (2016). *Response surface methodology: process and product optimization using designed experiments*. John Wiley & Sons.
- Nahar, S. J., Kazuhiko, S., & Haque, S. M. (2012). Effect of polysaccharides including elicitors on organogenesis in protocorm-like body (PLB) of *Cymbidium insigne* *in vitro*. *Journal of Agricultural Science and Technology*, 2, 1029-1033.
- Nair, R. S., Morris, A., Billa, N., & Leong, C.-O. (2019). An evaluation of curcumin-encapsulation chitosan nanoparticles for transdermal delivery. *American Association of Pharmaceutical Scientists*, 20(29), 1-13.
- Nerya, O., Musa, R., Khatib, S., Tamir, S., & Vaya, J. (2004). Chalcones as potent tyrosinase inhibitors: the effect of hydroxyl positions and numbers. *Phytochemistry*, 65, 1389-1395.
- Nguyen, V., Nguyen, V., & Hsieh, M.-F. (2013). Curcumin-loaded chitosan/gelatin composite sponge for wound healing application. *Polymeric Membrane Science and Technology*, 2013, 1-7.
- No, H. K., & Lee, M. Y. (1995). Isolation of chitin from crab shells waste. *Journal Korean Soc. Food Nutrition*, 24(1), 105-113.
- No, H. K., Meyers, S. P., & Lee, K. S. (1989). Isolation and characterization of chitin from crawfish shell waste. *Journal of Agricultural and Food Chemistry*, 37(3), 575-579.
- Ohta, K., Tanguchi, A., Konishi, N., & Hosoki, T. (1999). Chitosan treatment affects plant growth and flower quality in *Eustoma grandiflorum*. *HortScience*, 34, 233-234.
- Okaraonye, C. C., & Ikewuchi, J. C. (2009). Nutritional potential of *Oryctes rhinoceros* larva. *Pakistan Journal of Nutrition*, 8(1), 35-38.
- O'Toole, M. G., Henderson, R. M., Soucy, P. A., Fasciotto, B. H., Hoblitzell, P. J., Keynton, R. S., Gobin, A. S. (2012). Curcumin encapsulation in

- submicrometer spray-dried chitosan/tween 20 particles. *Biomacromolecules*, 13, 2309-2314.
- Packer, L. (1990). Methods in enzymology. Vol 90. New York: Academic press.
- Pădurețu, C.-C., Isopescu, R., Rău, I., Apetroaei, M. R., & Schröder, V. (2019). Influence of the parameters of chitin deacetylation process on the chitosan obtained from crab shell waste. *Korean Journal of Chemical Engineering*, 36(11), 1890-1899.
- Pandey, A., Singh, D., Dhas, N., Tewari, A. K., Pathak, K., Chatap, V., .Mutalik, S. (2020). Complex injectables: development, delivery, and advancement. In *Delivery of Drugs. Volume 2: Expectations and Realities of Multifunctional Drug Delivery Systems* (pp. 191-213). Elsevier Inc.
- Park, B. K., & Kim, M.-M. (2010). Applications of chitin and its derivatives in biological medicine. *International Journal of Molecular Sciences*, 11(12), 5152-5164.
- Park, C.-E., Park, D.-J., Kim, B.-K. (2015) Effects of a chitosan coating on properties of retinol-encapsulated zein nanoparticles. *Food Science and Biotechnology*, 24, 1725–1733.
- Paulino, A., Simionato, J. I., Garcia, J. C., & Nozaki, J. (2006). Characterization of chitosan and chitin produced by silkworm crysalides. *Carbohydrate Polymers*, 94, 98-103.
- Pavlichko, J. P. (1990). Polymer interactions to enhance the function of hyaluronic acid. *Drug and Cosmetics International Journal*, 147, 12-24.
- Peppas, N. A., Bures, P., Leobandung, W., & Ichikawa, H. (2000). Hydrogels in pharmaceutical formulations. *European Journal of Pharmaceutics and Biopharmaceutics*, 50, 27-46.
- Percot, A., Viton, C., & Domard, A. (2003). Optimization of chitin extraction from shrimp shells. *Biomacromolecules*, 4, 12-18.
