

PHYTOCHEMICALS ANALYSIS, ANTIOXIDANT,
AND ANTIBACTERIAL ACTIVITIES OF
Salvinia molesta D.S.Mitch (Salviniaceae)

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For my beloved *abah*, Md. Salleh bin Abdullah,
For my lovely sisters (Nur Asshura & Nur Ashira Solehah)

In the name of Allah, The Most Gracious and The Most Merciful,
This study dedicated to all of you.
May Allah grant these beautiful souls Jannah.



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ABSTRACT

The occurrence of invasive alien aquatic plant species in a habitat is recognized as the second largest threat to biodiversity after habitat loss. *Salvinia molesta* D.S.Mitch is classified as an obnoxious and noxious invasive aquatic plant in Malaysia, and it has been listed as the world's worst invasive alien species in Global Invasive Species Database (GISD) since 2013. The main goal of this study was to gain knowledge on the potential medicinal values of *S. molesta* extracts (aqueous and ethanol). Thus, the quantification of phytochemical contents, antioxidant, and antibacterial activities of *S. molesta* extracts were determined. The aqueous and ethanol extracts of *S. molesta* were prepared through the maceration technique. These extracts then were quantified for their total phenolic, flavonoid, tannin, alkaloid, and saponin contents following the standard methods. Besides, the antioxidant activity of the extracts was evaluated by DPPH, ABTS, and FRAP assays. Moreover, both aqueous and ethanol extracts of *S. molesta* were examined for their antibacterial activity by disc-diffusion, minimum inhibitory concentration (MIC), and minimum bactericidal concentration (MBC) methods against selected bacterial strains, such as *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa*. As a result, higher contents of phytochemicals were recorded in the ethanol extract at tannin followed by phenolic, saponin, flavonoid, and alkaloids. Thereupon, the extract obtained from ethanol exhibited high antioxidant capacity based on DPPH, ABTS, and FRAP assays. Moreover, the ethanol extract expressed high antibacterial activity against *B. cereus*, followed by *E. coli*, *S. aureus*, and *P. aeruginosa* as tested using disc-diffusion method. The MIC and MBC against *S. molesta* extracts also revealed that the ethanol extract can inhibit growth and kill all the bacteria strains tested. The study highlighted that ethanol extract exhibited higher phytochemical content, significant antioxidant, and antibacterial properties than aqueous extract.

ABSTRAK

Kewujudan spesies tumbuhan akuatik asing yang invasif dalam sesebuah habitat diiktiraf sebagai ancaman kedua terbesar kepada kepelbagaian biologi, selepas kehilangan habitat. *Salvinia molesta* D.S.Mitch diklasifikasikan sebagai tumbuhan akuatik invasif yang kurang menyenangkan dan berbahaya di Malaysia, dan ia telah disenaraikan sebagai spesies asing invasif terburuk di dunia dalam *Global Invasive Species Database* (GISD) sejak 2013. Tujuan utama kajian ini adalah untuk mendapatkan maklumat berkaitan potensi perubatan bagi ekstrak akueus dan etanol *S. molesta*. Oleh itu, kandungan fitokimia, aktiviti antioksidan dan antibakteria bagi ekstrak *S. molesta* telah ditentukan. Ekstrak akueus dan etanol *S. molesta* telah dihasilkan melalui teknik maserasi. Jumlah kandungan fenolik, flavonoid, tanin, alkaloid, dan saponin bagi kedua-dua ekstrak dikira secara kuantitatif mengikut kaedah piawai. Selain itu, aktiviti antioksidan ekstrak dinilai melalui ujian DPPH, ABTS, dan FRAP. Di samping itu, aktiviti antibakteria kedua-dua ekstrak akueus dan etanol *S. molesta* juga diuji melalui kaedah resapan cakera, kepekatan perencatan minimum (MIC), dan kepekatan membunuh bakteria minimum (MBC) terhadap beberapa bakteria terpilih, seperti *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli*, dan *Pseudomonas aeruginosa*. Hasil dapatan kandungan fitokimia yang lebih tinggi direkodkan dalam ekstrak etanol pada tanin diikuti oleh fenolik, saponin, flavonoid, dan alkaloid. Ekstrak daripada etanol menunjukkan kapasiti antioksidan yang tinggi berdasarkan ujian DPPH, ABTS, dan FRAP. Selain itu, ekstrak etanol *S. molesta* menunjukkan aktiviti antibakteria tertinggi terhadap *B. cereus*, diikuti oleh *E. coli*, *S. aureus*, dan *P. aeruginosa* apabila diuji menggunakan kaedah resapan cakera. MIC dan MBC terhadap ekstrak *S. molesta* juga mendedahkan bahawa ekstrak etanol boleh menghalang pertumbuhan bakteria pada dan membunuh semua bakteria yang diuji. Kajian ini menunjukkan bahawa ekstrak etanol mempunyai kandungan fitokimia yang tinggi, dan mempamerkan sifat antioksidan dan antibakteria yang ketara berbanding ekstrak akueus.

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LIST OF SYMBOLS AND ABBREVIATIONS

$^{\circ}\text{C}$	- Degree Celsius
%	- Percentage
<i>c</i>	- Concentration
<i>g</i>	- Gram
<i>m</i>	- Mass
<i>mm</i>	- Millimeter
<i>mg/g</i>	- Milligram per gram
<i>V</i>	- Volume
<i>w/v</i>	- Weight per volume
$\mu\text{g/mL}$	- Microgram per milliliter
<i>ABTS</i>	- 2, 2'-azino-bis-3-ethylbenzothiazoline-6-sulfonic acid
<i>BHT</i>	- Butylated hydroxytoluene
<i>CSIRO</i>	- Commonwealth Scientific and Industrial Research Organization of Australia
<i>DNA</i>	- Deoxyribonucleic acid
<i>DPPH</i>	- 2,2-diphenyl-1-picryl-hydrazyl-hydrate
<i>ET</i>	- Electron transfer
<i>EPPO</i>	- European and Mediterranean Plant Protection Organization
<i>FRAP</i>	- Ferric reducing antioxidant power
<i>GAE</i>	- Gallic acid equivalent
<i>GAS</i>	- Group A <i>Streptococcus</i>
<i>GISD</i>	- Global Invasive Species Database
<i>HAT</i>	- Hydrogen atom transfer
<i>IAAPs</i>	- Invasive alien aquatic plants
<i>INT</i>	- p-iodonitrotetrazolium chloride
<i>MBC</i>	- Minimum bactericidal concentration

<i>MHA</i>	-	Mueller-Hinton agar
<i>MHB</i>	-	Mueller-Hinton broth
<i>MIC</i>	-	Minimum inhibition concentration
<i>QE</i>	-	Quercetin equivalent
<i>ROS</i>	-	Reactive oxygen species
<i>RPF</i>	-	Radical protection factor
<i>RSF</i>	-	Radical skin factor
<i>TAC</i>	-	Total alkaloid content
<i>TAE</i>	-	Tannic acid equivalent
<i>TE</i>	-	Trolox equivalent
<i>TFC</i>	-	Total flavonoid content
<i>TPC</i>	-	Total phenolic content
<i>TSC</i>	-	Total saponin content
<i>TTC</i>	-	Total tannin content
<i>UTHM</i>	-	Universiti Tun Hussein Onn Malaysia



PTTA UTHM
PERPUSTAKAAN TUNKU TUN AMINAH

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CHAPTER 1

INTRODUCTION

1.1 Background of the study

Plants have been exploited as medicinal sources since ancient times. The World Health Organization (WHO) in 1993 stated that medicinal plants are not only important in the health care of ancient cultures for thousands of years due to their therapeutic potential but also in the current modern culture. Most consumers rely on herbal medications because alternative treatment options are more expensive and are frequently accompanied by dangerous negative effects (Mukherjee & Wahile, 2006). Accordingly, the use of natural plant products has been in greater demand nowadays and, as a result, there is a constant and urgent need to create new pharmaceuticals with a variety of chemical structures as well as brand-new mechanisms of action for infectious diseases that are currently emerging (Barbosa, Rall, & Fernandes, 2009).

Invasive alien aquatic plants (IAAPs) are non-native aquatic plant species that grow faster than native species, even in low-resource habitats (Jo, Fridley, & Frank, 2017). These plants are distinguished by their high level of adaptation in the absence of common predators, rapid proliferation, and phenotypic plasticity (Ismail *et al.*, 2019). Besides, other variables that contribute to the establishment and spread of IAAPs include increased nutrient levels in water and changes in flow regimes (Keller, Masoodi, & Shackleton, 2018). They have a variety of effects on the socioeconomic system, freshwater ecology, and water quality (Hassan & Nawchoo, 2020). The IAAPs may obstruct fishing activity and water transportation by growing dense mats on the surface, blocking streams, limiting sunlight, and outcompeting native floating plants for nutrients. In addition, the occurrence of IAAPS in a habitat is recognized as the second largest threat to biodiversity after habitat loss (Ismail *et al.*, 2019).

The rising of global trade and cross-border activity has enabled species to be transported to many parts of the world, including Malaysia and Indonesia. Both countries provide areas in which living organisms grow and prosper in humid and warm tropical regions. Some species colonize and naturalize well in their surroundings, while others become invasive (Ismail *et al.*, 2019). In Malaysia, some of the aquatic plants discovered are on the list of the world's 100 worst invasive alien species (Jo *et al.*, 2017), including water hyacinth (*Eichhornia crassipes*) and water fern (*Salvinia molesta*), and a few more that have yet to be named. As such, the introduction of these species would cause significant dangers to native species because they would face intense competition for resources, space, disease, and other factors (Ismail *et al.*, 2019).

Despite being a nuisance to the ecosystem, this aquatic weed has enormous untapped medicinal potential that could make them effective therapeutic agent for a variety of developing diseases. Moreover, there is a growing global interest to discover the medicinal benefits of aquatic weeds (Nithya, Jayanthi, & Raghunathan, 2016). Reactive oxygen species (ROS) such as hydroxyl radicals, peroxides, and superoxide anions are primary causes of oxidative damage that leads to mutagenesis, aging, carcinogenesis, and cardiovascular diseases (Hajam *et al.*, 2022). Both endogenous or exogenous antioxidants, whether synthetic or natural, protect the human body from oxidative stress-related disorders by inhibiting or delaying the oxidation process by limiting the onset or propagation of oxidizing chain reactions (Pisoschi *et al.*, 2021). Currently, plant-derived natural antioxidants are in high demand due to their numerous health benefits (Manzoor *et al.*, 2023).

