DEVELOPMENT OF FORMULATED FISH FEED SUPPLEMENTED WITH MICROALGAE OF PHYCOREMEDIATION ON THE GROWTH PERFORMANCE AND FATTY ACIDS CONTENT OF OREOCHROMIS NILOTICUS

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DEDICATION

I would like to dedicate this achievement to my family especially to my beloved parents, Nur Hanisah Sarjit Binti Abdullah and Hairuddin Bin Haris. To my family members, Muhammad Ridhwan, Nurul Atikah, Nur Aisyah, Orked Nur Alesha, Mawar Nur Raisha and my beloved Kitty. Thank you for all your supports and always being there for me.

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ABSTRACT

The arising issues on the use of fishmeal which causes over exploitation of the wild caught fish and the increment cost of aquaculture is increasing day by day. This has led to find new alternatives to replace fishmeal such as microalgae. In this study, 2 % of Scenedesmus sp. cultivated in Bold Basal Medium and wet market wastewater was partially subtituted to replace fishmeal and tested to Oreochromis niloticus fingerlings for 12 weeks. The physicochemical properties of pelletized feed was analysed and compared between control formulation, formulation 1 and formulation 2. The results found that, addition of Scenedesmus sp. in the diet would not alter the hardness of the feed, yet the small increment of hardness in the diet formulation 1 and 2 might be due to the failure of maintaining weight of the feed before and after pelletizing process. Besides, the analysis of variance showed that moisture content of formulation 1 and 2 were significantly difference (p<0.05) and the mineral composition of the feed shows an improvement for formulation 2, however, it is still not meet the requirement due to the absence of complete diet ingredients in the formulations. Moreover, the growth performance of *Oreochromis niloticus* was analysed weekly using analytical balance and the results obtained showed an improvement in the final weight of the fish in formulation 1 and 2 up to 43.38g and 44.85 g respectively. The concentration of fatty acids obtained in this study was acceptable, though, the concentration of omega 6 is higher compared to omega 3. To conclude this finding, the addition of Scenedesmus sp. in the diet formulation for Oreochromis niloticus gives a small difference in physicochemical properties of the feed pellet, yet it facilitates in maintaining and improving the growth and the quality of muscle tissue of Oreochromis niloticus.



ABSTRAK

Isu berkaitan pengunaan makanan ikan asli yang menyebabkan eksploitasi ikan liar dan kenaikan kos akuakultur semakin meningkat dari hari ke hari. Hal demikian telah membawa kepada pencarian alternatif baru untuk menggantikan makanan ikan asli seperti mikroalga. Dalam kajian ini, 2 % daripada Scenedesmus sp. di ternak menggunakan media *Bold Basal* dan air pasar untuk menggantikan makanan ikan asli dan diuji kepada Oreochromis niloticus selama 12 minggu. Sifat fizikokimia pellet makanan dianalisis dan dibandingkan antara formulasi kawalan, formulasi 1 dan formulasi 2. Bedasarkan keputusan analisis varians, penambahan Scenedesmus sp. dalam diet tidak mengubah kekerasan pelet makanan, namun peningkatan kecil kekerasan pelet dalam formulasi 1 dan 2 mungkin disebabkan oleh kegagalan mengekalkan berat pelet sebelum dan selepas proses pembuatan pelet. Selain itu, analisis varians menunjukkan bahawa kandungan kelembapan formulasi 1 dan 2 mempunyai perbezaan yang ketara (p<0.05) dan kandungan mineral di dalam pelet makanan menunjukkan peningkatan untuk formulasi 2. Walau bagaimanapun, ia masih tidak memenuhi nilai keperluan yang dicadangkan kerana ketiadaan diet yang seimbang dalam formulasi. Selain itu, prestasi pertumbuhan ikan Oreochromis niloticus dianalisis setiap minggu menggunakan penimbang elektronik dan data yang diperolehi menunjukkan peningkatan berat akhir ikan dalam formulasi 1 dan 2 masingmasing sebanyak 43.38g dan 44.85 g. Kepekatan asid lemak yang diperolehi dalam kajian ini boleh diterima, walaupun, kepekatan omega 6 adalah lebih tinggi berbanding omega 3. Sebagai kesimpulan, penambahan Scenedesmus sp. dalam formulasi diet untuk Oreochromis niloticus memberikan perbezaan kecil dalam sifat fizikokimia pelet suapan, namun ia membantu dalam mengekalkan dan meningkatkan pertumbuhan dan kualiti tisu otot Oreochromis niloticus.



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LIST OF SYMBOLS AND ABBREVIATIONS

AAFCO	-	Association of American Feed Control Official
ANOVA	-	Analysis of variance
AOAC	-	Association of Official Analytical Chemist
B1	-	Thiamine
B ₂	-	Riboflavin
B ₃	-	Niacin
B5	-	Pantothenic Acid
BBM	-	Bold Basal Medium
Cell/mL	-	Cell per milliliter
DHA	-	Docosahexaenoic Acid
DO	-	Dissolved oxygen
DOF	-	Department of Fisheries
Е	-	d-alpha-tocopherol
EPA	-	Eicosapentaenoic Acid
FAME	PUg	Fatty Acid Methyl Ester
FAO	-	Food and Agricultural Organization
FCR	-	Food Conversion Ratio
H ₂ SO ₄	-	Sulphuric Acid
HCL	-	Hydrochloric acid
Hg	-	Mercury
HMSO	-	Highline Medical Services Organization
HNO ₃	-	Nitric Acid
HPLC	-	High performance liquid chromatography
ICPMS	-	Inductive Coupled Plasma Mass Spectrometry
kg	-	Kilogram
КОН	-	Potassium Hydroxide
L	-	Liter

LC PUFA	-	Long Chain Polyunsaturated Fatty Acid
m	-	meter
MFAR	-	Malaysian Food Act and Regulation
mg/L	-	Milligram per liter
mL	-	Milliliter
mm	-	Millimeter
Mm Hg	-	Millimeter of Mercury
MUFA	-	Monounsaturated Fatty Acid
n3	-	Omega 3
n6	-	Omega 6
n9	-	Omega 9
NaOH	-	Sodium hydroxide
NH3	-	Ammonia
nm	-	Nanometer
NO ₃	-	Nitrate
NRC	-	National Research Council
NSP	-	Non-Starch Polysaccharide
PUFA	-	Polyunsaturated Fatty Acid
RPM	-	Revolution per minute
SFA	-	Saturated Fatty Acid
SGR	-19	Specific Growth Rate
SPSS	<u>9</u> 0	Statistical Package for Social Science
SR	-	Survival Rate
USA	-	United States of America
USEPA	-	United States Environmental Protection Agency
Uv-vis	-	Ultraviolet-Visible Spectroscopy
WHO	-	World Health and Organization
β sheet	-	Beta Sheet
μL	-	Microliter
μm	-	Micrometer



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- Arifin, S. N. H., Mohamed, R. M. S. R., Al- Gheethi, A. A. S., Lai, C. W., Gopalakrishnan, Y., Hairuddin, N. D. H. & Vo, D. (2021). Photocatalytic degradation of triclocarban in aqueous solution using a modified zeolite/TiO₂ composite: kinetic, mechanism study and toxicity assessment. *Environmental Science and Pollution Research*. <u>https://doi.org/10.1007/s11356-021-16732-y</u>
- Hairuddin, N. D., A Talip, B., Zing, N. Z., Mohamed, R. M. S. R., Al- Gheethi, A. A. S, Muhammad, N., Abdullah, N., Kahar, E. E. M. & Abdullah, S. (2020).
 Microalgae Production in Freshmarket Wastewater and Its Utilization as a Protein Substitute in Formulated Fish Feed for *Oreochromis* spp. in *Prospects of Freshmarket Wastes Management in Developing Countries*. Malaysia: Springer. pp. 77-88



- Hairuddin, N. D., A. Talip., B., Zing, N. Z., Mohamed, R. M. S. R., Al-Gheethi, A. A.
 S. (2019). The Analysis of Heavy Metals Accumulation in Oreochromis niloticus Muscle Tissue Feed with Formulated Fish Feed Supplemented with Microalgae. *Proc. of the Third Int Indonesia- Malaysia- Thailand Symposium on Innovation and Creativity on Cultivating Innovation and Creativity Culture*. Narathiwat. Princess of Naradhiwas University.p. 146
- A Talip, B., Zing, N. Z., Hairuddin, N. D., Mohamed, R. M. S. R., Muhammad, N., Basr, H., Abdullah, N., Kahar, E. E. M., Al Gheethi, A. A. S., A. & Abdullah, S. (2019). Determination of physicochemical properties of formulated fish feed supplemented with microalgae from bioremediation process. Jurnal Teknologi, *81* (3), 151-157

CHAPTER 1

INTRODUCTION

1.1 Research background

Aquaculture shows an increasing trend over the coming decades as it provides half of the fish for human consumption (Camacho-Rodriguez *et al.*, 2017). According to FAO (2018), aquaculture remains as one of the major food sectors worldwide due to its efficiency in producing edible protein in response to the substantial rise in the demand for fish. However, the feed inputs in aquaculture practices contribute up to 75% of total aquaculture production and the price of fishmeal as a main traditional ingredient used in the aquafeed keep on increasing due to its high protein and fatty acid content (FAO, 2018).