- Perrault, S. D., & Chan, W. C. (2010). In vivo assembly of nanoparticle components to improve targeted cancer imaging. *Proceedings of the National Academy of Sciences*, 107(25), 11194-11199.
- Pilai, K. S., Paul, W., & Sharma, C. P. (2009). Chitin and chitosan polymers: Chemistry, solubility and fiber formation. *Progress in Polymer Science*, 34, 641-678.

- Pratiwi, R., & Prinajati, D. P. (2018). Adsorption for lead removal by chitosan from shrimp shells. *Indonesian Journal of Urban and Environmental Technology*, 2(1), 35-46.
- Preetha, A., Kunnumakkara, A. B., Newman, R. A., & Aggarwal, B. B. (2007). Bioavailability of curcumin: Problems and Promises. *Molecular Pharmaceutics*, 4(6), 807-818.
- Pu, F., Chen, N., & Xue, S. (2016). Calcium intake, calcium homeostasis and health. *Food Science and Human Wellness*, 5(1), 8-16.
- Puvvada, Y. S., Vankayalapati, S., & Sukhavasi, S. (2012). Extraction of chitin from chitosan from exoskeleton of shrimp for application in the pharmaceutical industry. *International Current Pharmaceutical Journal*, 1(9), 258-263.
- Rai, D., Singh, J. K., Roy, N., & Pandan, D. (2008). Curcumin inhibits FtsZ assembly: an attractive mechanism for its antibacterial activity. *Biochemical Journal*, 410(1), 147-155.
- Ramasamy, P., Subhapradha, N., Shanmugam, V., & Shanmugam, A. (2014). Extraction, characterization and antioxidant property of chitosan from cuttlebone *Sepia Kobiensis* (Hoyle 1885). *International Journal of Biological Macromolecules*, 64, 202-212.
- Ramasamy, R. P., & Maliyekkal, S. M. (2014). Formation of gold nanoparticles upon chitosan leading to formation and collapse of gels. *New Journal of Chemistry*, 1-7.
- Ramírez, M. A., Rodriguez, A. T., Alfonso, L., & Peniche, C. (2010). Chitin and its derivatives as biopolymers with potential agricultural applications. *Biotechnologia Aplicada*, 27, 270-276.
- Rampino, A., Borgogna, M., Blasi, P., Bellich, B., & Cesaro, A. (2013). Chitosan nanoparticles: Preparation, size evolution and stability. *International Journal of Pharmacology*, 455, 219-228.
- Rangkadilok, N., Sitthimonchai, S., Worasuttayangkurn, L., Mahidol, C., Ruchirawut, M., & Satayavivad, J. (2007). Evaluation of free radical scavenging and antityrosinase activities of standardized longan fruit extract. *Food Chemistry and Toxicology*, 45, 328-336.
- Ravindranath, V., & Chandrasekhara, N. (1980). Absorption and tissue distribution of curcumin in rats. *Toxicology*, 16(3), 259-265.

- Ravindranath, V., & Chandrasekhara, N. (1981). *In vitro* studies on the intestinal absorption of curcumin in rats. *Toxicology*, 20(3), 251-257.
- Ravindranath, V., & Chandrasekhara, N. (1981). Metabolism of curcumin-studies with [3H] curcumin. *Toxicology*, 22(4), 337-344.
- Rinaudo, M. (2006). Chitin and chitosan: Properties and applications. *Progress in Polymer Science*, 31(7), 603-623.
- Robert, G. A. (1992). *Chitin chemistry*. London: Macmillan Press Ltd.
- Rodrigues, F., Oliveira, M. B. P. (2016). Cell-based *in vitro* models for dermal permeability studies. In *Concepts and Models for Drug Permeability Studies*. 155-167.
- Rosa, G. S., Moraes, M. A., & Pinto, L. A. (2010). Moisture sorption properties of chitosan. *LWT-Food Science and Technology*, 43, 145-420.
- Ross, A., & Willson, V. L. (2017). One-way anova. In *Basic and advanced statistical tests* (pp. 21-24). Brill Sense.
- Ross, M. B., Mirkin, C. A., & Schatz, G. C. (2016). Optical properties of one-, two-, and three-dimensional arrays of plasmonic nanostructures. *The Journal of Physical Chemistry C*, 120(2), 816-830.