Besides, since the introduction of antibacterial agents, they have played a critical role in reducing morbidity from infectious diseases. Nevertheless, infectious diseases continue to be the largest cause of death worldwide, and bacteria have become more resistant to conventional antibiotics in the recent years. The extensive usage of these compounds is expected to hasten the development of antibiotic resistance. In 2014, O'Neill published a high-profile assessment report estimating that antimicrobial resistance (AMR) might kill 10 million people per year by 2050. Accordingly, studies of natural products, with a focus on the isolation, purification, and characterization of secondary metabolites of plants, have has a significant impact on the discovery of substances with antibacterial action, because these metabolites have shown relevant benefits in the treatment of a variety of diseases (Bezerra *et al.*, 2019; Mulat, Pandita, & Khan, 2019).

S. molesta D.S. Mitch is a worldwide aquatic invader, named after Antonio Maria Salvini (1633 – 1729), belonging to the family Salviniaceae. The species name, *molesta*, was derived from the Latin word *molestus*, highlighting its weediness as it is a nuisance across the world, invading natural water bodies as well as artificial impoundments (Julien, Hill, & Tipping, 2009). In Malaysia, this aquatic plant is also known as kiambang *Salvinia* (Baehaki, Herpandi, & Anggraini, 2018; Sidek *et al.*, 2018; Maili, 2017). Based on the Global Invasive Species Database (GISD, 2022), in 2013, *S. molesta* was selected to replace the Rinderpest virus in the world's worst invasive alien species list, as the virus was declared eradicated in the wild in 2010.

Hence, in this study, *S. molesta* extracts, aqueous and ethanol, were examined to quantify the presence of phytochemicals and analyze the antioxidant and antibacterial activities. Understanding and evaluating the positive qualities of *S. molesta* extracts may be the initial step toward the effective exploitation of this plant as a natural alternative strategy for addressing IAAPs challenges in Malaysia. The knowledge of plant chemical contents is desirable for the identification of therapeutic agents as well as the discovery of new sources of economic phytochemicals for the synthesis of complex chemical substances and the actual significance of folklore medicines (Purushoth *et al.*, 2013).

1.2 Problem statement

Taman Botani Johor is a public recreation area where both locals and visitors can relax and stroll around the lakes. *S. molesta*, however, has successfully infested the lakes in recent years and the management was negatively impacted by the thick mats of this invasive aquatic plant. *S. molesta* was manually harvested from the lakes because it was a sustainable practice. Even so, since not all plants are removed, thus allowing the population to be regenerated through vegetative reproduction (Hill & Coetzee, 2017), the harvest requires manpower and regular follow-ups. The biomass of *S. molesta* was left on the edges of the lakes after harvesting. As a result, the aesthetic value of the lakes eventually becomes tarnished, and the biomass is wasted.

According to Alves *et al.* (2021), one of the explanations for a successful, exotic invasive plant species is a 'novel phytochemistry hypothesis'—exotic plants possessing potent secondary compounds that are unique or underrepresented in the plants' new range, making it more likely to develop into highly invasive. In addition,

plant materials for phytochemical studies are often expensive, scarce, and difficult to obtain. As suggested by Fan & Marston (2009), instead of focusing on rare or extinct species, research on the phytochemicals of common invasive species will utilize biomass as a source of potentially essential phytochemical compounds.

However, research and documentation on the phytochemicals and therapeutic values of *S. molesta* remain sparse and insufficient in Malaysia. Studies have focused more on the phytoremediation of this species on wastewater, as reported by Hayder & Mustafa (2021), Mustafa & Hayder (2021), Mohd-Nizam *et al.* (2020), Hanafiah, Mohamad, and Aziz (2018), Sidek *et al.* (2018), Ng, Samsudin, and Chan (2017), Ng & Chan (2017), and Ashraf, Maah, and Yusoff (2013). With the increasing interest in natural products, which may have less toxic and carcinogenic side effects than synthetic ones, researchers seek to assess and identify natural compounds as a green alternative (Johnston *et al.*, 2005). Thus, the study aims to investigate the active compounds and medicinal values of the different extracts of *S. molesta*, which are aqueous and ethanol.

The production of highly ROS by a single unpaired electron causes oxidative stress and is implicated in the pathophysiology of a variety of physiological diseases. These include cellular injury, cancer, cardiovascular, aging, and other problems (Losada-Barreiro & Bravo-Diaz, 2017). Natural antioxidants derived from plant sources have gained significant public attention in recent years due to the negative side effects of synthetic antioxidants such as toxicity and carcinogenicity (Taghvaei & Jafari, 2015). Besides, George & Gabriel (2016) also asserted that plants are possible sources of natural antioxidants that can be used to sabotage ROS. Thus, a study that searches into the antioxidant potential of *S. molesta* is crucial for understanding the ability of the plant to operate as a natural antioxidant.

Microorganisms are the primary reason of acute and chronic diseases, and the regular use of antibiotics increases the number of novel strains with antibiotic resistance. Antibiotic resistance and the need for new antibiotics have emerged as a major concern due to the rising demand (Zada *et al.*, 2021). As a result, the use of plant extracts is increasing. This is because plants contain a high concentration of antibacterial compounds and are being researched as a novel medication for antibiotic-resistant pathogens (Al-Rifai *et al.*, 2017). Hence, the evaluation of *S. molesta* against the resistant bacteria is necessary to better understand the ability of the plant to combat the bacteria.

1.3 Significance of the study

Although aquatic weeds are an environmental annoyance, they also possess a significant untapped medicinal potential that could make them an effective treatment for a variety of emerging diseases (Nithya *et al.*, 2016). Given the growing global interest in discovering the untapped medicinal properties of aquatic weeds, *S. molesta* might also be developed into a new, economically viable plant for medicine (Nithya *et al.*, 2016). Even though there is no citation on the traditional use of *S. molesta* in prior literature, this invasive aquatic plant was reported to have potential antioxidant (Nithya *et al.*, 2016) and antibacterial activities (Baehaki *et al.*, 2018; Nithya, Jayanthi, & Raghunathan, 2015). The findings from this study provide essential opportunities for discovery, which may result in new insights and a valuable option for future demands, especially in medicinal values of an invasive aquatic plant, *S. molesta*.

1.4 Limitations of study

There are few limitations observed during this study. Firstly, the samples of *S. molesta* were collected from the lake of Taman Botani Johor, which was not a controlled environment. The findings in this study could be affected by the external environmental factors. Generally, it is recognized that variety of external factors, including biotic and abiotic stressors, light, air, and substrate composition, can have an impact on the growth, development, uptake of micro- and macronutrients, and accumulation of phytochemicals of the plants (Syakilla *et al.*, 2022; Almuhayawi *et al.*, 2021). Apart from that, only secondary metabolites of aqueous and ethanol extracts of *S. molesta* were investigated in this study. Thus, a solid conclusion on percentage yield could not be derived from the findings. This is because the primary metabolites were not measured. According to Do *et al.* (2014), high-polarity solvents can extract compounds with a wider polarity, which allows primary metabolites, such as proteins and carbohydrates, to dissolve during the extraction process.

1.5 Research objectives

The study aimed to investigate scientifically the potential medicinal values of an invasive aquatic plant, *S. molesta*. This aim could be achieved through following specific objectives:

- i. To determine the phenolic, flavonoid, alkaloid, tannin, and saponin contents of aqueous and ethanol extracts of *S. molesta*.
- ii. To study the antioxidant properties of aqueous and ethanol extracts of *S. molesta* using DPPH, ABTS, and FRAP assays.
- iii. To examine the antibacterial properties of aqueous and ethanol extracts of *S. molesta* using the disc-diffusion method, minimum inhibition concentration (MIC), and minimum bactericidal concentration (MBC) assays.

1.6 Scope of the study

The scopes of the study are as follows:

- i. Taman Botani Johor, Batu Pahat was chosen as the study site for the collection of *S. molesta* samples.
- ii. *S. molesta* samples were extracted using aqueous and ethanol solvents.
- iii. The extracts were analyzed on phytochemical content including phenolics, flavonoids, tannins, alkaloids, and saponins.
- iv. The antioxidant activity of both extracts was measured using free radical scavenging assays.
- v. Four selected bacteria were used for the antibacterial activity test, specifically two Gram-negative: *Escherichia coli* and *Pseudomonas aeruginosa*, and two Gram-positive: *Bacillus cereus* and *Staphylococcus aureus*. Different bacteria were chosen to show the ability of the extracts as an antibacterial agent, using the disc diffusion method, MIC, and MBC assays.

CHAPTER 2

LITERATURE REVIEW

2.1 Aquatic plants

Aquatic plants, also called hydrophytic plants or hydrophytes, are plants that have adapted to or are living in aquatic environments (Muda, 2010). The three main divisions of aquatic plants include bryophytes (mosses, liverworts, and hornworts), pteridophytes (ferns and allies), and spermatophytes (seed-bearing plants) (Table 2.1) (Bornette & Puijalon, 2009; Chambers *et al.*, 2007). These aquatic plants are also classified into emergent, floating leaf, and submerged growth forms (Ismail, 2014; Chambers *et al.*, 2007).

Table 2.1: Aquatic plant divisions and representative genera (Chambers *et al.*, 2007)

Kingdom	Division	Descriptive term	Representative genera
Plantae	Bryophyta	Mosses and liverworts	<i>Fontinalis</i> , <i>Riella</i> , <i>Ricciocarpus</i>
	Pteridophyta	Ferns and allies	<i>Azolla</i> , <i>Salvinia</i> , <i>Isoetes</i>
	Spermatophyta	Seed-bearing plants	<i>Sagittaria</i> , <i>Alisma</i> , <i>Butomus</i> , <i>Brasenia</i> , <i>Cabomba</i> , <i>Callitriche</i> , <i>Ceratophyllum</i> , <i>Scirpus</i> , <i>Carex</i> , <i>Myriophyllum</i> , <i>Ellodea</i> , <i>Vallisneria</i> , <i>Juncus</i> , <i>Lemna</i> , <i>Utricularia</i> , <i>Nelumbo</i> , <i>Nymphaea</i> , <i>Nuphur</i> , <i>Sapartina</i> , <i>Eichhornia</i> , <i>Potamogeton</i> , <i>Ranunculus</i> , <i>Sparganium</i> , <i>Typha</i>

In the early 21st century, researchers have come to understand the critical role of plants that grow in or near water play in the development, maintenance, and provision of aquatic ecosystem services (Chambers *et al.*, 2007). In addition to interacting with and influencing the hydrological, geomorphological, and physio-

chemical environments, aquatic plants also engage in a wide range of interactions with other organisms from microbes to vertebrates, for example, by providing habitat and food (O'Hare *et al.*, 2018; Wood *et al.*, 2017). Aquatic plants are the foundation of aquatic food chains; they also actively contribute to the development and maintenance of freshwater ecosystem services and food webs (Smith, 2011; Scheffer & Jeppesen, 2007). These plants provide ideal habitat for large fish, besides serving as a shelter for the juvenile and food sources for fish, insects, waterfowls, and other wildlife.