Malaysia is encircled by sea, and have many rivers and lakes. These ecosystems provide natural sources of fish and other aquatic resources (Yusoff, 2015) for its residents. Therefore, fish has also been one of the important daily diets of Malaysians and a leading source of protein. Based on the consumption pattern by Yusoff (2015), an average one family spends about 20% of their food consumption on fish. In fact, the fish consumption index increased from 53.1 kg in 2011 and as in 2020, the consumption index has reached 61.1 kg. In Malaysia, the aquaculture industry contributes about 16% to the national seafood supply (Ng, 2009).

Moreover, the aquaculture industry contributes to national food security and bringing in foreign exchange for the country. Total fish production in 2012 contributed about 1.7 millions of tonnes cost at RM6 billion to the national fish supply (Yusoff, 2015). Development of the fisheries sector is expected to become one of the major contributors to the national economy, as Malaysia rely heavily on aquaculture as a source of income and animal protein. (Ibrahim *et al.*, 2018). According to Yusoff



(2015), the demand for fish in Malaysia is expected to increase from 1.7 million tonnes in 2011 to 1.93 million tonnes by 2020. Besides, in producing a good diet formulation for fish, protein is considered as one of major food components. It is undeniable that fishmeal is a good protein for the diet of aquatic organisms. However, due to the high demand of aquaculture production with the increasing issues of using fishmeal as a main protein source in the diet formulation has led to the need to find new alternatives, such as animal and plant sources of proteins to replace fishmeal for the needs of aquaculture (Sirakov *et al.*, 2015). Referring to Sirakov *et al.* (2015), the most accessible and inexpensive food component in aquaculture is microalgae.

Microalgae is photosynthetic organisms and it is the main source of cellular carbon and chemical energy for other organisms (Hattab et al, 2014). The popularity of microalgae in the aquaculture industry is increasing day by day due to their appropriate size for ingestion, high nutritional value, high growth rate, antioxidant property, and disease resistance. The nutritional value of microalgae is influenced by many factors such as size, shape, digestibility, and biochemical compositions (Guedes and Malcata, 2012). Hattab et al. (2014) has mentioned that, microalgae are cultivated for many purposes such as a bio filter to remove nutrients and other pollutants from wastewater and also as fish feed in aquaculture. Culturing of microalgae as feed in aquaculture develop since Allen and Nelson's triumph in 1910 by cultivating Chlorella sp. in Berlin, Germany (Preisig & Andersen, 2005). The common genus of microalgae used including, Chlorella, Tetraselmis, Scenedesmus, Pavlova, Phaeodactylum, Chaetoceros, Nannochloropsis, Skeletonema and Thalassiosira. As mentioned by Brown et al. (1997), analyses of microalgae in his study found that microalgae have similar contents of essential amino acids as in fishmeal. Many microalgae species have been used in fish feed formulations to assess their nutritional value, and its advantages in replacing the main protein source. For examples, Chlorella fed to Korean rockfish (Bai et al., 2002) and Scenedesmus fed to Tilapia (Tartiel et al., 2008).

Scenedesmus sp. is a green microalgae classified under family Chlorophyceae. The availability of essential amino acids required in fish growth and the high level of crude protein in this species make it possible to substitute fish meal (Bawdy *et al.*, 2008). There are three major nutrients to be considered in microalgae to meet the needs of feed ingredients which are protein, carbohydrate and lipid. The nutritional value of protein component for microalgae is considered high if their essential amino acid composition is relatively close or similar to the requirements of the feeding animals (Misurcova *et al.*, 2014). *Scenedesmus* is a source of single cell protein up to 40 to 70% by converting the solar energy, as claimed by Nasseri *et al.* (2011). Carbohydrates in microalgae are found in the form of starch, cellulose, sugars and other polysaccharides. In microalgae, carbohydrates serves two main functions which are, a structural components in the cell walls and as storage components inside the cell (Raven and Beardall, 2004). Lipids are highly significant at various growth stages of many aquaculture animals including fish larvae. In green algae, the average lipid content is 23% while in *Scenedesmus* sp. is 26% (Rakesh *et al.*, 2016).

In addition, wastewater which is commonly discharged without treatment contained excess nutrients that are good for the growth of microalgae. Thus, due to the flexibility and adaptability of this microalgae towards cultivation in extreme conditions such as in wet market wastewater, has allowed it to undergo phycoremediation process which may results in microalgae biomass with a higher nutrient content that is ideal as a fish feed supplement and indirectly helped in reducing cost by lowering the use of fishmeal in the diet formulation (Mobin & Alam, 2014). Therefore, these strengthen that the phycoremediation process is good to improve the nutrient in microalgae to be used as fishmeal substitutes.



O. niloticus known as Nile tilapia, is one of the most cultured tropical fish worldwide (World Bank, 2013). O. niloticus possess five characteristics that make it as one of the selected fish species in the aquafarming industry which are high growth rate, versatility in environmental tolerance, stress and disease resistance, high reproductive rate and high acceptance for different diets (El-Sayed, 2006). The global production of this species was 4.5 million tonnes in 2013 and aimed to exceed 6.6 million tonnes by 2030 (FAO, 2014). Specific in Malaysia, a study by amal and Zamri-Saad (2011) has stated that, the production of O. niloticus reaches up to 20.8 metric tonnes. Moreover, this fish species is highly favoured in local and international market. Large scale production of *O. niloticus* through aquaculture practices would help to meet the demand of this species for both local supply and exportation. Overall, finding new alternative protein sources is important to minimize the problems of relying on fish meal and indirectly reduce cost in aquaculture practices. The phycoremediation process of Scenedesmus sp. in wetmarket wastewater would be one of the effective ways to improve the nutrient content in microalgae to replace fishmeal in fish feed diet formulations.

1.2 Problem statement

The ocean derived fishmeal and fish oil in aqua feeds has given rise to sustainability concerns as it will not meet the growing demand and will constraint aquaculture growth. As the aquaculture practices increased worldwide, the demand for fishmeal increases linearly, with prices rising by almost 300% in the past 10 years (Origin oil, 2014). It is inarguable that fishmeal is one of the best protein sources in the diet for fish, however, continuous exploitation of this natural resource will definitely lead to environmentally and economically unsustainable. Therefore, it is important to find alternative protein sources to replace fishmeal to fulfil the demand for fish in aquaculture production.

Currently, microalgae have received more attention because some of the species contain higher protein source that is almost similar to fishmeal. Comparing with animal by-product, microalgae is more preferable since animal by product requires the rendering process before using in the farm (Jedrejek *et al.*, 2016). Moreover, according to Shah *et al.* (2017), the composition of commercial microalgae species such as proximate analysis, amino acids and fatty acids and their mineral content is comparable to the available feed ingredients in the aqua feed industry (Kent *et al.*, 2015). Therefore, in this study, microalgae *Scenedesmus* sp. was cultivated in wet market wastewater as a protein replacer to partially replace fishmeal.



1.3 Significant of study

The State of World Fisheries and Aquaculture reported by FAO (2012) outlined that aquaculture is one of the fastest growing feed sectors, having almost 60 million tonnes globally for the past decades, and it is expected to grow up to 50% in the next years. According to FAO, a continuous expansion of the human population is predicted to pass nine billion by 2050, simultaneously the economic development gives a greater proportion with an income that allows them to choose their diet (Obach, 2012). Fishmeal has been used as a source of protein in the past decades in formulating artificial fish feed. About 36% of fisheries catch worldwide was processed into fishmeal to be incorporated into feeds for farmed fish every year (Ogello et *al.*, 2014). Tacon and Metian (2008) stated that a global survey estimated aquaculture

consumption of fishmeal is 3724 thousand tonnes in 2006. Aladetohun and Sogbesan (2013) added that, aquaculture practices are predicted to grow over the next 20 years indirectly would increase the demand for fishmeal and fish oil which indirectly may increase the pressure on the already threatened stocks of wild fish. Hence, the increasing demand of the production for aquatic animals must be supported by a corresponding increase in the production of formulated diets for culturing aquatic animals (Rahman et *al.*, 2013). It is undeniable that fishmeal would fulfil the requirement of essential amino acids of the farmed animals to maximize growth. However, the use of fishmeal should be controlled to ensure food security and sustainability of the food chain.

In Asia, fish feed production increased from 40% in 2000 to 60% in 2008 while the usage of fishmeal for tilapia culture risen from 0.8 million tonnes to 1.7 million tonnes during the same period (Tacon & Metian, 2008). In addition, fish feed accounts for the highest operational cost in aquaculture with protein being the most expensive diet (Munguti et *al.*, 2012) and fishmeal is the most expensive protein source in aquaculture feeds (Egerton *et al.*, 2020). Moreover, the availability of fish meal and formulated fish feed in Malaysia is low and depending on importation which cause increasing of the operating cost (Chun, 2007).