- Saewan, N., Thakam, A., Jintaisong, A., & Kittigowitana, K. (2014). Anti-tyrosinase and cytotoxicity activities of curcumin-metal complexes. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(10), 270-273.
- Sagheer, F. A., Al-Sughayer, M. A., Muslim, S., & Elsabee, M. Z. (2009). Extraction and characterization of chitin and chitosan from marine sources in Arabian Gulf. *Carbohydrate Polymers*, 77, 410-419.
- Sahu, D., Kannan, G. M., Tailang, M., Vijayaraghavan, R. (2016). *In vitro* cytotoxicity of nanoparticles: A comparison between particle size and cell type. *Journal of Nanoscience*. 1-9.
- Sajomsang, W., & Gonil , P. (2010). Preparation and characterization of alpha chitin from cicada sloughs. *Materials Science and Engineering C*, 30, 357-363.
- Sakai, T., & Alexandridis, P. (2005). Mechanism of gold metal ion reduction, nanoparticle growth and size control in aqueous amphiphilic block copolymer solutons at ambient conditions. *The Journal of Physical Chemistry*, 109(16), 7766-7777.

- Samsudin, A., Chew, P. S., & Mohamad, M. M. (1993). *Oryctes rhinoceros*: Breeding and damage on oil palms to oil palm replanting situation. *The Planter*, 69(813), 583-591.
- Sayari, N., Sila, A., Abdelmalek, B. E., Abdallah, R. B., Ellouz-Chaabouni, S., Bougatef, A., & Balti, R. (2016). Chitin and chitosan from the Norway lobster by-products: Antimicrobial and anti-proliferative activities. *International Journal of Biological Macromolecules*, 87, 163-171.
- Schipper, C. M. (1976). Mass rearing the coconut rhinoceros beetle, *Oryctes rhinoceros* L. (Scarab., Dynastinae). *Zeitschrift fur Angewandte Entomologie*, 81(1), 21-25.
- Schipper, N. G., Olsson, S., Hoogstraate, J. A., deBoer, A. G., Varum, K. M., & Artursson, P. (1997). Chitosan as absorption enhancers for poorly absorbable drugs: 2. Mechanism of absorption enhancement. *Pharmaceutical Research*, 14, 923-929.
- Schipper, N., Varum, K. M., & Artursson, P. (1996). Chitosans as absorption enhancers for poorly absorbable drugs: 1. Influence of molecular weight and degree of deacetylation on drug transport across human intestinal epithelial (Caco-2) cells. *Pharmaceutical Research*, 13, 1686-1692.
- Schutz, C. A., Juillerat-Jeanneret, L., Kauper, P., & Wandrey, C. (2011). Cell response to the exposure to chitosan-TPP//alginate nanogels. *Biomacromolecules*, 12, 4153-4161.
- Scimeca, M., Bischetti, S., Lamsira, H. K., Bonfiglio, R., & Bonanno, E. (2018). Energy Dispersive X-ray (EDX) microanalysis: A powerful tool in biomedical research and diagnosis. *European Journal of Histochemistry*, 62(1), 2841.
- Shafiqah, A. R., Abdul Aziz, A., & Mehrdel, B. (2018). Nanoparticle optical properties: Size dependence of a single gold spherical nanoparticle. *Journal of Physics: Conference Series*, 1083, 012040.
- Shah, B. R., Li, Y., Jin, W., An, Y., He, L., Li, Z. H., Li, B. (2016). Preparation and optimization of Pickering emulsion stabilized by chitosan-tripolyphosphate nanoparticles for curcumin encapsulation. *Food Hydrocolloids*, 52, 369-377.
- Shah, P., Jogani, V., Mishra, P., Mishra, A. K., Bagchi, T., & Misra, A. (2007). Modulation of ganciclovir intestinal absorption in presence of absorption enhancers. *Journal of Pharmaceutical Science*, 96, 2710-2722.

- Shahidi, F., & Abuzaytoun, R. (2005). Chitin, chitosan, and co-products: Chemistry, production, applications, and health effects. *Advance Food Nutrition Research*, 49, 93-135.