All plants, including those that grow underwater, produce oxygen due to photosynthesis; thus, aquatic plants are also the primary source of oxygen for other aquatic life (Bonvechio & Bonvenchio, 2006). Moreover, aquatic plants act as important bioindicators of environmental conditions and long-term ecological changes in water quality (Lacoul & Freedman, 2006; Solimini, Cardoso, & Heiskanen, 2006). Rooted aquatic plants help to stabilize shorelines and bottom sediments. They absorb nutrients and filter pollutants from runoff, resulting in cleaner water. Lembi (2009) added that a diverse aquatic plant population enhances the beauty of the water body. Thus, many people acknowledge and appreciate the aesthetic value of aquatic plant life, whether in a backyard fishpond, around a retention pond, or along the shoreline of a lake.

2.1.1 Invasive alien aquatic plants (IAAPs)

The introduction of non-native aquatic plants to the new habitat has frequently become a nuisance by impeding human water use and endangering the structure and function of diverse native aquatic ecosystems (Richardson *et al.*, 2000). Invasive alien aquatic plants (IAAPs) are defined as non-native aquatic plant species that grow more quickly than native species, even in habitats with limited resources. The rapid growth of invasive plants is associated with a set of plant traits that can boost nutrient uptake and photosynthetic capacity (Jo *et al.*, 2017). According to Gettys (2014), some of the IAAPs are purposely introduced by humans as ornamentals, aquarium decorations, and souvenirs. Some introduced aquatic plant species are colonized and naturalized well to the native environment, while some become invasive. Besides, the invasion of these species in the native habitats was also caused by the expanding global trade and cross-boundary transportation services (Havel *et al.*, 2015).

REFERENCES

- Aboul-Enein, A. M., Shanab, S. M., Shalaby, E. A., Zahran, M. M., Lightfoot, D. A., & El-Shemy, H. A. (2014). Cytotoxic and antioxidant properties of active principals isolated from water hyacinth against four cancer cells lines. *BMC complementary and alternative medicine*, 14(1), 1-11.
- Agu, K. C., & Okolie, P. N. (2017). Proximate composition, phytochemical analysis, and in vitro antioxidant potentials of extracts of *Annona muricata* (Soursop). *Food science & nutrition*, 5(5), 1029-1036.
- Ajanal, M., Gundkalle, M. B., & Nayak, S. U. (2012). Estimation of total alkaloid in Chitrakadivati by UV-Spectrophotometer. *Ancient science of life*, 31(4), 198.
- Al-Rifai, A., Aqel, A., Al-Warhi, T., Wabaidur, S. M., Al-Othman, Z. A., & Badjah-Hadj-Ahmed, A. Y. (2017). Antibacterial, antioxidant activity of ethanolic plant extracts of some Convolvulus species and their DART-ToF-MS profiling. *Evidence-Based Complementary and Alternative Medicine*, 2017.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2002). Genesis, modulation, and regeneration of skeletal muscle. In *Molecular Biology of the Cell*. 4th edition. Garland Science.
- Aleksic, V., & Knezevic, P. (2014). Antimicrobial and antioxidative activity of extracts and essential oils of *Myrtus communis* L. *Microbiological research*, 169(4), 240-254.
- Almuhayawi, S. M., Almuhayawi, M. S., Al Jaouni, S. K., Selim, S., & Hassan, A. H. (2021). Effect of laser light on growth, physiology, accumulation of phytochemicals, and biological activities of sprouts of three Brassica cultivars. *Journal of Agricultural and Food Chemistry*, 69(22), 6240-6250.
- Alves, D., Duarte, S., Arsénio, P., Gonçalves, J., Rodrigues, C. M. P., Lourenço, A., & Máximo, P. (2021). Exploring the Phytochemicals of *Acacia melanoxylon* R. Br. *Plants* 2021, 10, 2698.

- Andama, M., Ongom, R., & Lukubye, B. (2017). Proliferation of *Salvinia molesta* at lake Kyoga landing sites as a result of anthropogenic influences. *Journal of Geoscience and Environment Protection*, 5(11), 160.
- Anokwuru, C. P., Anyasor, G. N., Ajibaye, O., Fakoya, O., & Okebugwu, P. (2011). Effect of extraction solvents on phenolic, flavonoid and antioxidant activities of three Nigerian medicinal plants. *Nature and Science*, 9(7), 53-61.
- Anusiya, G., Bharathi, S., Mukesh Praveen, K., & Sainandhini, G. (2020). Extraction and molecular characterization of biological compounds from water hyacinth. *Journal of Medicinal Plants*, 8(5), 14-19.
- Aquino, R., Morelli, S., Lauro, M. R., Abdo, S., Saija, A., & Tomaino, A. (2001). Phenolic constituents and antioxidant activity of an extract of *Anthurium versicolour* leaves. *Journal of Natural Products*, 64(8), 1019-1023.
- Arawande, J. O., Akinnusotu, A., & Alademeyin, J. O. (2018). Extractive value and phytochemical screening of ginger (*Zingiber officinale*) and turmeric (*Curcuma longa*) using different solvents. *Int. J. Trad. Nat. Med*, 8(1), 13-22.
- Ashraf, M. A., Maah, M. J., & Yusoff, I. (2013). Evaluation of natural phytoremediation process occurring at ex-tin mining catchment. *Chiang Mai J. Sci*, 40(2), 198-213.
- Azalework, H. G., Sahabjada, A. J., & Md Arshad, T. M. (2017). Phytochemical investigation, GC-MS profile and antimicrobial activity of a medicinal plant *Ruta graveolens* L. from Ethiopia. *International Journal of Pharmacy and Pharmaceutical Sciences*, 9(6), 29-34.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., Jahurul, M., H., A., Ghafoor, K., Norulaini, N., A., N., & Omar, A. K. M. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of food engineering*, 117(4), 426-436.
- Baehaki, A., Herpandi, H., & Anggraini, A. (2018). Aktivitas antibakteri ekstrak tumbuhan perairan kiambang (*Salvinia molesta*). *Saintekmol: Jurnal Sains dan Teknologi*, 16(2), 125-134.
- Baehaki, A., Tampubolon, N. H., & Lestari, S. D. (2020). Phytochemical Compounds and Antioxidant Activity of Water Chestnut (*Eleocharis dulcis*) and Giant Molesta (*Salvinia molesta*) Extract. *Systematic Reviews in Pharmacy*, 11(12).
- Bakar, B. H. (2004). Invasive weed species in Malaysian agro ecosystems: species, impacts and management. *Malaysian Journal of Science*, 23(1), 1-42.

- Balasundram, N., Sundram, K., & Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food chemistry*, 99(1), 191-203.
- Bandiola, T. M. B. (2018). Extraction and qualitative phytochemical screening of medicinal plants: A brief summary. *International Journal of Pharmacy*, 8(1), 137-143.
- Barakat, M. Z., Shahab, S. K., Darwin, N., & Zahemy, E. I. (1993). Determination of ascorbic acid from plants. *Anal. Biochem*, 53, 225-245.
- Baral, B., & Vaidya, G. S. (2011). Biological and chemical assessment of water hyacinth (*Eichhornia crassipes* (mart.) Solms.) of Phewa Lake, Nepal. *Scientific World*, 9(9), 57-62.
- Baral, B., Vaidya, G. S., and Bhattarai, N. (2012). Bioactivity and Biochemical Analysis of Water Hyacinth (*Eichhornia crassipes*). *Botanica Orientalis*, 8, 33-39.
- Barbosa, L. N., Rall, V. L. M., Fernandes, A. A. H., Ushimaru, P. I., da Silva Probst, I., & Fernandes Jr, A. (2009). Essential oils against foodborne pathogens and spoilage bacteria in minced meat. *Foodborne pathogens and disease*, 6(6), 725-728.
- Bendary, E., Francis, R. R., Ali, H. M. G., Sarwat, M. I., & El Hady, S. (2013). Antioxidant and structure-activity relationships (SARs) of some phenolic and anilines compounds. *Annals of Agricultural Sciences*, 58(2), 173-181.
- Benzie, I. F., & Strain, J. J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. *Analytical biochemistry*, 239(1), 70-76.
- Bezerra, J. W. A., Costa, A. R., de Freitas, M. A., Rodrigues, F. C., de Souza, M. A., da Silva, A. R. P., Dos Santos, A. T. L., Linhares, K. V., & Morais-Braga, M. F. B. (2019). Chemical composition, antimicrobial, modulator and antioxidant activity of essential oil of *Dysphania ambrosioides* (L.) Mosyakin & Clemants. *Comparative Immunology, Microbiology and Infectious Diseases*, 65, 58-64.
- Bhuyar, P., Rahim, M. H., Sundararaju, S., Maniam, G. P., & Govindan, N. (2020). Antioxidant and antibacterial activity of red seaweed *Kappaphycus alvarezii* against pathogenic bacteria. *Global Journal of Environmental Science and Management*, 6(1), 47-58.