The usage of microalgae in aquaculture as feed is uncommon. Microalgae are known as promising living cells which are at the base of the aquatic food chain due to their good nutritional value (Sathasivam *et al.*, 2019). According to Allen and Nelson (1910), the first reports of the microalgae as feed in aquaculture practices were published since 1910, and the use of microalgae has developed widely (Duerr *et al.*, 1998; Santos-Ballardo *et al.*, 2015). The main application of microalgae in aquaculture could relate directly or indirectly to the nutrition impacts on many aquatic farmed organisms, for instance, rotifers and fish. Moreover, due to some characteristics such as lipids and carotenoids content, microalgae are known as the next generation of sustainable feedstock (Murray *et al.*, 2013; Perez-Lopez *et al.*, 2014).

In this study, freshwater microalgae from the genus *Scenedesmus* sp. was chosen to replace fishmeal in formulated fish feed. *Scenedesmus* sp. has a relatively similar essential amino acid profile to fishmeal. It is inarguable that additional cost will be used to produce formulated fish feed as Bold Basal's Medium (BBM) also will be used to cultivate *Scenedesmus* sp. However, the cultivation of *Scenedesmus* sp. using wet market wastewater will resulted in effluent with low nutrient and microalgae with higher nutrient composition. It is an economically fesible way to cultivate microalgae on a large scale. The ability of *Scenedesmus* sp. to perform phycoremediation would also ensure the nutrient recovery and indirectly prevent environmental pollution.

Therefore, the findings of this study are beneficial especially to the aquaculture and fish farming industry to enhance an increasing production of the eco- friendly formula of fish feed which consists of lower usage of fishmeal while maintaining the nutritional value for growth performance of *O. niloticus*.

1.4 Objective of study

The objectives of this study are:

- I. To compare and formulate fish feed with 2% replacement of fishmeal with *Scenedesmus* sp. cultivated in Bold Basal Medium and wet market wastewater.
- II. To study the physicochemical properties of the microalgae substitution cultivated in Bold's Basal Medium and wet market wastewater formulated feed.
- III. To investigate the effect of microalgae supplemented products on fish growth performance, and fatty acids content in *O. niloticus* muscle tissue.

1.5 Scope of study

- I. The study was focused on the formulation of feed with partial replacement of fishmeal with locally obtained *Scenedesmus* sp. that was cultivated in two different mediums which are standard medium and wet market wastewater.
- II. In this study, the physicochemical properties of the formulated feed produced was analysed and compared with the control formulation and the feed prototype was tested to *O. niloticus* fingerlings.
- III. The growth performance of *O. niloticus* and the water quality of the experimental tank was analysed weekly for 12 weeks. At the end of experimental period, the analysis on the fatty acids methyl ester in the muscle tissue of the fish was done to ensure a good quality of the fillet produced.

CHAPTER 2

LITERATURE REVIEW

2.1 Phycoremediation concept

The use of algae, either macroalgae or microalgae to treat wastewater has been discovered and practices for over 40 years (Ahmad *et al.*, 2013). Phycoremediation term was introduced by John (2000), which defines a bioremediation process performed by microorganisms such as microalgae (John, 2000; Gani *et al.*, 2015). Besides, the use of microalgae culture to treat wastewater is firstly studied by Oswald *et al.* (1957). These photosynthetic microorganisms in wastewater aid in the bio transform the pollutant loads in their cell and clean the wastewater before being released to the environment (Rawat *et al.*, 2011). According to Gani *et al.* (2016), there are some benefits of using microalgae in wastewater treatment which are effective in cost, environmentally friendly and not producing any toxic gases to the environment. Moreover, the algal biomass has the potential of high nutrient value, heavy metals removal and can be used as a live feed and formulated feed for both agriculture and aquaculture practices.

However, the removal efficiency of the pollutants by microalgae is depending on the species. According to Mohamed *et al.* (2017), phycoremediation of wastewater using microalgae occurs within 2 to 26 days to reduce pollutants to a minimum level. Only a few researchers have been done on phycoremediation of microalgae using various types of wastewater. Table 2. 1 below shows the study of using microalgae in wastewater treatment.



Based on Table 2.1, various microalgae species such as *Chlorella vulgaris*, *Nostoc commune*, *Scenedesmus* sp., *Botrycoccus* sp., and *Oscillatoria* sp. are used in wastewater for phycoremediation treatment. Latiffi *et al.* (2017) and Jais *et al.* (2015) studied the phycoremediation of meat processing wastewater and wet market wastewater by using the same type of microalgae which is *Scenedesmus* sp. while different studies on the physicochemical parameters' removal by *Botryococcus* sp. on two other type of wastewater which are dairy wastewater and Grey wastewater were conducted by Gani *et al.* (2015).

Sources	Phycoremediation summary	Reference
Meat processing wastewater	Preliminary assessment of growth rates on different concentrations of microalgae <i>Scenedesmus</i> sp. in industrial meat food	Latiffi et al. (2017)
Wet market wastewater	Removal of nutrients and selected heavy metals in wet market wastewater by using microalgae <i>Scenedesmus</i> sp	Jais <i>et al</i> . (2015)
Dairy and Grey wastewater	Phycoremediation of greywater and dairy wastewater by <i>Botryococcus</i> sp. for physiochemical parameters removal	Gani <i>et al</i> . (2015)
Sewage treatment plant wastewater	Application of <i>Chlorella vulgaris</i> for reduction of organic and inorganic pollutant in sewage treatment plant wastewater	Sahu, (2014)
Industrial wastewater	Application of microalgal bacterial flocs for industrial wastewater treatment using sequencing batch reactors	Hende <i>et al.</i> (2014)
Municipal wastewater	Phycoremediation of municipal wastewater using Oscillatoria and Nostoc commune obtained from Mula Mutha river	Azarpira <i>et al</i> . (2014)
Domestic wastewater	Domestic wastewater bioremediation using microalgae <i>Chlorella vulgaris</i> and <i>Scenedesmus quadricauda</i> for physiochemical reduction	Kshirsagar, (2013)

Table 2.1: Phycoremediation study using microalgae

The aforementioned study above, is almost similar to the research performed by Sahu (2014); Hende *et al.* (2014); Azarpira *et al.* (2014) and Kshirsagar (2013), which focus on the efficiency of the microalgae to remove the pollutants in the wastewater. It is common to study the wastewater treatment using microalgae. However, the applications of the by- product and its benefit should not be ignored. The illustration in Figure 2.1 shows the mechanism of wastewater treatment with the presence of microalgae, its biomass application, and the benefits of the wastewater after treatment.



Figure 2.1: Mechanisms of wastewater treatment using microalgae

The phycoremediation of wastewater using microalgae starts with organic matter which enhance the growth of bacteria and carbon dioxide released. The carbon dioxide is then used by microalgae for photosynthesis with the aid of light from the surrounding. Moreover, this photosynthetic microalga will treat the wastewater by absorbing all the excess nutrients inside the medium and the oxygen released to the environment will be cycled for bacterial oxidation. Commonly, the microalgae byproduct can be applied for many purposes such as for bio fertilization and as animal feed in aquaculture industry. Apart from that, the treated wastewater will ensure a discharge of wastewater into the aquatic ecosystem such as rivers and lakes with a low

REFERENCES

- Aas, T. S., Terjesen, B. F., Sigholt, T., Hillestad, M., Holm, J., Restfie, S., Baeverfjord, K., Rorvik, A., Sorenson, M., Oehme, M. & Asgard, T. (2011). Nutritional responses in rainbow trout (*Oncorhynchus mykiss*) fed diets with different physical qualities at stable or variable environmental conditions *Aquaculture Nutrition*, *17*, 657-670
- Abdel-Tawwab, M., Mousa, M. A. A., Ahmad, M. H. & Sakr, S. F. M. (2007). The use of calcium pre-exposure as a protective agent against environmental copper toxicity for juvenile Nile tilapia, *Oreochromis niloticus*. *Aquaculture*, 264, 236–246
- Abiona, O., Awojide, S. & Odesanmi, O. (2018). Assessment of heavy and trace metal contents of internal organs of tilapia (*Oreochromis niloticus*) and catfish (*Clarias gariepinus*). *Journal of Life Physical Science*, 10 (1), 61–66
- Agbozu, I. E., Ekweozor, I. K. E. & Opuene, K. (2007). Survey of heavy metals in the catfish synodontis claria. *International Journal of Environmental Science and Technology*, 4 (1), 94-97
- Ahmad, F., Khan, A. U. & Yasar, A. (2013). Comparative phycoremediation of sewage water by various species of algae. *Pakistan Academy Science*, 50, 131–139
- Aladetohun, N. F. & Sogbesan, O. A. (2013). Utilization of blood meal as a protein ingredient from animal waste product in the diet of Oreochromis niloticus. *International Journal of Fisheries and Aquaculture*, 5 (9), 234 – 237
- Ali, A. & Al-Asgah, N. A. (2001). Effect of feeding different carbohydrate to lipid ratios on the growth performance and body composition of Nile tilapia
 Amal, M. N. A. & Zamri-Saad, M. (2011). Streptococcosis in tilapia (*Oreochromis niloticus*): a review. *Pertanika Journal Tropical Agricultural Science*, 34 (2), 195-206