- Shahidi, F., & Naczk, M. (1995). Method of analysis and quantification of phenolic compound. In *Food phenolics: sources, chemistry, effects and applications* (pp. 287-293). Lanchester, PA. USA: Technomic Publishing Company.
- Shin, C.-S., Kim, D.-Y., & Shin, W.-S. (2019). Characterization of chitosan extracted from mealworm beetle (*Tenebrio molitor*, *Zophobas morio*) and rhinoceros beetle (*Allomyrina dichotoma*) and their antibacterial activities. *International Journal of Biological Macromolecules*, 125, 72-77.
- Shishodia, S., Sethi, G., & Aggarwal, B. B. (2005). Curcumin: getting back to the roots. *Annals of the New York Academy of Sciences*, 1056, 206-217.
- Silva, F. F., Dore, C. G., Marques, C. T., Nascimento, M. S., Benevides, N. B., Rocha, H. O., Leite, E. L. (2010). Anticoagulant activity, paw edema and pleurisy induced carrageenan: Action of major types of commercial carrageenans. *Carbohydrate Polymers*, 79, 26-33.
- Singh, A., Benjakul, S., & Prodpran, T. (2019). Ultrasound-assisted extraction of chitosan from squid pen: Molecular characterization and fat binding capacity. *Journal of Food Science*, 84(2), 224-234.
- Solati, E., & Dorranian, D. (2015). Comparison between silver and gold nanoparticles prepared by pulsed laser ablation in distilled water. *Journal of Cluster Science*, 26(3), 727-742.
- Song, C., Yu, H., Zhang, M., Yang, Y., & Zhang, G. (2013). Physicochemical properties and antioxidant activity of chitosan from the blowfly *Chrysomya megacephala* larvae. *International Journal of Biological Macromolecules*, 60, 347-354.
- Soon, C. Y., Tee, Y. B., Tan, C. H., Rosnita, A. T., & Khalina, A. (2018). Extraction and physicochemical characterization of chitin and chitosan from *Zophobas morio* larvae in varying sodium hydroxide concentration. *International Journal of Biological Macromolecules*, 108, 135-142.
- Stankiewicz, B. A., van Bergen, P. F., Duncan, I. J., Carter, J., Briggs, D., & Evershed, R. P. (1996). Chemical composition of paleozoic and mesozoic fossil invertebrate cuticles as revealed by Pyrolysis-Gas

- Chromatography/Mass Spectrometry. *Rapid Communications in Mass Spectrometry*, 10, 1747-1757.
- Stuchinskaya, T., Moreno, M., Cook, M. J., Edwards, D. R., & Russell, D. A. (2011). Targeted photodynamic therapy of breast cancer cells using antibody-phthalocyanine-gold nanoparticle conjugates. *Photochemical & Photobiological Sciences*, 10, 822.
- Sun, P., Huang, W., Jin, M., Wang, Q., Fan, B., Kang, L., & Gao, Z. (2016). Chitosan-based nanoparticles for survivin targeted siRNA delivery in breast tumor therapy and preventing its metastasis. *International Journal of Nanomedicine*, 11, 4931-4945.
- Sun, T., Yao, Q., Zhou, D., & Mao, F. (2008). Antioxidant activity of N-carbozymethyl chitosan oligosaccharides. *Bioorganic & Medical Chemistry Letters*, 18(21), 5774-5776.
- Synowiecki, J., & Abdul Quawi Al-Khateeb, N. A. (2000). The recovery of protein hydrolysate during enzymatic isolation of chitin from shrimp *Crangon crangon* processing discards. *Food Chemistry*, 68(2), 147-152.
- Tada, T., Ohnishi, K., Suzuki, K., Tomita, H., Okamori, M., Katuzaki, H., Imai, K. (2002). Potential cosmetic whitening agents from insect cuticle: Tyrosinase inhibitory activity of N-acetyldopamine dimers from exuviae of cicada, *Cryptotympana tustulata* FABR. *Journal of Oleo Science*, 51(5), 355-358.