- Birt, D. F., & Jeffery, E. (2013). Flavonoids. *Advances in Nutrition*, 4(5), 576-577.
- Bolou, G. E. K., Bagré, I., Ouattara, K., & Djaman, A. J. (2011). Evaluation of the antibacterial activity of 14 medicinal plants in Côte d'Ivoire. *Tropical Journal of Pharmaceutical Research*, 10(3).
- Bonvechio, K. I., & Bonvechio, T. F. (2006). Relationship between habitat and sport fish populations over a 20-year period at West Lake Tohopekaliga, Florida. *North American Journal of Fisheries Management*, 26(1), 124-133.
- Booy, O., Wade, M., & Roy, H. (2015). *Field guide to invasive plants and animals in Britain*. Bloomsbury Publishing.
- Bornette, G., & Puijalon, S. (2009). Macrophytes: ecology of aquatic plants. *eLS*.
- Bravo, L. (1998). Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition reviews*, 56(11), 317-333.
- Broadhurst, R. B., & Jones, W. T. (1978). Analysis of condensed tannins using acidified vanillin. *Journal of the Science of Food and Agriculture*, 29(9), 788-794.
- Burri, S. C., Ekholm, A., Håkansson, Å., Tornberg, E., & Rumpunen, K. (2017). Antioxidant capacity and major phenol compounds of horticultural plant materials not usually used. *Journal of functional foods*, 38, 119-127.
- Cai, Y., Luo, Q., Sun, M., & Corke, H. (2004). Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life sciences*, 74(17), 2157-2184.
- Carocho, M., & Ferreira, I. C. (2013). A review on antioxidants, prooxidants and related controversy: Natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food and chemical toxicology*, 51, 15-25.
- Cary, P. R., & Weerts, P. G. (1983). Growth of *Salvinia molesta* as affected by water temperature and nutrition II. Effects of phosphorus level. *Aquatic botany*, 17(1), 61-70.
- Chambers, P. A., Lacoul, P., Murphy, K. J., & Thomaz, S. M. (2007). Global diversity of aquatic macrophytes in freshwater. In *Freshwater animal diversity assessment* (pp. 9-26). Springer, Dordrecht.
- Chandrasekaran, A., Idelchik, M. D. P. S., & Melendez, J. A. (2017). Redox control of senescence and age-related disease. *Redox biology*, 11, 91-102.

- Chang, C. C., Yang, M. H., Wen, H. M., & Chern, J. C. (2002). Estimation of total flavonoid content in propolis by two complementary colourimetric methods. *Journal of food and drug analysis*, 10(3).
- Chantiratikul, P., Meechai, P., & Nakbanpotec, W. (2009). Antioxidant activities and phenolic contents of extracts from *Salvinia molesta* and *Eichhornia crassipes*. *Research Journal of Biological Sciences*, 4(10), 1113-1117.
- Chao, P. Y., Lin, S. Y., Lin, K. H., Liu, Y. F., Hsu, J. I., Yang, C. M., & Lai, J. Y. (2014). Antioxidant activity in extracts of 27 indigenous Taiwanese vegetables. *Nutrients*, 6(5), 2115-2130.
- Chemat, F., Abert Vian, M., Ravi, H. K., Khadhraoui, B., Hilali, S., Perino, S., & Fabiano Tixier, A. S. (2019). Review of alternative solvents for green extraction of food and natural products: Panorama, principles, applications and prospects. *Molecules*, 24(16), 3007.
- Chikezie, P. C., Ibegbulem, C. O., & Mbagwu, F. N. (2015). Medicinal potentials and toxicity concerns of bioactive principles. *Med Aromat Plants*, 4(3), 1-15.
- Choudhary, M. I., Naheed, N., Abbaskhan, A., Musharraf, S. G., & Siddiqui, H. (2008). Phenolic and other constituents of freshwater fern *Salvinia molesta*. *Phytochemistry*, 69(4), 1018-1023.
- Cilliers, C. J. (1991). Biological control of water hyacinth, *Eichhornia crassipes* (Pontederiaceae), in South Africa. *Agriculture, ecosystems & environment*, 37(1-3), 207-217.
- Cueva, C., Moreno-Arribas, M. V., Martín-Álvarez, P. J., Bills, G., Vicente, M. F., Basilio, A., ... & Bartolomé, B. (2010). Antimicrobial activity of phenolic acids against commensal, probiotic and pathogenic bacteria. *Research in microbiology*, 161(5), 372-382.
- Cushnie, T. T., Cushnie, B., & Lamb, A. J. (2014). Alkaloids: An overview of their antibacterial, antibiotic-enhancing and antivirulence activities. *International journal of antimicrobial agents*, 44(5), 377-386.
- de Almeida, C. G., Garbois, G. D., Amaral, L. M., Diniz, C. C., & Le Hyaric, M. (2010). Relationship between structure and antibacterial activity of lipophilic N-acyldiamines. *Biomedicine & pharmacotherapy*, 64(4), 287-290.
- De Bruyne, T., Pieters, L., Deelstra, H., & Vlietinck, A. (1999). Condensed vegetable tannins: biodiversity in structure and biological activities. *Biochemical Systematics and Ecology*, 27(4), 445-459.

- Diop, O., & Hill, M. P. (2009). Quantitative post-release evaluation of biological control of floating fern, *Salvinia molesta* DS Mitchell (Salviniaceae), with *Cyrtobagous salviniae* Calder and Sands (Coleoptera: Curculionidae) on the Senegal River and Senegal River Delta. *African Entomology*, 17(1), 64-70.
- Djeridane, A., Yousfi, M., Nadjemi, B., Boutassouna, D., Stocker, P., & Vidal, N. (2006). Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. *Food chemistry*, 97(4), 654-660.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of food and drug analysis*, 22(3), 296-302.
- Doeleman, J. A. (1989). *Biological control of Salvinia molesta in Sri Lanka: an assessment of costs and benefits* (No. 436-2016-33783).
- Dorman, H. J. D., Peltoketo, A., Hiltunen, R., & Tikkanen, M. J. (2003). Characterisation of the antioxidant properties of de-odourised aqueous extracts from selected Lamiaceae herbs. *Food chemistry*, 83(2), 255-262.
- Dudonne, S., Vitrac, X., Coutiere, P., Woillez, M., & Mérillon, J. M. (2009). Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *Journal of agricultural and food chemistry*, 57(5), 1768-1774.
- Ekam, V. S., & Ebong, P. E. (2007). Serum protein and enzyme levels in rats following administration of antioxidant vitamins during caffeinated and non-caffeinated paracetamol induced hepatotoxicity. *Nigerian Journal of Physiological Sciences*, 22(1-2).
- El Far, M. M., & Taie, H. A. (2009). Antioxidant activities, total anthocyanins, phenolics and flavonoids contents of some sweetpotato genotypes under stress of different concentrations of sucrose and sorbitol. *Australian Journal of Basic and Applied Sciences*, 3(4), 3609-3616.
- Elumalai, A., & Eswaraiah, M. C. (2011). A pharmacological review on *Garcinia indica* Choisy. *Int. J. Univers. Pharm. Life Sci*, 1, 508-206.
- Elvira, K., Fachriyah, E., & Kusriani, D. (2018). Isolation of Flavonoid Compounds from Eceng Gondok (*Eichhornia crassipes*) and Antioxidant Tests with DPPH (1, 1-Diphenyl-2-Picrylhydrazyl) Method. *Jurnal Kimia Sains dan Aplikasi*, 21(4), 187-192.

- Fan, P., & Marston, A. (2009). How can phytochemists benefit from invasive plants?. *Natural Product Communications*, 4(10), 1934578X0900401018.
- Fawole, F. J., Sahu, N. P., Pal, A. K., & Lakra, W. S. (2013). Evaluation of antioxidant and antimicrobial properties of selected Indian medicinal plants. *International Journal of Medicinal and Aromatic Plants*, 3(1), 69-77.
- Fellegrini, N., Ke, R., Yang, M., & Rice-Evans, C. (1999). [34] Screening of dietary carotenoids and carotenoid-rich fruit extracts for antioxidant activities applying 2, 2'-azinobis (3-ethylenebenzothiazoline-6-sulfonic acid radical cation decolourization assay. In *Methods in enzymology* (Vol. 299, pp. 379-389). Academic Press.
- Fernandez-Panchon, M. S., Villano, D., Troncoso, A. M., & Garcia-Parrilla, M. C. (2008). Antioxidant activity of phenolic compounds: from in vitro results to in vivo evidence. *Critical reviews in food science and nutrition*, 48(7), 649-671.
- Ferrer, J. L., Austin, M. B., Stewart Jr, C., & Noel, J. P. (2008). Structure and function of enzymes involved in the biosynthesis of phenylpropanoids. *Plant Physiology and Biochemistry*, 46(3), 356-370.
- Forno, I. W. (1983). Native distribution of the *Salvinia auriculata* complex and keys to species identification. *Aquatic Botany*, 17(1), 71-83.
- Forno, I. W., & Bourne, A. S. (1984). Studies in South America of arthropods on the *Salvinia auriculata* complex of floating ferns and their effects on *S. molesta*. *Bulletin of entomological research*, 74(4), 609-621.
- Forno, I. W., & Harley, K. L. S. (1979). The occurrence of *Salvinia molesta* in Brazil. *Aquatic Botany*, 6(2), 185-187.
- Friedman, M. (2006). Antibiotic activities of plant compounds against non-resistant and antibiotic-resistant foodborne human pathogens.
- Frijhoff, J., Winyard, P. G., Zarkovic, N., Davies, S. S., Stocker, R., Cheng, D., Knight, A., R., Taylor, E., L., Oettrich, J., Ruskovska, T., Gasparovic, A., C., Cuadraro, A., Weber, D., Poulsen, H., E., Grune, T., Schmidt, H., H., H., W., & Ghezzi, P. (2015). Clinical relevance of biomarkers of oxidative stress. *Antioxidants & redox signaling*, 23(14), 1144-1170.
- Furukawa, S., Fujita, T., Shimabukuro, M., Iwaki, M., Yamada, Y., Nakajima, Y., ... & Shimomura, I. (2017). Increased oxidative stress in obesity and its impact on metabolic syndrome. *The Journal of clinical investigation*, 114(12), 1752-1761.