- Allen, E. J. & Nelson, E. W. (1910). The artificial culture of marine plankton organisms. *Journal of Marine Biology Association*, 8421-8474
- Allen, K. (2016). Evaluating Spirulina as a protein source in Nile Tilapia (*Oreochromis niloticus*) grow-out Diets. Queen's University Kingston: Master's Project Report
- Amal, M. N. A. & Zamri-Saad, M. (2011). Streptococcosis in tilapia (Oreochromis niloticus): A review. Pertanika Journal Tropical Agricultural Science, 34 (2), 195-206
- Amankwaah, D., Cobbina, S. J., Tiwaa, Y. A., Bakobie, N. & Millicent, E. A. B. (2014). Assessment of pond effluent effect on water quality of the .Asuofia Stream, Ghana. *African Journal of Environmental Science and Technology*, 8 (5), 306-311
- Anyanwu, L. T. & Solomon, J. R. (2015). Physiochemical parameters of fish pond use for Dutch Clarias hybrid fed coconut chaff and Bambara nuts. *Aquaculture*, 11, 1-10
- Association of Official Analytical Chemists (AOAC). (1990). Official Methods of Analysis of the Association of Official Analytical Chemists, 15th ed. Virginia: Association of Official Analytical Chemist
- Association of American Feed Control Officials. (2000). Retrived from <u>http://www.fao.org/3/y1453e0m.htm</u>
- Atwood, H. L., Fontenot, Q. C., Tomasso, J. R. & Isely, J. J. (2001). Toxicity of nitrite to Nile tilapia: effect of fish and environmental chloride. *North American Journal of Aquaculture*, 63, 49-51
- Azarpira, H., Behdarvand, P., Dhumal, K. & Pondhe, G. (2014). Potential use of cyanobacteria species in phycoremediation of municipal wastewater. *International Journal of Bioscience*, (4) 4, 105-111
- Azaza, M. S., Dhraief, M. N., Kraiem, M. M. & Baras, E. (2010). Influences of food particle size on growth, size heterogeneity, food intake and gastric evacuation in juvenile Nile tilapia, *Oreochromis niloticus*, L., 1758. *Aquaculture*, 309, 193- 202
- Azzaydi, M., Martines, F. J., Zamora, S., Sanchez-Vazquez, F. J. & Madrid, J. A. (2000). The influence of nocturnal vs. diurnal feeding condition under winter condition on growth and feed conversion of European sea bass, *Dicentrarchus labrax. Aquaculture*, 182, 329–38

- Badr, A. M., Mahana, N. A. & Eissa, A. (2014). Assessment of Heavy metal levels in water and their toxicity in some tissues of Nile Tilapia (*Oreochromis niloticus*) in River Nile Basin at Greater Cairo, Egypt. Global. Veterinarian, 13 (4), 432-443
- Baeverfjord, G., Refstie, S., Krogedal, P. & Asgard, T. (2006). Low feed pellet water stability and fluctuating water salinity cause separation and accumulation of dietary oil in the stomach of rainbow trout (*Oncorhynchus mykiss*). Aquaculture, 261, 1335-1345
- Baharuddin, N. N. D. E., Aziz, N. S., Sohif, H. N., Karim, W. A. A., Al- Obaidi,
 J. R. & Basiran, M. N. (2016). Marine microalgae flocculation using plant:
 The case of Nannochloropsis oculata and Moringa oleifera. *Pakistan Journal* of Biotechnology, 48 (2), 831-840
- Bai, S. C., Choi, S. M. & Wang, X. J. (2002). Apparent protein and phosphorus digestibilities of five diferent dietry protein sources in Korean rockfish, *Sebastes schlegeli* (Hilgendorf). *Aquaculture Research*, 32 (1), 99-105
- Bailey. J., Alanara, A. & Crampton, V. (2003). Do delivery rate and pellet size affect growth rate in Atlantic salmon (*Salmo salar* L.) raised under semicommercial farming conditions. *Aquaculture*, 224, 79–88
- Bawdy, T. M., Ibrahim, E. M. & Zeinhom, M. M. (2008). Partial replacement of fishmeal with dried microalga (*Chlorella* spp and *Scenedesmus* spp) in Nile tilapia (*Oreochromis niloticus*) diets. *Proc. Of the 8th Int. Conf. on Tilapia in Aquaculture*. Cairo, Egypt. pp. 801-811
- Becker, E. (2007) Microalgae as a source of protein. *Biotechnology Advanced*, 25, 207–210
- Becker. W. (2004). *Microalgae in Human and Animal Nutrition*. Cambridge: Blackwell Science
- Begum, A., Mondal, S., Ferdous, Z., Zafar, M. A. & Ali, M. M. (2014). Impact of water quality parameters on monosex tilapia (*Oreochromis niloticus*) production under pond condition. *International Journal Animal Science*, 2 (1), 14-21
- Beveridge, M. C. M. & McAndrew, B. J. (2000). *Tilapias: Biology and Exploitation*. Dordrecht: Kluwer Academic Publishers

- Bhatnagar, A. & Singh, G. (2010). Culture fisheries in village ponds: a multilocation study in Haryana, India. Agriculture and Biology Journal of Norrth America, 1 (5), 961–968
- Boonanuntanasaran, S., Jangprai, A., Kumkhong, S., Plagnes-Juan, E., Veron, V.,
 Burel, C., Marandel, L. & Panserat, S. (2018). Adaptation of Nile tilapia (*Oreochromis niloticus*) to different levels of dietary carbohydrates: New insights from a long term nutritional study. Aquaculture, 496, 58-65
- Boyd, C. E. (1998). Water Quality for Pond Aquaculture, Research and Development Series. Alabama: Auburn University
- Boyd, E. C. (2004). *Farm-Level Issues in Aquaculture Certification: Tilapia*. United States: Report commissioned by WWF
- Bringas, S. C., Plassen, L., Lekang, O. I. & Schuller, R. B. (2007). Measuring physical quality of pelleted feed by texture profile analysis, a new pellet tester and comparisons to other common measurement devices. *Annual Transaction* of The Nordic Rheology Society, 15
- Brown, M. R., Jeffrey, S. W., Volkman, J. K. & Dunstan, G. A. (1997). Nutritional properties of microalgae for marineculture. *Aquaculture*, 151, 315-331
- Brown, M., Robert, R. (2002). Preparation and assessment of microalgal concentrates as feeds for larval and juvenile Pacific oyster (*Crassostrea gigas*). Aquaculture, 207, 289-309
- Caldini, N. N., Cavalcante, D. D. H., Filho, P. R. N. R. & Docarmo, M.V. (2015).
 Feeding Nile Tilapia with artificial diet and dried bioflocs biomass. *Animal Science*, *37* (*4*), 335-341
- Camacho- Rodriguez, J., Macias- Sanchez, M. D., Ceron- Garcia, M. C., Alarcon, F. J. & Molina- Grima, E. (2017). Microalgae as a potential ingredient for partial fishmeal replacement in aquafeeds: nutrient stability under different storage conditions. *Journal of Applied Phycology*, *30* (2), 1049-1059
- Canonico, G. C., Arthington, A., McCrary, J. K. & Thieme, M. L. (2005). The effects of introduced tilapias on native biodiversity. *Aquatic Conservation Marine and Freshwater Ecosystems*, 15 (5), 463-483

73

- Castro- Gonzalez, M. I. & Mendez- Armenta, M. (2008). Heavy metals: implications associated to fish consumption. *Environmental Toxicology and Pharmacology*, 26, 263- 271
- Celik, E. (2012). *Tilapia Culture Review*. Norwegian University of Life Science: Master's Thesis
- Cerezuela, R., Guardiola, F. A., Meseguer, J. & Esteban, M. A. (2012). Enrichment of gilthead seabream (*Sparus aurata* L.) diet with microalgae: Effects on the immune system. *Fish Physiology and Biochemistry*, 38 (6), 1729-1739
- Charo-Karisa, H. (2006). Selection for growth of Nile Tilapia (Oreochromis niloticus) in low-inputs environments. Wageningen University: PhD Thesis
- Chen, G., Zhao, L., Qi, Y. & Lu Cui, Y. (2014). Chitosan and its derivative applied in harvesting microalgae or biodiesel production: An outlook. *Journal of Nanomaterials*, 1-9
- Chun, K. H. (2007). Comparison of nutritional values of small- scale processed and commercial fishmeal for marine fish culture in Sabah, Malaysia. Universiti Malaysia Sabah: Master's Project
- Daniel, S., Larry, W. D. & Joseph, H. S. (2005). Comparative oxygen consumption and metabolism of striped bass (Moronesaxaatilis) and its hybrid. *Journal* of World Aquaculture Society, 36 (4), 521-529