- Tettey, C. O., Nagajyothi, P. C., Lee, S. E., Ocloo, A., Minh An, T. N., Sreekanth, T. M., & Lee, K. D. (2012). Anti-melanoma, tyrosinase inhibitory and anti-microbial activities of gold nanoparticles synthesised from aqueous leaf extracts of *Teraxacum officinale*. *International Journal of Cosmetic Science*, 34, 150-154.
- Thompson, D. T. (2007). Using gold nanoparticles for catalysis. *Nanotoday*, 2(4), 40-43.
- Tiwari, A. D., Mishra, A. K., Mishra, S. B., Arotiba, O. A., & Mamba, B. B. (2011). Green synthesis and stabilization of gold nanoparticles in chemically modified chitosan matrices. *International Journal of Biological Macromolecules*, 48, 682-687.
- Tiyaboonchai, W., Tungpradit, W., & Plianbangchang, P. (2007). Formulation and characterization of curcuminoids loaded solid lipid nanoparticles. *International Journal of Pharmacology*, 337(1-2), 299-306.

- Tolaimate, A., Desbrieres, J., Rhazi, M., & Alagui, A. (2003). Contribution to the preparation of chitins and chitosans with controlled physico-chemical properties. *Polymer*, 44, 7939-7952.
- Trapani, A., Lopedota, A., Franco, M., Cioffi, N., Ieva, E., Garcia-Fuentes, M., & Alonso, M. J. (2010). A comparative study of chitosan and chitosan/cyclodextrin nanoparticles as potential carriers for the oral delivery of small peptides. *European Journal of Pharmacy and Biopharmacy*, 75, 26-32.
- Triunfo, M., Tafi, E., Guarneri, A., Scieuzzo, C., Hahn, T., Zibek, S., Falabella, P. (2021). Insect chitin-based nanomaterials for innovative cosmetics and cosmeceuticals cosmetics. *Cosmetics*, 8(2), 40.
- Tu, C. X., Lin, M., Lu, S. S., Qi, X. Y., Zhang, R. X., & Zhang, Y. Y. (2012). Curcumin inhibits melanogenesis in human melanocytes. *Phytotherapy Research*, 26(2), 174-179.
- Tyagi, P., Singh, M., Kumari, H., & Mukhopadhyay, K. (2015). Bacterial activity of curcumin I is associated with damaging of bacterial membrane. *PLoS One*, 10(3), e0121313.
- Tzoumaki, M. V., Moschakis, T., Kiosseoglou, V., & Biliaderis, C. G. (2011). Oil-in-water emulsions stabilized by chitin nanocrystal particles. *Food Hydrocolloids*, 25, 1521-1529.
- Tzoumaki, M. V., Moschakis, T., Scholten, E., & Biliaderis, C. G. (2012). *In vitro* lipid digestion of chitin nanocrystal stabilized o/w emulsions. *Food & Function*.
- Vakili, T., Iranshahi, M., Arab, H., Riahi, B., Roshan, M., & Kairimi, G. (2017). Safety evaluation of auraptene in rats in acute and subacute toxicity studies. *Regulatory Toxicology and Pharmacology*, 91, 159-164.
- van der Kaaden, A., Boon, J. J., de Leeuw, J. W., de Lange, F., Schuyl, P., Schulten, H. R., & Bahr, U. (1984). Comparison of analytical pyrolysis techniques in the characterization of chitin. *Analytical Chemistry*, 56, 2160.
- Vijayan, S. R., Santhiyagu, P., Singamuthu, M., Kumari, A., Jayaraman, R., & Ethiraj, K. (2014). Synthesis and characterization of silver and gold nanoparticles using aqueous extract of seaweed, *Turbinaria conoides*, and their antimicrofouling activity. *The Scientific World Journal*, 938272.

- Vino, A. B., Ramasamy, P., Shanmugam, V., & Shanmugam, A. (2012). Extraction, characterization and in vitro antioxidative potential of chitosan and sulfated chitosan from Cuttlebone of *Sepia aculeata* Orbigny, 1848. *Asian Pacific Journal of Tropical Biomedicine*, 2, S334-S341.
- Wan, S., Sun, Y., Qi, X., & Tan, F. (2012). Improved bioavailability of poorly water-soluble drug curcumin in cellulose acetate solid dispersion. *Journal of the American Association of Pharmaceutical Scientists*, 13, 159-166.