- Galam, D., Silva, J., Sanders, D., & Oard, J. H. (2015). Morphological and genetic survey of Giant *Salvinia* populations in Louisiana and Texas. *Aquatic Botany*, 127, 20-25.
- Genestra, M. (2007). Oxyl radicals, redox-sensitive signalling cascades and antioxidants. *Cellular signalling*, 19(9), 1807-1819.
- George, G. T., & Gabriel, J. J. (2016). *In vitro* antioxidant potential of secondary and tertiary growth forms of *Salvinia molesta* Mitchell. *World Journal of Pharmaceutical Sciences*, 4(3), 450-462.
- Gettys, L. A. (2014). Aquatic weed management: control methods. *SRAC Publication—Southern Regional Aquaculture Centre*.
- Ghani, A. (2003). *Medicinal plants of Bangladesh: chemical constituents and uses*. 2nd Edition. Asiatic society of Bangladesh. Nimtali, Dhaka, Bangladesh.
- Godstime, O. C., Enwa, F. O., Jewo A. O., & Eze, C. O. (2014). Mechanisms of antimicrobial actions of phytochemicals against enteric pathogens—a review. *J Pharm Chem Biol Sci*, 2(2), 77-85.
- Abarca-Vargas, R., Malacara, C. F. P., & Petricevich, V. L. (2016). Characterization of chemical compounds with antioxidant and cytotoxic activities in *Bougainvillea x Buttiana holttum* and *standl*, (Var. rose) extracts. *Antioxidants*, 5(4), 45.
- Gopal, B. (1987). *Water hyacinth*. Elsevier Science Publishers.
- Gopalakrishnan, R., & Elumalai, N. (2022). Preliminary Phytochemical Screening And In Vitro Antioxidant Activity of Methanol Ethanol And Aqueous Extract of Polyherbal Formulation (PHF). *European Journal of Biomedical*, 9(12), 293-301.
- Gulcin, I., Kirecci, E., Akkemik, E., Topal, F., & Hisar, O. (2010). Antioxidant, antibacterial, and anticandidal activities of an aquatic plant: duckweed (*Lemna minor* L. Lemnaceae). *Turkish Journal of Biology*, 34(2), 175-188.
- Gülçin, İ., Oktay, M., Kireççi, E., & Küfrevioğlu, Ö. İ. (2003). Screening of antioxidant and antimicrobial activities of anise (*Pimpinella anisum* L.) seed extracts. *Food chemistry*, 83(3), 371-382.
- Habibi, N., Grieger, J. A., & Bianco-Miotto, T. (2020). A review of the potential interaction of selenium and iodine on placental and child health. *Nutrients*, 12(9), 2678.

- Hajam, Y. A., Rani, R., Ganie, S. Y., Sheikh, T. A., Javaid, D., Qadri, S. S., ... & Reshi, M. S. (2022). Oxidative stress in human pathology and aging: Molecular mechanisms and perspectives. *Cells*, 11(3), 552.
- Haller W. 2009. Hydrilla. In: Gettys LA, Haller WT, Bellaud M (eds). Introduction to the Plant Monographs. Biology and Control of Aquatic Plants: A Best Management Practices Handbook. Aquatic Ecosystem Restoration Foundation, Marietta GA, USA.
- Hamid, H. H., Ghaima, K. K., & Najem, A. M. (2013). Photochemical, antioxidant and antibacterial activities of some extracts of water hyacinth (*Eichhornia crassipes*) leaves. *Int. J. Adv. Pharm. Res*, 4, 1847-1851.
- Hanafiah, M. M., Mohamad, N. H. S. M., & Aziz, N. I. H. A. (2018). *Salvinia molesta* dan *Pistia stratiotes* sebagai agen fitoremediasi dalam rawatan air sisa kumbahan. *Sains Malaysiana*, 47(8), 1625-1634.
- Harborne, J. B. (1973). Phytochemical methods London Chapman and Hall.
- Harborne, J. B. (1980). Secondary Plant Products: Encyclopedia of Plant Physiology, New Series Volume 8: edited by EA Bell and BV Charlwood. Springer, Berlin, 1980. 674+ xvi pp. DM 198 (ca£ 50).
- Harley, K. L. S., & Mitchell, D., S. (1981). The Biology of Australian Weeds. 6: *Salvinia molesta* DS Mitchell.
- Harvey, A. L. (2008). Natural products in drug discovery. *Drug discovery today*, 13(19-20), 894-901.
- Haslam, E. (1996). Natural polyphenols (vegetable tannins) as drugs: possible modes of action. *Journal of natural products*, 59(2), 205-215.
- Hassan, A., & Nawchoo, I. A. (2020). Impact of invasive plants in aquatic ecosystems. In *Bioremediation and Biotechnology*, Springer, Cham., 55-73.
- Havel, J. E., Kovalenko, K. E., Thomaz, S. M., Amaltano, S., & Kats, L. B. 2015. Aquatic invasive species: challenges for the future. *Hydrobiologia*, 750(1), 147170.
- Hayder, G., & Mustafa, H. M. (2021). Cultivation of aquatic plants for biofiltration of wastewater. *Lett. Appl. NanoBioScience*, 10, 1919-1924.v
- Heleno, S. A., Martins, A., Queiroz, M. J. R., & Ferreira, I. C. (2015). Bioactivity of phenolic acids: Metabolites versus parent compounds: A review. *Food chemistry*, 173, 501-513.

- Hikmawanti, N. P. E., Fatmawati, S., & Asri, A. W. (2021, April). The effect of ethanol concentrations as the extraction solvent on antioxidant activity of Katuk (*Sauropus androgynus* (L.) Merr.) leaves extracts. In *IOP Conference Series: Earth and Environmental Science* (Vol. 755, No. 1, p. 012060). IOP Publishing.
- Hill, M. P. (2003). The impact and control of alien aquatic vegetation in South African aquatic ecosystems. *African Journal of Aquatic Science*, 28(1), 19-24.
- Hill, M. P., & Coetzee, J. (2017). The biological control of aquatic weeds in South Africa: Current status and future challenges. *Bothalia-African Biodiversity & Conservation*, 47(2), 1-12.
- Houghton, P. J., & Raman, A. (1998). Methods for extraction and sample clean-up. *Laboratory Handbook for the Fractionation of Natural Extracts*, 22-53.
- Hussen, E. M., & Endalew, S. A. (2023). In vitro antioxidant and free-radical scavenging activities of polar leaf extracts of *Vernonia amygdalina*. *BMC Complementary Medicine and Therapies*, 23(1), 1-12.
- Ionela, D. C., & Ion, I. B. (2007). Plant products as antimicrobial agents. *Section Genetics Molecular Biology*, 8, 151.
- Iqbal, E., Salim, K. A., & Lim, L. B. (2015). Phytochemical screening, total phenolics and antioxidant activities of bark and leaf extracts of *Goniothalamus velutinus* (Airy Shaw) from Brunei Darussalam. *Journal of King Saud University-Science*, 27(3), 224-232.
- Ismail, S. N. (2014). *A Comparative Study Between Pistia Stratiotes L. and Salvinia molesta Mitch. Based on Life Cycle and Salinity Tolerance*. Universiti Sains Malaysia: PhD. Thesis.
- Ismail, S. N., Subehi, L., Mansor, A., & Mashhor, M. (2019, November). Invasive Aquatic Plant Species of Chenderoh Reservoir, Malaysia and Jatiluhur Reservoir, Indonesia. In *IOP Conference Series: Earth and Environmental Science* (Vol. 380, No. 1, p. 012004). IOP Publishing.
- Jabatan Landskap Negara. (2021). *Taman Botani Johor*. Retrieved on December 15, 2022, from <http://www.jln.gov.my/index.php/pages/view/504>
- Jaganjac, M., Milkovic, L., Zarkovic, N., & Zarkovic, K. (2022). Oxidative stress and regeneration. *Free Radical Biology and Medicine*, 181, 154-165.
- Jakopic, J., & Veberic, R. (2009). Extraction of phenolic compounds from green walnut fruits in different solvents. *Acta Agriculturae Slovenica*, 93(1), 11.

- Jang, H. D., Chang, K. S., Huang, Y. S., Hsu, C. L., Lee, S. H., & Su, M. S. (2007). Principal phenolic phytochemicals and antioxidant activities of three Chinese medicinal plants. *Food chemistry*, 103(3), 749-756.
- Jasmine, R., Daisy, P., & Selvakumar, B. N. (2007). A Novel Terpenoid from *Elephantopus scaber* with Antibacterial Activity against Beta lactamase-Producing Clinical Isolates. *Research Journal of Microbiology*, 2(10), 770-775.
- Jayanthi, P., Lalitha, P., & Shubashini, K. S. (2011). Phytochemical investigation of the extracts of *Eichhornia crassipes* and its solvent fractionates. *Journal of Pharmacy Research*, 4(5), 1405-1406.
- Jo, I., Fridley, J. D., & Frank, D. A. (2017). Invasive plants accelerate nitrogen cycling: evidence from experimental woody monocultures. *Journal of Ecology*, 105(4), 1105-1110.
- Johnston, J. E., Sepe, H. A., Miano, C. L., Brannan, R. G., & Alderton, A. L. (2005). Honey inhibits lipid oxidation in ready-to-eat ground beef patties. *Meat science*, 70(4), 627-631.
- Julien, M. H., Hill, M. P., & Tipping, P. W. (2009). *Salvinia molesta* DS Mitchell (Salviniaceae). *Weed biological control with arthropods in the tropics*. Cambridge University Press, Cambridge, 378-407.
- Julien, M., McFadyen, R., & Cullen, J. (2012). *Salvinia molesta* DS Mitchell—salvinia. *Biological control of weeds in Australia*, 518-525.
- Kamaraj, M., Dhana Ranges Kumar, V., Nithya, T. G., & Danya, U. (2020). Assessment of antioxidant, antibacterial activity and phytoactive compounds of aqueous extracts of avocado fruit peel from Ethiopia. *International Journal of Peptide Research and Therapeutics*, 26(3), 1549-1557.
- Kashiwada, Y., Huang, L., Kilkuskie, R. E., Bodner, A. J., & Lee, K. H. (1992). New hexahydroxydiphenyl derivatives as potent inhibitors of HIV replication in H9 lymphocytes. *Bioorganic & medicinal chemistry letters*, 2(3), 235-238.
- Kaur, G. J., & Arora, D. S. (2009). Antibacterial and phytochemical screening of *Anethum graveolens*, *Foeniculum vulgare* and *Trachyspermum ammi*. *BMC complementary and alternative medicine*, 9(1), 1-10.
- Kchaou, W., Abbès, F., Blecker, C., Attia, H., & Besbes, S. (2013). Effects of extraction solvents on phenolic contents and antioxidant activities of Tunisian