Department of Fisheries Malaysia (2013). Penyelidikan Makanan Ikan. Putrajaya: Department of Fisheries Malaysia. Retrived from <u>http://www.dof.gov.my/</u>

Department of Fisheries Malaysia (2015). Penyelidikan Makanan Ikan. Putrajaya: Department of Fisheries Malaysia. Retrieved from <u>http://www.dof.gov.my/</u>

Deswati, Suyani, H., Muchtar, A. K., Abe, E. F., Yusuf. Y. & Pardi, H. (2019). Copper, Iron and Zinc contents in water, Pakcoy (*Brassica rapa*) and Tilapia (*Oreochromis niloticus*) in the presence of aquaponics. *Rasayan Journal of Chemistry*, 12 (1), 40-49

- Devi, P. A., Padmavathy, P., Aanand, S. & Aruljothi, K. (2017). Review on water quality parameters in freshwater cage fish culture. *International Journal of Applied Research*, *3* (5), 114-120
- Dhaneesh, K. V., Noushad, K. M. & Kumar, T. T. A. (2012). Nutritional evaluation of commercially important fish species of Lakshadweep Archipelago, India. *PLOS One*, 7 (9)

- Dong, G. (2015). Wastewater sampling and characteristion-raw sewage monitoring and result analysis. Proc. of the 9th Int. Conf. on Annual Water Industry Operations Conference & Exhibition. NSW Public Works. pp. 40
- Dragonovic, V., Goot, A. J., Boom, R. & Jonkers, J. (2011). Assessment of the effects of fishmeal, wheat gluten, soy protein concentrate and feed moisture on extruder system parameters and the technical quality of fish feed. *Animal Feed Science and Technology*, 165, 238-250
- Duerr, E. O., Molnar, A. & Sato, V. (1998). Cultured microalgae as aquaculture feeds. *Journal of Marine Biotechnology*, 6, 65-70
- Dullah, H., Malek, M. A. & Hanafiah, M. M. (2020). Life cycle assessment of Nile Tilapia (*Oreochromis Niloticus*) farming in Kenyir Lake, Terrenganu. *Sustainability*, 12, 2268
- Duncan, P. L., Klesius, P. H. (1996). Effects of feeding *Spirulina* on specific and non-specific immune responses of channel catfish. *Journal of Aquatic Animal Health*, 8, 308-313
- Duong, V. T., Ahmed, F., Thomas-Hall, S. R., Quigley, S., Nowak, E. & Schenk,
 P. M. (2015). High protein- and high lipid- producing microalgae from northern Australia as potential feedstock for animal feed and biodiesel. *Frontiers in Bioengineering and Biotechnology*, 3 (53), 53-59
- Durmus, M. (2018). Fish oil for human health: omega- 3 fatty acid profiles of marine seafood species. *Food Science and Technology*, 1-8
- Egerton, S., Wan, A., Murphy, K., Collins, F., Ahern, G., Sugrue, I., Busca, K.,
 Egan, F., Muller, N., Whooley, J., McGinnity, P., Culloty, S., Ross, R. P. &
 Stanton, C. (2020). Replacing fishmeal with plant protein in Atlantic salom
 (Salmo salar) diets by supplementation with fish protein hydrolysate. *Scientific Reports*, 10 (4194)
- Enyidi, U., Pirhonen, J., Kettunen, J. & Vielma, J. (2017). Effect of Feed Protein:
 Lipid Rtio on Growth Parameters of African Catfish *Clarias gariepinus* after
 Fish Meal Substitution in the Diet with Bambaranut (*Voandzeia subterranea*)
 Meal and Soybean (*Glycine max*) Meal. *Fishes*, 2, 1
- Ekubo, A. A. & Abowei, J. F. N. (2011). Review of some water quality management principles in culture fisheries. *Resource Journal of Applied Science, Engineering and Technology*, 3 (2), 1342

El-Sayed, A. F. M. (2006). Tilapia Culture. Cambridge: CABI Publishing

- El-Serafy, S. S., El-Ezabi, M. M., Shehata, T. M. & Esmael, N. (2007). Dietary iron requirement of the Nile tilapia, Oreochromis niloticus (L.) fingerlings in intensive fish culture. Egypt Journal of Aquatic Biology & Fish, 11(2), 165-184
- Emparan, Q., Harun, R. & Danquah, M. K. (2019). Role of phycoremediation for nutrient removal from wastewater: A review. Applied Ecology and Envionmental Research, 17 (1), 889-915
- Fabregas, J. & Herrero, C. (2003). Marine microalgae as a potential source of minerals in fish diets. *Aquaculture*, 51 (33), 237-243
- Fadl, S. E., El- Habashi, N.g Gad, D. M., Elkassas, Z. I. E., Abdelhady, D. H. & Hegazi, S. M. (2019). Effect of adding Dunaliella algae to fish diet on lead acetate toxicity and gene expression in the liver of Nile tilapia. *Toxin Review*, 1-14
- Fernandes, C. E., Da Silva Vasconcelos, M. A., De Almeida Ribeiro, M., Sarubbo, L. A., Andrade, S. A. C. & De Melo Filho, A. B. (2014). Nutritional and lipid profiles in marine fish species from Brazil. *Food Chemistry*, 160, 67–71
- Food and Agricultural Organization. (2012). *The State of World Fisheries and Aquaculture*. Rome: Food and Agriculture Organization of the United Nations
- Food and Agricultural Organization of the United Nations. (2014). *The State of World Fisheries and Aquaculture*. Rome: Food and Agriculture Organization of the United Nations
- Food and Agricultural Organization. (2004), *The state of world fisheries and aquaculture 2004*. Rome: Food and Agriculture Organization of the United Nations
- Food and Agricultural Organization. (2017). *Aquaculture feed and Fertilizer Resources Information System*. Rome: Food and Agricultural Organization of the United Nations.
- Food and Agricultural Organization. (2018). *The state of world fisheries and Aquaculture: Meeting the Sustainable Development Goals*. Rome: Food and Agriculture Organization of the United Nations



- Food and Agriculture Organization. (2002). FAO Technical Guidelines for Responsible Fisheries, 5, Supplement 1. Rome: Food and Agriculture Organization of the United Nations
- Food and Agriculture Organization of the United Nations. (2019). Aquaculture feed and Fertilizer Resources Information System. Retrived from http://www.fao.org/fileadmin/user_upload/affris/img/Niletilapia_table/table _10.jpg
- Gall, J. E., Boyd, R. S. & Rajakaruna, N. (2015). Transfer of heavy metals through terrestrial food webs: A review. *Environmental Monitoring and Assessment*, 187, 201
- Gani, P., Sunar, N. M., Matias-Peralta, H., Jamaian, S. S. & Latiff, A.A.A. (2016).
 Effects of different culture conditions on the phycoremediation efficiency of domestic wastewater, *Journal of environmental chemical engineering*, 4 (4), 4744-4753
- Gani, P., Sunar, N. M., Matias-Peralta, H. M., Latiff, A. A. & Razak, A. R. A. (2016a). Influence of initial cell concentrations on the growth rate and biomass productivity of microalgae in domestic wastewater. *Applied Ecology* and Environmental Research, 14(2), 399-409.

Gani, P., Sunar, N. M., Matias-Peralta, H., Latiff, A. A. A., Parjo, U. K. & Razak,
A. R. A. (2015). Phycoremediation of wastewaters and potential hydrocarbon
from microalgae: A review. *Advances in Environmental Biology*, 9 (20), 1-8

Ganuza, E. (2016). Microalgae as a Mineral Vehicle in Aqua feeds. U.S. Patent

- Gladyshev, M., Lepskaya, E. V., Sushchik, N. N. & Makhutova, O. N. (2012). Comparison of Polyunsaturated Fatty Acids content in fillets of Anadromous and Landlocked Sockeye Salmon Oncorhynchus Nerka. *Journal of Food Science*, 77 (12), 1307-1310
- Gorgun, S. & Akpinar, M. A. (2012). Effect of season on the fatty acid composition of the liver and muscle of *Alburnus chalcoides* from Todurge Lake (Sivas, Turkey). *Turkish Journal of Zoology*, *36* (5), 691–698
- Gorlach-Lira, K., Pacheco, C., Carvalho, L. C. T., Junior, H. N. M. & Crispim, M.
 C. (2013). The influence of fish culture in floating net cages on microbial indicators of water quality. *Brazilian Journal of Biology*, 73 (3), 457-463