- Wang, C., Mathiyalagan, R., Kim, Y., Aceituno, V. C., Singh, A., Ahn, S., Yang, D. C. (2016). Rapid green synthesis of silver and gold nanoparticles using *Dendropanax morbifera* leaf extract and their anticancer activities. *International Journal of Nanomedicine*, 11, 3691-3701.
- Wang, R., Billone, P. S., & Mullet, W. M. (2013). Nanomedicine in action: An overview of cancer nanomedicine on the market and in clinical trials. *Journal of Nanomaterials*, 1-12.
- Wang, W., Bo, S., Li, S., & Qin, W. (1991). Determination of the Mark-Houwink equation for chitosans with different degrees of deacetylation. *International Journal of Biology and Macromolecules*, 13, 281-285.
- Water, J. J., Kim, Y., Maltesen, M. J., Franzyk, H., Foged, C., & Nielsen, H. M. (2015). Hyaluronic acid-based nanogels produced by microfluidics-facilitated self-assembly improves the safety profile of the cationic host defense peptide novicidin. *Pharmacology Research*, 32, 2727-2735.
- Wheat, P. F. (2001). History and development of antimicrobial susceptibility testing methodology. *Journal of Antimicrobial Chemotherapy*, 48(Supplement 1), 1-4.
- Wojnarowska-Nowak, R., Polit, J., Broda, D., & Sheregii, E. M. (2015). Gold nanoparticles like a matrix for covalent immobilization of cholesterol oxidase – application for biosensing. *Archive of Metallurgy and Materials*, 60(3), 2289-2296.
- Wolkers, W. F., Oliver, A. E., Tablina, F., & Crowea, J. H. (2004). A Fourier-transform infrared spectroscopy study of sugar glasses. *Carbohydrate Research*, 339, 1077-1082.
- Wood, B. J. (1968a). *Pest of oil palms in Malaysia and their control*. Kuala Lumpur, Malaysia: Incorporated Society of Planters.

- Wood, B. J. (1968b). Studies on the effect of ground vegetation on infestations of *Oryctes rhinoceros* (L.) (Col., Dynastidae) in oyung oil palm replantings in Malaysia. *Buletin of Entomological Research*, 59, 85-96.
- Wood, B. J., Corley, R. H., & Goh, K. H. (1973). Studies on the effect of pest damage on oil palm yield. In: Wastie RL, Earp DA, eds. Advances in oil palm cultivation. Kuala Lumpur, Malaysia: Incorporated society of planters. 360-379.
- Xu, H., Yang, X., & Cai, J. (2020). A novel approach of curcumin loaded chitosan/dextran nanocomposite for the management of complicated abdominal wound dehiscence. *Journal of Cluster Science*, 31, 823-830.
- Yadav, A., Lomash, V., Samim, M., & Flora, S. J. (2012). Curcumin encapsulated in chitosan nanoparticles: A novel strategy for the treatment of arsenic toxicity. *Chemico-Biological Interactions*, 199(1), 49-61.
- Yang, B.-Y., Hu, C.-H., Huang, W.-C., Ho, C.-Y., Yao, C.-H., & Huang, C.-H. (2019). Effect of bilayer nanofibrous scaffolds containing curcumin/lithospermi radix extract on wound healing in streptozotocin-induced diabetic rats. *Polymers*, 11(1745), 1-17.
- Yoshinori, K., Hiraku, O., & Yoshiharu, M. (2003). Application of chitin and chitosan derivatives in the pharmaceutical field. *Current Pharmaceutical Biotechnology*, 4(5), 303-309.
- Younes, I., Ghorbel-Bellaaj, O., Chaabouni, M., Rinaudo, M., Souard, F., Vanhaverbeke, C., Nasri, M. (2014). Use of a fractional factorial design to study the effects of experimental factors on the chitin deacetylation. *International Journal of Biological Macromolecules*, 70, 385-390.
- Younes, I., Hajji, S., Franchet, V., Rinaudo, M., Jellouli, K., & Nasri, M. (2014). Chitin extraction from shrimp shell using enzymatic treatment. Antitumor, antioxidant and antimicrobial activities of chitosan. *International Journal of Biological Macromolecules*, 69, 489-498.