- date varieties (*Phoenix dactylifera* L.). *Industrial crops and products*, 45, 262-269.
- Keller, R. P., Masoodi, A., & Shackleton, R. T. (2018). The impact of invasive aquatic plants on ecosystem services and human well-being in Wular Lake, India, *Reg. Environ. Chang.*, 18(3), 847–857, doi: 10.1007/s10113-017-1232-3.
- Khanbabaee, K., & Van Ree, T. (2001). Tannins: classification and definition. *Natural product reports*, 18(6), 641-649.
- Khare, C.P. (2005). *Encyclopedia of Indian medicinal plants*. Berlin Heidelberg, Germany: Springer-Verlag. p. 372.
- Kiruba, R. M., Christobel, G. J., Mini, J. J., Sankaralingam, S., Venkatesh, S., Sundari, A., & Soorya, C. (2023). Phytochemical Analysis, Antioxidant and Antimicrobial Activity of Aquatic Plant *Trapa natans* L.-Medicinal Aquatic Plant. *Asian Journal of Biochemistry, Genetics and Molecular Biology*, 13(3), 1-14.
- Kittakoop, P., Mahidol, C., & Ruchirawat, S. (2014). Alkaloids as important scaffolds in therapeutic drugs for the treatments of cancer, tuberculosis, and smoking cessation. *Current topics in medicinal chemistry*, 14(2), 239-252.
- Kuete, V., Ngameni, B., Simo, C. F., Tankeu, R. K., Ngadjui, B. T., Meyer, J. J. M., ... & Kuate, J. R. (2008). Antimicrobial activity of the crude extracts and compounds from *Ficus chlamydocarpa* and *Ficus cordata* (Moraceae). *Journal of ethnopharmacology*, 120(1), 17-24.
- Lacoul, P., & Freedman, B. (2006). Environmental influences on aquatic plants in freshwater ecosystems. *Environmental Reviews*, 14(2), 89-136.
- Lafay, S., & Gil-Izquierdo, A. (2008). Bioavailability of phenolic acids. *Phytochemistry Reviews*, 7(2), 301-311.
- Lal, A. (2016). *Salvinia molesta*: an assessment of the effects and methods of eradication.
- Lalitha, T., P. & Jayanthi, P. (2012). Preliminary studies on phytochemicals and antimicrobial activity of solvent extracts of *Eichhornia crassipes* (Mart.) Solms. *Asian Journal of Plant Science and Research*, 2(2), 115-122.
- Lata, N., & Dubey, V. (2010). Quantification and identification of alkaloids of *Eichhornia crassipes*: the world's worst aquatic plant. *J Phar Res*, 3, 1229-1231.

- Lata, N., & Veenapani, D. (2010). Isolation of flavonoids from *Eichhornia crassipes*: the world's worst aquatic plant. *Journal of Pharmacy Research*, 3(9), 2116-2118.
- Lee, J. H., Park, K. H., Lee, M. H., Kim, H. T., Seo, W. D., Kim, J. Y., ... & Ha, T. J. (2013). Identification, characterisation, and quantification of phenolic compounds in the antioxidant activity-containing fraction from the seeds of Korean perilla (*Perilla frutescens*) cultivars. *Food chemistry*, 136(2), 843-852.
- Lembi, C. A. (2009). Identifying and Managing Aquatic Vegetation. Aquatic plant management. Formerly Purdue Extension. Publication WS-21-W. *Purdue University*, APM-3-W No: 03/09, USA, 19.
- Li, X., Wu, X., & Huang, L. (2009). Correlation between antioxidant activities and phenolic contents of radix *Angelicae sinensis* (Danggui). *Molecules*, 14(12), 5349-5361.
- Liu, H. W., He, L. Y., Gao, J. M., Ma, Y. B., Zhang, X. M., Peng, H., & Chen, J. J. (2008). Chemical constituents from the aquatic weed *Pistia stratiotes*. *Chemistry of Natural Compounds*, 44(2), 236-238.
- Liu, R., Xing, L., Fu, Q., Zhou, G. H., & Zhang, W. G. (2016). A review of antioxidant peptides derived from meat muscle and by-products. *Antioxidants*, 5(3), 32.
- Lobo, V., Patil, A., Phatak, A., & Chandra, N. (2010). Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy reviews*, 4(8), 118.
- Losada-Barreiro, S., & Bravo-Diaz, C. (2017). Free radicals and polyphenols: The redox chemistry of neurodegenerative diseases. *European journal of medicinal chemistry*, 133, 379-402.
- Luque, G. M., Bellard, C., Bertelsmeier, C., Bonnaud, E., Genovesi, P., Simberloff, D., & Courchamp, F. (2014). The 100th of the world's worst invasive alien species. *Biological invasions*, 16(5), 981-985.
- Madsen, J. D., & Wersal, R. M. (2008). Growth regulation of *Salvinia molesta* by pH and available water column nutrients. *Journal of Freshwater Ecology*, 23(2), 305-313.
- Maharjan, N. (2008). *Evaluation of antibacterial activities of medicinal plants* (Doctoral dissertation, Department of Microbiology).
- Mahdi-Pour, B., Jothy, S. L., Latha, L. Y., Chen, Y., & Sasidharan, S. (2012). Antioxidant activity of methanol extracts of different parts of *Lantana camara*. *Asian Pacific journal of tropical biomedicine*, 2(12), 960-965.

- Mali, S. (2017). Kiambang *Salvinia molesta* Pembunuh Senyap Biodiversiti. 9. 1-3.
- Manzoor, A., Yousuf, B., Pandith, J. A., & Ahmad, S. (2023). Plant-derived active substances incorporated as antioxidant, antibacterial or antifungal components in coatings/films for food packaging applications. *Food Bioscience*, 102717.
- Mardawati, E., Filianty, F., & Marta, H. (2008). Antioxide activity study of manggis skin extract (*Garcinia mangostana* L) in order to use manggis skin waste in puspahiang district, tasikmalaya regency. *Teknotan: Jurnal Industri & Teknologi Pertanian*, 2(3), 1-5.
- Maseko, Z., Coetzee, J. A., & Hill, M. P. (2019). Effect of shade and eutrophication on the biological control of *Salvinia molesta* (Salviniaceae) by the weevil *Cyrtobagous salviniae* (Coleoptera: Eirirhinidae). *Austral Entomology*, 58(3), 595-601.
- Mashhor, M., Noreha, M. N., & Ahyaudin, A. (2002). Effects of clearing on the succession of aquatic weeds along irrigation and drainage canals in the MUDA area. In *Rice agroecosystem of the Muda irrigation scheme, Malaysia*.
- McDonald, S., Prenzler, P. D., Antolovich, M., & Robards, K. (2001). Phenolic content and antioxidant activity of olive extracts. *Food chemistry*, 73(1), 73-84.
- Mcfarland, D.G., Nelson, L.S., Grodowitz, M.J., Smart, R.M., & Owens, C.S. (2004). *Salvinia molesta* D. S. Mitchell (Giant Salvinia) in the United States: A Review of Species Ecology and Approaches to Management.
- Mogana, R., Adhikari, A., Tzar, M. N., Ramliza, R., & Wiart, C. (2020). Antibacterial activities of the extracts, fractions and isolated compounds from *Canarium patentinervium* Miq. against bacterial clinical isolates. *BMC complementary medicine and therapies*, 20(1), 1-11.
- Mohd-Nizam, N. U., Mohd-Hanafiah, M., Mohd-Noor, I., & Abd-Karim, H. I. (2020). Efficiency of five selected aquatic plants in phytoremediation of aquaculture wastewater. *Applied Sciences*, 10(8), 2712.
- Mogana, R., Adhikari, A., Tzar, M. N., Ramliza, R., & Wiart, C. (2020). Antibacterial activities of the extracts, fractions and isolated compounds from *Canarium patentinervium* Miq. against bacterial clinical isolates. *BMC complementary medicine and therapies*, 20(1), 1-11.

- Molyneux, S. L., Lister, C. E., & Savage, G. P. (2004). An investigation of the antioxidant properties and colour of glasshouse grown tomatoes. *International Journal of Food Sciences and Nutrition*, 55(7), 537-545.
- Mostafa, A. A., Al-Askar, A. A., Almaary, K. S., Dawoud, T. M., Sholkamy, E. N., & Bakri, M. M. (2018). Antimicrobial activity of some plant extracts against bacterial strains causing food poisoning diseases. *Saudi journal of biological sciences*, 25(2), 361-366.
- Mtewa, A. G., Deyno, S., Ngwira, K., Lampiao, F., Peter, E. L., Ahovegbe, L. Y., ... & Sesaaazi, D. C. (2018). Drug-like properties of anticancer molecules elucidated from *Eichhornia crassipes*.
- Muda, S. A. (2010). *Effects of Nymphaea caerulea on Wastewater Quality from Palm Oil Production*. Universiti Malaysia Pahang: Degree Thesis.
- Muhammad, G., Muammad, I., Sobia, K., Dawood, A., Muhammad, J. A., Kashis, S. A., ... & Sajid, M. (2014). A comparative study of antimicrobial and antioxidant activities of garlic (*Allium sativum* L.) extracts in various localities of Pakistan. *African Journal of Plant Science*, 8(6), 298-306.
- Mukheriee, P. K., & Wahile, A. (2006). Integrated approaches towards drug development from Ayurveda and other Indian system of medicines. *Journal of Ethnopharmacology*, 103(1), 25-35.
- Mulat, M., Pandita, A., & Khan, F. (2019). Medicinal plant compounds for combating the multi-drug resistant pathogenic bacteria: a review. *Current pharmaceutical biotechnology*, 20(3), 183-196.
- Mustafa, H. M., & Hayder, G. (2020). Performance of *Pistia stratiotes*, *Salvinia molesta*, and *Eichhornia crassipes* aquatic plants in the tertiary treatment of domestic wastewater with varying retention times. *Applied Sciences*, 10(24), 9105.
- Mustafa, H. M., & Hayder, G. (2021). Performance of *Salvinia molesta* plants in tertiary treatment of domestic wastewater. *Heliyon*, 7(1), e06040.
- Naczka, M., & Shahidi, F. (2004). Extraction and analysis of phenolics in food. *Journal of chromatography A*, 1054(1-2), 95-111.
- Nagaprashantha, L. D., Vatsyayan, R., Singhal, J., Fast, S., Roby, R., Awasthi, S., & Singhal, S. S. (2011). Anti-cancer effects of novel flavonoid vicenin-2 as a single agent and in synergistic combination with docetaxel in prostate cancer. *Biochemical pharmacology*, 82(9), 1100-1109.