- Green, A. J. & Planchart, A. (2018). The neurological toxicity of heavy metals: A fish perspective. Comparative biochemistry and physiology part c. *Toxicology and Pharmacology*, 208, 12-19
- Guedes, A. C. & Malcata, F. X. (2012). Nutritional Value and Uses of Microalgae in Aquaculture. Croatia: Universiti Campus Step Slavka Krautzeka
- Guiry, M. D. & Guiry, G. M. (2015). Algae Base. World-wide electronic publication, National University of Ireland. Retrieved from http://www.algaebase.org
- Guiry, M. D. & Guiry, G. M. (2014). AlgaeBase. World-wide electronic publication, National University of Ireland. Retrived from <u>www.algaebase.org</u>
- Gyimah, E., Akoto O. & Nimako, C. (2018). Health risk assessment of heavy metals contamination in edible fish species from the Barekese Reservoir in Kumasi, Ghana. American Journal of Environmental Science, 2 (1), 1-7
- HACH. (1979). Standard Method for the Examination of Water and Wastewater. USA: HACH
- Hamzah, A., Nguyen, N. H. & Ponzoni, R. W. (2008). Performance and survival of three red Tilapia strains (*Oreochromis* spp.) in pond environment in Kedah State, Malaysia. *Proc. of the 8th Int. Conf. on Tilapia in Aquaculture*. Kedah, Malaysia. pp. 199-211
- Hattab, M. A. & Ghaly, A. (2014). Effects of Light Exposure and Nitrogen Source on the Production of Oil from Freshwater and Marine Water Microalgae. American *Journal of Biochemistry and Biotechnology*, *10* (4), 211-233
- Hende, S. V. D., Carre, E., Cocaus, E., Beelen, V., Boon, N. & Vervaeren, H. (2014). Treatment of industrial wastewaters by microalgal bacterial flocs in sequencing batch reactors. *Bioresource Technology*, 161, 245-254
- Hosseini Tafreshi, A. & Shariati, M. (2009). Dunaliella biotechnology: methods and applications. *Journal of Applied Microbiology*, *107* (1), 14-35
- Hu, C. J., Chen, S. M., Pan, C. H. & Huang, C. H. (2006). Effects of dietary vitamin A or b-carotene on growth of juvenile hybrid tilapia, *Oreochromis niloticus* x *O. aureus*. *Aquaculture*, 253, 602-607
- Ibrahim, A. Z., Hassan, K. H., Kamarudin, R. & Anuar, A. R. (2018). Measuring sustainable livelihood for Malaysia's poor: the sustainable livelihoods Index

and the B40 group of coastal fishermaen in northen Peninsular Malaysia. International Journal of Society, 2, 39-47

- Ibrahem, M. D., Mohamed, F. M., Marwa, A. & Ibrahim, M. A. (2013). The role of *Spirulina platensis (Arthrospira platensis)* in growth and immunity of Nile tilapia (*Oreochromis niloticus*) and its resistance to bacterial infection. *Journal of Agricultural Science*, 5 (6), 109-117
- Ighwela, K. A., Ahmad, A. & Abol- Munafi, A. B. (2013). Water stability and nutrient leaching of different levels of maltose formulated fish pellets. *Global Veterinaria*, *10* (6), 638-642
- International Union of Pure and Applied Chemistry (IUPAC). (1987). Preparation of Fatty Acid Methyl Ester in Standard Methods for Analysis of Oils, Fats and Derivative, 7th ed. Oxford: Blackwell
- Jabeen, F. & Chaudhry, A. S. (2011). Chemical compositions and fatty acid profiles of three freshwater fish species. *Food Chemistry*, *125*, 991-996
- Jais, N, M., Mohamed, R. M. S. R., Apandi, W. A. W. M. & Peralta. H. M. M. (2015). Removal of nutrients and selected heavy metals in wet market wastewater by using microalgae *Scenedesmus* sp. *Applied Mechanics and Materials*, 773-774
- Jaya-Ram, A., Fuad, F., Zakeyuddin, M. S. & Md Shah, A. S. R. (2018). Muscle fatty acid content in selected freshwater fish from Bukit Merah reservoir, Perak, Malaysia. *Tropical Life Sciences Research*, 29 (2), 103-117
- Jedrejek, D., Levic, J., Wallace, J. & Oleszek, W. (2016). Animal by product for feed: Characteristics, European regulatory framework, and potential impacts on human and animal health and the environment. *Journal of Animal and Feed Sciences*, 25 (3), 189-202
- John, J. (2000). A Self-Sustainable Remediation System for Acidic Mine Voids. *Proc. Of the Fourth Int. Conf. on Diffuse Pollution*. pp. 506–11
- Kandyliari, A., Mallouchos, A., Papandroulakis, N., Golla, J. P., Lam, T. T., Sakellari, A., Karavoltsos, S., Vasilis, V. & Kapsokefalou, M. (2020).
 Nutrient composition and fatty acid and protein profiles of selected fish by-products. *Foods*, 9 (2), 190



- Karthikeyan, S. (2012). A critical review: microalgae as a renewable source for biofuel production. *International Journal of Engineering Research and Technology*, 1, 1-6
- Kasozia, N., Tandlichc, R., Fickd, M., Kaisere, H. & Wilhelmia, B. (2019). Iron supplementation and management in aquaponic systems: A review. *Aquaculture Reports*, 15
- Kay, R. A. (1991). Microalgae as food and supplement. Critical reviews. Food Science and Nutrition, 30, 555-573
- Kent, M., Welladsen, H. M., Mangott, A. & Li, Y. (2015) Nutritional evaluation of Australian microalgae as potential human health supplements. *PLoS One*, 10 (2), 1-14
- Khandaker, M. U., Heffny, N. A., Amin, Y. M. & Bradley, D. A. (2019). Elevated concentration of radioactive potassium in edible algae cultivated in Malaysian seas and estimation of ingestion dose to humans. *Algal Resource*, 38
- Kim, S. S, Rahimnejad, S., Kim, K. W. & Lee, K. J. (2013) Partial replacement of fishmeal with Spirulina pacifica in diets for parrot fish (Oplegnathus fasciatus). Turkish Journal of Fisheries and Aquatic Science, 13, 197–204
- Kinsella, J., Shimp, J., Mai, J. & Weihrauch, J. (1977). Fatty acid content and composition of freshwater finfish. *Journal of the American Oil Chemists Society*, 54 (10), 424-429
- Kortei, N. K., Heymann, M. E., Essuman, E. K., Kpodo, F. M., Akonor, P. T., Lokpo, S. Y. & Tettey, C. (2020). Health risk assessment and levels of toxic metals in fishes (*Oreochromis niloticus* and *Clarias anguillaris*) from Ankobrah and Pra basins: Impact of illegal mining activities on food safety. *Toxicology Reports*, 7, 360-369
- Kris-Etherton, P. M., Grieger, J. A. & Etherton, T. D. (2009). Dietary reference intakes for DHA and EPA. *Prostaglandins*, *Leukotrienes and Essential Fatty Acids*, 81, 99–104
- Kshirsagar, A. D. (2013). Bioremediation of wastewater by using microalgae: An experimental study. *International Journal of Life Sciences Biotechnology and Pharma Research*, 2 (3), 339-346

- Lang, I., Hodac, L., Friedl, T. & Feussner, I. (2011). Fatty acid profiles and their distribution patterns in microalgae: a comprehensive analysis of more than 2000 strains from the SAG culture collection. *BMC Plant Biology*, 11, 124
- Latiffi, N. A. A., Mohamed, R. M. S. R., Apandi, N. & Tajuddin, R. M. (2017). Preliminary Assessment of Growth Rates on Different Concentration of Microalgae Scenedesmus sp. in Industrial Meat Food. Protozoology, 103 (1)
- Leaf, A. & Weber, P. C. (1988). Cardiovascular effects of n-3 fatty acids. *The New Journal of Medicine*, 318
- Lee, J. Y. (2003). Vitamin K requirements of juvenile tilapia, Oreochromis niloticus x O. aureus and grouper, Epinephelus malabaricus. National Taiwan Ocean University: Master's thesis
- Lee, S., Moniruzzaman, M., Bae, J., Seong, M., Jin Song, Y., Dosanjh, B. & Bai, S. C. (2016). Effects of extruded pellet and moist pellet on growth performance, body composition, and hematology of juvenile olive flounder, and *Paralichthys olivaceus*. *Fisheries and Aquatic Sciences*, 32
- Lieke, T., Meinelt, T., Hoseinifar, S. H., Pan, B., Straus, D. L. & Steinberg, C. E.
 W. (2019). Sustainable aquaculture requires environmental friendly treatment strategies for fish disease. *Reviews in Aquaculture*, 12 (2), 943-965
- Lemarie, G., Dosdat, A., Coves, D., Dutto, G., Gasset, E. & Person-Le Ruyet, J. (2004). Effect of chronic ammonia exposure on growth of European seabass (*Dicentrarchus labrax*) juveniles. *Aquaculture*, 229, 479-491
- Li, G., Sinclair, A. J. & Li, D. (2011). Comparison of lipid content and fatty acid composition in the edible meat of wild and cultured freshwater and marine fish and shrimps from China. *Journal of Agricultural and Food Chemistry*, 59 (5), 1871-1881
- Li, J., Fan, Z., Qu, M., Qiao, X., Sun, J., Bai, D. & Cheng, Z. (2015). Applications of microalgae as feed additives in aquaculture. *International Symposium on Energy Science and Chemical Engineering*, 352-356
- Li, M. & Huang, C. (2016). Effect of dietary zinc level on growth, enzyme activity andbody trace elements of hybrid tilapia, *Oreochromis niloticus* x O. *aureus*, fed soya bean meal-based diets. *Aquaculture Nutrition*, 22, 1320–1327