- Young, E. C. (1986). The rhinoceros beetle project: History and review of the research programme. *Agriculture, Ecosystems and Environment*, 15, 149-166.
- Yukuyama, M. N., Kato, E. M., Lobenberg, R., & Bou-Chakra, N. A. (2017). Chanlanges and future prospects of nanoemulsion as a drug delivery system. *Current Pharmaceutical Design*, 23, 495-508.

- Zainol Abidin, N., Kormin, F., Zainol Abidin, N., Mohamed Anuar, N., & Abu Bakar, M. (2020). The potential of insects as alternative sources of chitin: An overview on the chemical method of extraction from various sources. *International Journal of Molecular Sciences*, 21(4978).
- Zhang, L., Mazouzi, Y., Salmain, M., Liedberg, B., & Boujday, S. (2020). Anti-body-gold nanoparticle bioconjugates for biosensors: Synthesis, characterization and selected applications. *Biosensors and Bioelectronics*, 165, 112370.
- Zhang, M., Haga, A., Sekiguchi, H., & Hirano, S. (2000). Structure of insect chitin isolated from beetle larva cuticle and silkworm (*Bombyx mori*) pupa exuvia. *International Journal of Biological Macromolecules*, 40, 99-105.
- Zhang, W., Zhang, J., Jiang, Q., & Xia, W. (2012). Physicochrmical and structural characteristics of chitosan nanopowders prepared by ultrafine milling. *Carbohydrate Polymers*, 87, 309-313.
- Zhang, Y., Jiang, J., Liu, L., Zheng, K., Yu, S., & Fan, Y. (2015). Preparation, assessment, and comparison of α -chitin nano-fiber films with different surface charges. *Nanoscale Research Letters*, 10(26), 1-11.
- Zhang, Y., Xue, C., Xue, Y., Gao, R., & Zhang, X. (2005). Determination of the degree of deacetylation of chitin and chitosan by X-ray powder diffraction. *Carbohydrate Research*, 340, 1914-1917.
- Zhang, Y., Zhang, X., Ding, R., Zhang, J., & Liu, J. (2011). Determination of the degree of deacetylation of chitosan by potentiometric titration preceded by enzymatic pretreatment. *Carbohydrate Polymers*, 83, 813-817.
- Zhao, L., Du, J., Duan, Y., Zhang, Y., Zhang, H., & Yang, C. (2012). Curcumin loaded mixed micelled composed of Pluronic P123 and F68: preparation, optimization and *in vitro* characterization. *Colloids and Surfaces.*, 97, 101-108.
- Zhou, X., Jiang, T., Wang, L., Yang, H., Zhang, S., & Zhou, P. (2010). Interaction of curcumin with Zn(II) and Cu(II) ions based on experiment and theoretical calculation. *Journal of Molecular Structure*, 984, 316-325.

VITA

Nurul Alyani Binti Zainol Abidin is from Malaysia and was born on February 13th, 1994. She completed her bachelor's degree in science (Food Technology) with honours from Universiti Tun Hussein Onn Malaysia in 2017 with a great CGPA. She was offered a fast-track course for her doctorate studies at UTHM. Straightaway, she started her PhD in the same year she graduated with her bachelor's degree. The area of biopolymers has struck her fancy since then. She studied the extraction method for chitin and chitosan from a rare source, which was *O. rhinoceros*, a pest for oil palm trees. Continuing with the high potential of chitosan from the rhinoceros beetle, she continued to synthesis and characterise the chitosan-curcumin gold nanoparticles (CCG-NP). To support the claim stating that chitosan from pest had great value, she elucidated the cosmeceutical potential of the CCG-NP mixture through *in vitro* drug release, antimicrobial, and anti-tyrosinase studies. To completely determine that the mixture is safe for human contact, cytotoxicity for CCG-NP was studied as well. Throughout her PhD journey, she had completed, assisted, and participated in eight different grants, both internal and national. The fields involved were food science, nutrition, nanotechnology, product development, pharmacology, and nutraceuticals.