- Narayani, M., Johnson, M., Sivaraman, A., & Janakiraman, N. (2012). Phytochemical and antibacterial studies on *Jatropha curcas* L. *Journal of Chemical and Pharmaceutical Research*, 4(5), 2639-2642.
- Nascimento, G. G., Locatelli, J., Freitas, P. C., & Silva, G. L. (2000). Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. *Brazilian journal of microbiology*, 31, 247-256.
- Negi, P. S. (2012). Plant extracts for the control of bacterial growth: Efficacy, stability and safety issues for food application. *International journal of food microbiology*, 156(1), 7-17.
- Neha, K., Haider, M. R., Pathak, A., & Yar, M. S. (2019). Medicinal prospects of antioxidants: A review. *European journal of medicinal chemistry*, 178, 687-704.
- Nelson, L. S. (2009). Giant and common salvinia. *Biology and control of aquatic plants*. Gainesville: Cover photograph courtesy of SePRO Corporation.
- Ng, Y. S., & Chan, D. J. C. (2017). Wastewater phytoremediation by *Salvinia molesta*. *Journal of Water Process Engineering*, 15, 107-115.
- Ng, Y. S., Samsudin, N. I. S., & Chan, D. J. C. (2017, June). Phytoremediation capabilities of *Spirodela polyrhiza* and *Salvinia molesta* in fish farm wastewater: A preliminary study. In *IOP Conference Series: Materials Science and Engineering* (Vol. 206, No. 1, p. 012084). IOP Publishing.
- Ngo, T. V., Scarlett, C. J., Bowyer, M. C., Ngo, P. D., & Vuong, Q. V. (2017). Impact of different extraction solvents on bioactive compounds and antioxidant capacity from the root of *Salacia chinensis* L. *Journal of Food Quality*.
- Nithya, T. G., Jayanthi, J., & Raghunathan, M. G. (2015). Phytochemical, Antibacterial and GC MS analysis of a floating fern *Salvinia molesta* DS Mitchell (1972). *International Journal of PharmTech Research*, 8(9), 85-90.
- Nithya, T. G., Jayanthi, J., & Raghunathan, M. G. (2016). Antioxidant activity, total phenol, flavonoid, alkaloid, tannin, and saponin contents of leaf extracts of *Salvinia molesta* DS Mitchell (1972). *Asian J Pharm Clin Res*, 9(1), 200-203.
- Nyananyo, B. L., Gijo, A. H., & Ogamba, E. N. (2007). The Physico-chemistry and Distribution of Water Hyacinth (*Eichhornia crassipes*) on the river Nun in the Niger Delta. *Journal of Applied Sciences and Environmental Management*, 11(3).

- O'Hare, M. T., Aguiar, F. C., Asaeda, T., Bakker, E. S., Chambers, P. A., Clayton, J. S., ... & Wood, K. A. (2018). Plants in aquatic ecosystems: current trends and future directions. *Hydrobiologia*, 812(1), 1-11.
- O'Neill, J. (2014). Antimicrobial resistance: tackling a crisis for the health and wealth of nations. *Rev. Antimicrob. Resist.*
- Obadoni, B. O., & Ochuko, P. O. (2002). Phytochemical studies and comparative efficacy of the crude extracts of some haemostatic plants in Edo and Delta States of Nigeria. *Global Journal of pure and applied sciences*, 8(2), 203-208.
- Oladunmoye, M. K. (2006). Comparative evaluation of antimicrobial activities and phytochemical screening of two varieties of *Acalypha wilkesiana*. *Int J Trop Med*, 1(3), 1348-1356.
- Oliver, J. D. (1993). A review of the biology of giant Salvinia (*Salvinia molesta* Mitchell). *Journal of Aquatic Plant Management*, 31, 227.
- Omar, K., Geronikaki, A., Zoumpoulakis, P., Camoutsis, C., Soković, M., Ćirić, A., & Glamočlija, J. (2010). Novel 4-thiazolidinone derivatives as potential antifungal and antibacterial drugs. *Bioorganic & medicinal chemistry*, 18(1), 426-432.
- Oosterhout, E., V. (2006). *Salvinia Control Manual: Management and Control Options for Salvinia (Salvinia Molesta) in Australia*. NSW Department of Primary Industries.
- Otitolaiye, C., Omonkhua, A., Oriakhi, K., Okello, E., Onoagbe, I., & Okonofua, F. (2023). Phytochemical analysis and in-vitro antioxidant potential of aqueous and ethanol extracts of *Irvingia gabonensis* stem bark. *Pharmacognosy Research*.
- Oyaizu, M. (1986). Studies on products of browning reaction antioxidative activities of products of browning reaction prepared from glucosamine. *The Japanese journal of nutrition and dietetics*, 44(6), 307-315.
- Ozawa, T., Lilley, T. H., & Haslam, E. (1987). Polyphenol interactions: astringency and the loss of astringency in ripening fruit. *Phytochemistry*, 26(11), 2937-2942.
- Parekh, J., & Chanda, S. (2007). In vitro antibacterial activity of the crude methanol extract of *Woodfordia fruticosa* Kurz. flower (Lythraceae). *Brazilian Journal of Microbiology*, 38, 204-207.

- Park, Y., Lee, H. J., Lee, S. S., & Choi, D. H. (2003). Studies on biological activity of wood extractives-chemical components and antioxidative activity of the leaves of *Sophora japonica*. *Modchae Konghak*, 31, 43-48.
- Paul, R. K., Dutta, D., Chakraborty, D., Nayak, A., Dutta, P. K., & Nag, M. (2019). Antimicrobial agents from natural sources: An overview. *Advance Pharmaceutical Journal*, 4(2), 41-51.
- Pisoschi, A. M., Pop, A., Iordache, F., Stanca, L., Predoi, G., & Serban, A. I. (2021). Oxidative stress mitigation by antioxidants-an overview on their chemistry and influences on health status. *European Journal of Medicinal Chemistry*, 209, 112891.
- Płotka-Wasyłka, J., Rutkowska, M., Owczarek, K., Tobiszewski, M., & Namieśnik, J. (2017). Extraction with environmentally friendly solvents. *TrAC Trends in Analytical Chemistry*, 91, 12-25.
- Podsędek, A. (2007). Natural antioxidants and antioxidant capacity of Brassica vegetables: A review. *LWT-Food Science and Technology*, 40(1), 1-11.
- Powers, S. K., Ji, L. L., Kavazis, A. N., & Jackson, M. J. (2011). Reactive oxygen species: impact on skeletal muscle. *Comprehensive physiology*, 1(2), 941.
- Prasad, R. N., Viswanathan, S., Devi, J. R., Nayak, V., Swetha, V. C., Archana, B. R., Parathasarathy, N., & Rajkumar, J. (2008). Preliminary phytochemical screening and antimicrobial activity of *Samanea saman*. *Journal of Medicinal Plants Research*, 2(10), 268-270.
- Purushoth, P. T., Panneerselvam, P., Suresh, R., Clement, A. W., & Balasubramanian, S. (2013). GC-MS analysis of ethanolic extract of *Canthium parviflorum* Lamk Leaf.
- Raja, K. S., Taip, F. S., Azmi, M. M. Z., & Shishir, M. R. I. (2019). Effect of pre-treatment and different drying methods on the physicochemical properties of *Carica papaya* L. leaf powder. *Journal of the Saudi Society of Agricultural Sciences*, 18(2), 150-156.
- Rastogi, R. P., & Mehrotra, B. N. (1993). *Compendium of Indian medicinal plants*. Central Drug Research Institute. New Delhi, India.
- Razak, N. A., Shaari, A. R., Jolkili, M., & Leng, L. Y. (2016). Drying curves and colour changes of *Cassia alata* leaves at different temperatures. In *MATEC Web of Conferences* (Vol. 78, p. 01020). EDP Sciences.

- Rebey, I. B., Borgou, S., Debez, I. B. S., Karoui, I. J., Sellami, I. H., Msaada, K., Limam, F., & Marzouk, B. (2012). Effects of extraction solvents and provenances on phenolic contents and antioxidant activities of cumin (*Cuminum cyminum* L.) seeds. *Food and Bioprocess Technology*, 5, 2827-2836.
- Regitz, M., & Falbe, J. (1995). CD-Römpf Chemie Lexikon, Version 1.0.
- Rhoades, D. F. (1979). Evolution of plant chemical defense against herbivores. *Herbivores: their interaction with secondary plant metabolites*, 3-54.
- Richardson, D. M., Pyšek, P., Rejmanek, M., Barbour, M. G., Panetta, F. D., & West, C. J. (2000). Naturalization and invasion of alien plants: concepts and definitions. *Diversity and distributions*, 6(2), 93-107.
- Room, P. M. (1988). Effects of temperature, nutrients and a beetle on branch architecture of the floating weed *Salvinia molesta* and simulations of biological control. *The Journal of Ecology*, 826-848.
- Room, P. M. (1990). Ecology of a simple plant-herbivore system. Biological control of *Salvinia*. *Trends in Ecology & Evolution*, 5(3), 74-79.
- Sagbo, I., & Mbeng, W. (2018). Plants used for cosmetics in the Eastern Cape Province of South Africa: A case study of skin care. *Pharmacognosy Reviews*, 12(24), 139-156.
- Salisbury, D., & Bronas, U. (2015). Reactive oxygen and nitrogen species: impact on endothelial dysfunction. *Nursing research*, 64(1), 53-66.
- Santhosh, P., Nithya, T. G., Lakshmi, S. G., Marino, G. L. S., Balavaishnavi, B., & Kamaraj, M. (2022). Assessment of phytochemicals, antioxidant, antibacterial activity, and profiling of functional molecules in novel freshwater fern *Salvinia cucullata* Roxb. *South African Journal of Botany*.
- Sarker, S. D., Latif, Z., & Gray, A. I. (2005). Natural product isolation: an overview. *Natural Products Isolation*, 1-25.
- Sarmah, P., Dan, M. M., Adapa, D., & Sarangi, T. K. (2018). A review on common pathogenic microorganisms and their impact on human health. *Electronic Journal of Biology*, 14(1), 50-58.
- Scheffer, M., & Jeppesen, E. (2007). Regime shifts in shallow lakes. *Ecosystems*, 10(1), 1-3.