- Li, Y., Bordinhon, A. M., Davis, D. A., Zhang, W. & Zhu, X. (2012). Protein: energy ratio in practical diets for Nile tilapia *Oreochromis niloticus*. *Aquaculture International*, 21 (5), 1109-1119
- Litchkoppler, B. F. (1999). *Water Quality Management for Pond Fish Culture*. New York: El Sevier Scientific Publishing Company
- Lim, C. E. & Webster, C. D. (2006). *Tilapia: Biology, Culture and Nutrition*. New York: Food Products Press
- Lim, C., Yildirim, M. & Klesius, P. (2011). Lipid and Fatty acids requirements of Tilapia. North American Journal of Aquaculture, 73, 188-193
- Liu, H, Liu, D. & Yao, M. (2004). Fishery Sciences. Chinese Academy of Fishery Science of China. 3, 15- 25
- Lorenzen, K. (2000). Population Dynamics and Management. Klewer Academic Publishers. pp 163-225
- Madhumathi, M. & Rengasamy, R. (2011). Antioxidant status of *Penaeus* monodon fed with *Dunaliella salina* supplemented diet and resistance against WSSV. International Journal of Engineering, Science and Technology, 3, 7249–7259
- Makwinja, R. & Geremew, A. (2020). Roles and requirements of trace elements in tilapia nutrition: Review. *Egyptian Journal of Aquatic Research*, 46 (3), 281-287
- Makwinja, R., Kapute, F., Singini, W. & Zidana, H. (2015). Effect of dietary protein levels on ammonia concentration and growth of *Tilapia rendalli* raised in concrete tanks. *International Journal of Fisheries and Aquaculture*, 7 (11), 178-184
- Malaysia Food Act and Regulation. (1985). *Malaysian Law on Food and Drugs*. Malaysia: Malaysian Law Publishers
- Medina-Felix, D., Lopez-Elias, J. A., Martinez-Cordova, L. R., Lopez-Torres, M. A., Hernandez-Lopez, J., Rivas-Vega, M. E. & Mendoza-Cano, F. (2014) Evaluation of the productive and physiological responses of Litopenaeus vannamei infected with WSSV and fed diets enriched with Dunaliella sp. *Journal Invertebrate Pathology*, 117, 9–12

- Misurcova, L., Bunka, F., Vavra Ambrozova, J., Machu, L., Samek, D. & Kracmar, S. (2014). Amino acid composition of algal products and its contribution to RDI. *Food Chemistry*, 151, 120-125.
- Mjoun, K., Rosentrater, K. & Brown, M. L. (2010). Tilapia: environmental biology and nutrition requirements. Fact Sheets, paper 164
- Mobin, S., & Alam, F. (2014). Biofuel production from algae utilizing wastewater. Proc. of the 19th Int. Conf. on Australian Fluid Mechanics. Melbourne, Australia. pp. 8-11
- Mohamed, R. M., Al-Gheethi, A. A., Aznin, S. S., Hasila, A. H., Wurochekke, A. A. & Kassim, A. H. (2017). Removal of nutrients and organic pollutants from household greywater by phycoremediation for safe disposal. *International Journal of Energy Environment Engineering*, 8, 259–272
- Munguti, J. M., Charo- Karisa, H., Opiyo, M. A., Ogello, E. O., Marijani, E. & Nzayisenga, L. (2012) Nutritive value and availability of commonly used feed ingredients for farmed Nile Tilapia (Oreochromis niloticus) and African catfish (Clarias gariepinus, burchell) in Kenya, Rwanda and Tanzania. *African Journal of Food Agriculture, Nutrition and Development*, *12* (*3*), 1–22
- Murray, P. M., Moane, S. & Collins, C. (2013). Sustainable production of biologically active molecules of marine based origin. *New Biotechnology*, *30* (6), 839-85
- Nandeesha, M. C., Gangadhar, B., Varghese, T. J. & Keshavanath, P. (1998). Effect of feeding Spirulina platensis on the growth, proximate composition and organoleptic quality of common carp, *Cyprinus carpio* L. *Aquaculture Research*, 29, 305-312
- Nasseri, A. T., Rasoul- Amini, S., Morowvat, M. H. & Younes, G. (2011). American Journal of Food and Technology, 6.
- National Research Council (1993). Nutrient Requirements of Fish. Washington: National Academy Press
- Ng, W. & Romano, N. (2013). A review of the nutrition and feeding management of farmed tilapia throughout the culture cycle. *Reviews in Aquaculture*, 5 (4), 220-254

- Ng, W. K. (2009). The current status and future prospects for the aquaculture industry in Malaysia. *World Aquaculture*, 40 (3), 26-30
- Nielsen, S. (2014). Food Analysis. New York: Springer
- Noman, S. (2018). *Use of Spirulina in Fish Culture*. Master's thesis. Bangabandhu Sheikh Mujibur Rahman Agricultural University Salna: Master's Thesis
- Nyanti, L., Hii, K. M., Sow, A., Norhadi, I. & Ling, T. Y. (2012). Impacts of aquaculture at different depths and distances from cage culture sited in Batang Ai Hydroelectric Dam Reservoir Sarawak, Malaysia. World Applied Sciences Journal, 19 (4), 451-456
- Obach, A. (2012). Options and challenges of Alternative Protein an Energy Resources for Aquafeed. Norway: Skretting Aquaculture Research Centre
- Obirikorang, K. A., Amisah, S., Fialor, S. C. & Skov, P. V. (2015). Effects of dietary inclusions of oilseed meals on physical characteristics and feed intake of diets for the Nile tilapia, *Oreochromis niloticus*. *Aquaculture Reports*, 1, 43-49
- Oehme, M., Aas, T. S., Olsen, H. J., Sorensen, M., Hillestad, M., Li, Y. & Asgard, T. (2012). Effects of dietary moisture content of extruded diets on physical feed quality and nutritional response in Atlantic salmon (*Salmo salar*). *Aquaculture nutrition*, 20 (4), 451-465
- Ogello, E. O., Munguti, J. M., Sakakura, Y. & Hagiwara, A. (2014). Complete replacement of fishmeal in the diet of Nile Tilapia (Oreochromis niloticus) grow-out with alternative protein sources. *International Journal of Advanced Research*, 2 (8), 962- 978
- Origin Oil. (2014). Origin Oil White Paper Algae as Aquafeed. Retrived from http://www.originclear.com/pdf/OriginOil-White-Paper-Algae-AsAquafeed.pdf
- Oswald, W. J. & Gotaas, H. B. (1957). Photosynthesis in sewage treatment. *Transaction of the American Society of Civil Engineering*, 122, 73–105
- Ozogul, Y., Ozogul, F. & Erguden, S. A. (2007). Fatty acids profiles and fat contents of commercially important seawater and freshwater fish species of Turkey: A comparative study. *Food Chemistry*, 103, 217-223

- Ozparlak, H. (2013). Effect of seasons on fatty acid composition and n-3/n-6 ratios of muscle lipids of some fish species in Apa Dam Lake, Turkey. *Pakistan Journal of Zoology*, 45 (4), 1027-1033
- Paolucci, M., Fasulo, G. & Volpe, M. G. (2015). Employment of marine polysaccharides to manufacture functional bio composites for aquaculture feeding applications. *Marine Drugs*, 13, 2680-2693
- Parvin, A., Hossain, M. K., Islam, S., Das, S. S., Munshi, J. L., Suchi, P. D., Moniruzzaman, M., Saha, B. & Mustafa, M. G. (2019). Bioaccumulation of heavy metals in different tissues of Nile Tilapia (*Oreochromis niloticus*) in Bangladesh. *Malaysian Journal of Nutrition*, 25 (2)
- Patel, S. R. J. (2014). Optimization for Production of Fish Protein Hydrolysate from Nile Perch (Lates niloticus) by-products. University of Nairobi: Master's Project Report
- Patterson, D., Delbert, M. & Gatlin, D. M. (2013) Evaluation of whole and lipidextracted algae meals in the diets of juvenile red drum (*Sciaenops* ocellatus). Aquaculture, 416-417
- Patnaik, R., Singh, N. K., Bagchi, S. K., Rao, P. S. & Mallick, N. (2019).
 Utilization of Scenedesmus obliquus protein as a replacement of the commercially available fish meal under an algal refinery approach. *Frontiers in Microbiology*, 10, 2114
- Pereira, H., Barreira, L., Figueiredo, F., Custodio, L., Vizetto-Duarte, C., Polo, C.,
 Resek, E., Engelen, A. & Varela, J. (2012). Polyunsaturated fatty acids of marine macroalgae: potential for nutritional and pharmaceutical applications. *Marine Drugs*, *10*, 1920–1935
- Perez-Lopez, P., Gonzalez-Garcia, S. & Jeffryes. (2014). Life cycle assessment of the production of the red antioxidant carotenoid astaxanthin by microalgae: from lab to pilot scale. *Journal of Cleaner Production*, 64, 332-44
- Perumal, P., Prasath, B., Santhanam, P. & Selvaraju, A. (2012). Isolation and culture of microalgae. Workshop on Advance in Aquaculture Technology, 166-181
- Popma, T. & Masser, M. (1999). *Tilapia life story and biology*. Southern Regional Aquaculture Center Publication