- Sen, A., & Batra, A. (2012). Evaluation of antimicrobial activity of different solvent extracts of medicinal plant: *Melia azedarach* L. *Int J Curr Pharm Res*, 4(2), 67-73.
- Sen, R., Nayak, L., & De, R. K. (2016). A review on host–pathogen interactions: classification and prediction. *European Journal of Clinical Microbiology & Infectious Diseases*, 35, 1581-1599.
- Senhaji, S., Lamchouri, F., & Toufik, H. (2020). Phytochemical content, antibacterial and antioxidant potential of endemic plant *Anabasis aretioides* Coss. & Moq. (Chenopodiaceae). *BioMed Research International*, 87(2), 67-78.
- Shaheen, S., Naseer, S., Ashraf, M., & Akram, N. A. (2013). Salt stress affects water relations, photosynthesis, and oxidative defense mechanisms in *Solanum melongena* L. *Journal of Plant Interactions*, 8(1), 85–96.
- Shan, B., Cai, Y. Z., Brooks, J. D., & Corke, H. (2007). The in vitro antibacterial activity of dietary spice and medicinal herb extracts. *International Journal of food microbiology*, 117(1), 112-119.
- Shanab, S. M. M., Ameer, M. A., Fekry, A. M., Ghoneim, A. A., & Shalaby, E. A. (2011). Corrosion resistance of magnesium alloy (AZ31E) as orthopaedic biomaterials in sodium chloride containing antioxidantly active compounds from *Eichhornia crassipes*. *Int. J. Electrochem. Sci*, 6, 3017-3035.
- Shanab, S. M., Shalaby, E. A., Lightfoot, D. A., & El-Shemy, H. A. (2010). Allelopathic effects of water hyacinth [*Eichhornia crassipes*]. *PloS one*, 5(10), e13200.
- Sheikh, N., Kumar, Y., Misra, A. K., & Pfoze, L. (2013). Phytochemical screening to validate the ethnobotanical importance of root tubers of *Dioscorea* species of Meghalaya, North East India. *Journal of Medicinal Plants*, 1(6), 62-69.
- Shi, Q. I. U., Hui, S. U. N., Zhang, A. H., Hong-Ying, X. U., Guang-Li, Y. A. N., Ying, H. A. N., & Xi-Jun, W. A. N. G. (2014). Natural alkaloids: basic aspects, biological roles, and future perspectives. *Chinese Journal of Natural Medicines*, 12(6), 401-406.
- Sidek, N. M., Abdullah, S. R. S., Draman, S. F. S., Rosli, M. M. M., & Sanusi, M. F. (2018). Phytoremediation of abandoned mining lake by water hyacinth and water lettuces in constructed wetlands. *Jurnal Teknologi*, 80(5).
- Silva, V., Igrejas, G., Falco, V., Santos, T. P., Torres, C., Oliveira, A. M., ... & Poeta, P. (2018). Chemical composition, antioxidant and antimicrobial activity of

- phenolic compounds extracted from wine industry by-products. *Food control*, 92, 516-522.
- Slinkard, K., & Singleton, V. L. (1977). Total phenol analysis: automation and comparison with manual methods. *American journal of enology and viticulture*, 28(1), 49-55.
- Smith, J. E. (2011) Algae, [in:] Simberloff D., Rejmanek M. (eds) *Encyclopedia of biological invasions*, University of California Press, Los Angeles: 11-15.
- Sodipo, O. A., Akinniyi, J. A., & Ogunbameru, J. V. (2000). Studies on certain characteristics of extracts of bark of *Pausinystalia johimbe* and *Pausinystalia macroceras* (K Schum) Pierre ex Beille. *Global Journal of Pure and Applied Sciences*, 6(1), 83-88.
- Soerjani, M., Pancho, J. V., & Voung, N. V. (1975). Aquatic weed problems and control in Southeast Asia. *Hyacinth Control Journal*, 13, 2-3.
- Solimini, A. G., Cardoso, A. C., & Heiskanen, A. S. (2006). Indicators and methods for the ecological status assessment under the Water Framework Directive. *Linkages between chemical and biological quality of surface waters Joint Research Centre, European Commission*, 1-262.
- Sullivan, P. R., Postle, L. A., & Julien, M. (2011). Biological control of *Salvinia molesta* by *Cyrtobagous salviniae* in temperate Australia. *Biological Control*, 57(3), 222-228.
- Sultana, B., Anwar, F., & Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*, 14(6), 2167-2180.
- Surendraraj, A., Farvin, K. S., & Anandan, R. (2013). Antioxidant potential of water hyacinth (*Eichornia crassipes*): In vitro antioxidant activity and phenolic composition. *Journal of Aquatic Food Product Technology*, 22(1), 11-26.
- Syakilla, N., George, R., Chye, F. Y., Pindi, W., Mantihal, S., Wahab, N. A., Fadzwi, F. M., Gu, P., H., & Matanjun, P. (2022). A Review on Nutrients, Phytochemicals, and Health Benefits of Green Seaweed, *Caulerpa lentillifera*. *Foods*, 11(18), 2832.
- Taghvaei, M., & Jafari, S. M. (2015). Application and stability of natural antioxidants in edible oils in order to substitute synthetic additives. *Journal of food science and technology*, 52(3), 1272-1282.

- Thomas, P. A., & Room, A. P. (1986). Taxonomy and control of *Salvinia molesta*. *Nature*, 320(6063), 581-584.
- Tilak, J. C., Adhikari, S., & Devasagayam, T. P. (2004). Antioxidant properties of *Plumbago zeylanica*, an Indian medicinal plant and its active ingredient, plumbagin. *Redox report*, 9(4), 219-227.
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G., & Kaur, H. (2011). Phytochemical screening and extraction: a review. *Internationale pharmaceutica sciencia*, 1(1), 98-106.
- Toki, K., Saito, N., Iimura, K., Suzuki, T., & Honda, T. (1994). (Delphinidin 3-gentiobiosyl) (apigenin 7-glucosyl) malonate from the flowers of *Eichhornia crassipes*. *Phytochemistry*, 36(5), 1181-1183.
- Truong, D. H., Nguyen, D. H., Ta, N. T. A., Bui, A. V., Do, T. H., & Nguyen, H. C. (2019). Evaluation of the use of different solvents for phytochemical constituents, antioxidants, and in vitro anti-inflammatory activities of *Severinia buxifolia*. *Journal of food quality*, 2019.
- Tsao, R. (2010). Chemistry and biochemistry of dietary polyphenols. *Nutrients*, 2(12), 1231-1246.
- Tsuzuki, J. K., Svidzinski, T. I., Shinobu, C. S., Silva, L. F., Rodrigues-Filho, E., Cortez, D. A., & Ferreira, I. C. (2007). Antifungal activity of the extracts and saponins from *Sapindus saponaria* L. *Anais da Academia Brasileira de Ciências*, 79, 577-583.
- Tyagi, T., & Agarwal, M. (2017). Antioxidant properties and phenolic compounds in methanolic extracts of *Eichhornia crassipes*. *Research Journal of Phytochemistry*, 11(2), 85-89.
- Umesh, C. V., Jamsheer, A. M., & Prasad, M. A. (2018). The role of flavonoids in drug discovery—review on potential applications. *Research Journal of Life Sciences, Bioinformatic, Pharmaceutical and Chemical Sciences*, 4(1), 70-77.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M., & Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The international journal of biochemistry & cell biology*, 39(1), 44-84.
- Venkataraman, K., Khurana, S., & Tai, T. C. (2013). Oxidative stress in aging-matters of the heart and mind. *International journal of molecular sciences*, 14(9), 17897-17925.

- Villaño, D., Fernández-Pachón, M. S., Moyá, M. L., Troncoso, A. M., & García-Parrilla, M. C. (2007). Radical scavenging ability of polyphenolic compounds towards DPPH free radical. *Talanta*, 71(1), 230-235.
- Vuolo, M. M., Lima, V. S., & Junior, M. R. M. (2019). Phenolic compounds: Structure, classification, and antioxidant power. In *Bioactive compounds* (pp. 33-50). Woodhead Publishing.
- Wang, T. Y., Li, Q., & Bi, K. S. (2018). Bioactive flavonoids in medicinal plants: Structure, activity, and biological fate. *Asian journal of pharmaceutical sciences*, 13(1), 12-23.
- Wersal, R. M., & Madsen, J. D. (2012). Aquatic plants, their uses and risks: a review of the global status of aquatic plants.
- Westendarp, H. (2006). [Effects of tannins in animal nutrition]. *DTW. Deutsche Tierärztliche Wochenschrift*, 113 7, 264-8.
- Whiteman, J. B., & Room, P. M. (1991). Temperatures lethal to *Salvinia molesta* Mitchell. *Aquatic Botany*, 40(1), 27-35.
- Widyaningrum, I., Wibisono, N., & Kusumawati, A. H. (2020). Effect of extraction method on antimicrobial activity against staphylococcus aureus of tapak liman (elephantopus scaber l.) leaves.
- Williamson, G. (2017). The role of polyphenols in modern nutrition. *Nutrition bulletin*, 42(3), 226-235.
- Wood, K. A., O'Hare, M. T., McDonald, C., Searle, K. R., Daunt, F., & Stillman, R. A. (2017). Herbivore regulation of plant abundance in aquatic ecosystems. *Biological Reviews*, 92(2), 1128-1141.
- World Health Organization. (1993). *Research guidelines for evaluating the safety and efficacy of herbal medicines*. WHO Regional Office for the Western Pacific.
- Yadav, B. S., Yadav, R., Yadav, R. B., & Garg, M. (2016). Antioxidant activity of various extracts of selected gourd vegetables. *Journal of food science and technology*, 53(4), 1823-1833.
- Yisa, J. (2009). Phytochemical analysis and antimicrobial activity of *Scoparia dulcis* and *Nymphaea lotus*. *Australian Journal of Basic and Applied Sciences*, 3(4), 3975-3979.
- Zada, S., Sajjad, W., Rafiq, M., Ali, S., Hu, Z., Wang, H., & Cai, R. (2021). Cave microbes as a potential source of drugs development in the modern era. *Microbial ecology*, 1-14.

- Zargoosh, Z., Ghavam, M., Bacchetta, G., & Tavili, A. (2019). Effects of ecological factors on the antioxidant potential and total phenol content of *Scrophularia striata* Boiss. *Scientific Reports*, 9(1), 1-15.
- Zennie, T. M., & McClure, J. W. (1977). The flavonoid chemistry of *Pistia stratiotes* L. and the origin of the Lemnaceae. *Aquatic Botany*, 3, 49-54.
- Zgoda, J. R., & Porter, J. R. (2001). A convenient microdilution method for screening natural products against bacteria and fungi. *Pharmaceutical Biology*, 39(3), 221-225.
- Zhang, Y., Wu, S., Qin, Y., Liu, J., Liu, J., Wang, Q., Ren, F., & Zhang, H. (2018). Interaction of phenolic acids and their derivatives with human serum albumin: Structure–affinity relationships and effects on antioxidant activity. *Food chemistry*, 240, 1072-1080.
- Zulhaimi, H. I., Rosli, I. R., Kasim, K. F., Akmal, H. M., Nuradibah, M. A., & Sam, S. T. (2017, September). A comparative study of *Averrhoabilimbi* extraction method. In *AIP Conference Proceedings* (Vol. 1885, No. 1, p. 020178). AIP Publishing LLC.



PTTA UTHM
PERPUSTAKAAN TUNKU TUN AMINAH

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