- Prabhu, A. J. (2015). Minerals in Fish: Does the Source Matter. Wageningen University: Ph.D. Thesis
- Pratiwi, D. Y. (2020). A mini review: Effect of Dunaliella salina on the growth and health of fish. *Asian Journal of Fisheries and Aquatic Research*, *10* (2), 1-8
- Preisig, H. R. & Andersen, R. A. (2005). *Historical review of algal culturing techniques*. Algal Culturing Techniques. London: Elsevier
- Priatni, S., Ratnanungrum, D., Kosasih, W., Sriendah, E., Srikandace, Y., Rosmalina, T. & Pudjiraharti, S. (2018). Protein and fatty acid profile of marine fishes from Java sea, Indonesia. *Biodiversitas*, 19 (5), 1737-1742
- Radhika, D., Veerabahu, C. & Marippandi, M. (2009). Effect of sewage cultured Lemna minor on growth of Mollinesia latipinna fed on mixed diets. *Current World Environment*, 4(2), 399-403
- Rahman, M. M., Choi, J. & Lee, S. (2013). Use of distillers dried grain as partial replacement of wheat flour and corn gluten meal in the diet of juvenile Black Seabream (Acanthopagrus schlegeli). *Turkish Journal of Fisheries and Aquatic Sciences*, 13, 699 706
- Raja, R., Shanmugam, H., Ganesan. & V., Carvalho, I. S. (2014). Biomass from Microalgae: An Overview. *Journal of Oceanography and Marine Research*, 2 (1), 118
- Raja, R., Hemaiswarya, S., Kumar, N. A., Sridhar, S. & Rengasamy, R. (2008). A perspective on the biotechnological potential of microalgae: Criticial Review.
 Microbiology, 34, 77-88
- Rakesh, S. G., Chawla, A., Singh, H., Chauhan, R. S. & Kant, A. (2016). Characterization and screening of native Scenedesmus sp. isolates suitabl for biofuel feedstock. *PLOS ONE*, 11 (5).
- Raven, J. A. & Beardall, J. (2004). Carbohydrate metabolism and respiration in algae. Springer, Netherlands, pp 205-224.
- Rawat, R., Ranjith, K., Mutanda, T. & Bux, F. (2011). Dual role of microalgae: Phycoremediation of domestic wastewater and biomass production for sustainable biofuels production. *Applied Energy*, 88, 3411–3424
- Reigh, R., Robinson, E. & Brown, P. (2007). Effects of dietary magnesium on growth and tissue magnesium content of blue tilapia Oreochromis aureus. Journal of the World Aquaculture Society, 22 (3), 192-200



- Rincon, D. D., Velasquez, H. A., Davila, M. J., Semprun, A. M., Morales, E. D. & Hernandez, J. L. (2012). Substitution levels of fishmeal by *Arthrospira spirulina* maxima meal in experimental diets for red tilapia fingerlings (*Oreochromis* sp.). *Revista Colombiana de Ciencias Pecuarias*, 25, 430–437
- Renuka, N., Sood, A., Prasanna, R. & Ahluwalia, A. S. (2015). Phycoremediation of wastewaters: A synergistic approach using microalgae for bioremediation and biomass generation. *International Journal Environmental Science Technology*, 12, 1443-1460
- Ruangsomboon, S., Choochote, S. & Taveekijakarn, P. (2010). Growth performance and nutritional composition of red tilapia (*Oreochromis niloticus* x O. *mossambicus*) fed diets containing raw *Spirulina platensis*. The International Conference on Sustainable Community Development 2010. 21–23 January 2010. Khon Kaen University, Nongkhai Campus, Thailand and Vientiane, Lao PDR
- Roem, A. J., Kohler, C. C. & Stickney, R. R. (1990). Vitamin E requirements of the blue tilapia, *Oreochromis aureus* in relation to the dietary lipid level. *Aquaculture*, 87, 155-164
- Ross, L. L. (2002). Environmental physiology and energetic. *Fish and Fisheries Series*, 25, 89–128
- Royes, J. B. & Chapman, F. A. (2003). Preparing your own fish feeds, document circular, department of fisheries and aquatic science, Florida Cooperative Extension Service. *Institute of Food and Agricultural Sciences*, 6 (2), 1-9
- Rubino, J. & Franz, K. (2012). Coordination chemistry of cooper proteins: How nature handles a toxic cargo for essential function. *Journal of Inorganic Biochemistry*, 107, 129-143
- Russo, G. L. (2009). Dietary n-6 and n-3 polyunsaturated fatty acids: from biochemistry to clinical implications in cardiovascular prevention. *Biochemical Pharmacology*, 77 (6), 937-946
- Safafar, H., Hass, M., Moller, P., Holdt, S. & Jacobsen, C. (2016). High- EPA biomass from Nannochloropsis salina cultivated in a flat panel photo bioreactor on a process water enriched growth medium. *Marine drugs*, 14 (8), 144



- Sahu, O. (2014). Reduction of Organic and Inorganic Pollutant from Wastewater by Algae. *International Letters of Natural Sciences*, *13*, 1-8
- Saiyasaeng, S., Yuangsoi, B., Wiriyapattanasub, P. & Wongmaneeprateep, S. (2014). Effects of dietary *Schizochytrium* sp. supplementation on morphological characters and growth performance of Nile tilapia (*Oreochromis niloticus*). *Khon Kaen Agriculture Journal*, 42, 45–50
- Santos, S. K. A. D., Schorer, M., Moura. G. D. S., Lanna, E. A. T. & Pedreira, M. M. (2019). Evaluation of growth and fatty acid profile of Nile tilapia (*Oreochromis niloticus*) fed with *Schizochytrium* sp. *Aquaculture Research*, 50 (4), 1068-1074
- Santos-Ballardo, D. U., Rossi, S. & Hernández, V. (2015). A simple spectrophotometric method for biomass measurement of important microalgae species in aquaculture. *Aquaculture*, 448, 87-92
- Sarker, B., Rahman, M. M. & Alam, M. N. (2015). A study on fish feed manufacture with its nutritional quality and impacts on fish production. *Research in Agriculture Livestock and Fisheries*, 2 (2), 353
- Sathasivam, R., Radhakrishnan, R., Hashem, A. & Abd_Allah, E. F. (2019). Microalgae metabolites: A rich source for food and medicine. *Saudi Journal* of Biological Sciences, 26 (4), 709-722
- Setiadi, E., Widyastuti, Y. R. & Prihadi, T. H. (2018). Water quality, survival, and growth of red tilapia, *Oreochromis niloticus* cultured in aquaponics System. Web of Conference, 47
- Shah, M. R. M. D., Lutzu, G. A., Alam, A. M. D., Sarker, P., Chowdhury, M. A.
 K. & Parsaeimehr, A. (2017). Microalgae in aquafeeds for a sustainable aquaculture industry. *Journal of Applied Phycology*, 30, 197–213
- Shiau, S. & Hsu. (1999). Quantification of vitamin C requirement for juvenile tilapia, Oreochromis niloticus x O. aureus, with L-ascorbyl-2monophosphate-Na and L- ascorbyl-2-monophosphate-Mg. Aquaculture, 175, 317-326
- Shiau, S. & Lin, Y. (2006). Vitamin requirements of Tilapia: A review. *Aquaculture*, 103, 311-320
- Shiau, S. & Suen, G. S. (1992). Estimation of the niacin requirements for tilapia fed diets containing glucose or dextrin. *The Journal of Nutrition*, 122, 2030-2036

- Shiau, S. & Lu, L. (2004). Dietary sodium requirement determined for juvenile hybrid tilapia (*Oreochromis niloticus* × O. *aureus*) reared in fresh water and seawater. *British Journal of Nutrition*, 91, 585-590
- Sioen, I., Henauw, S. D., Verdonck, F., Thuyne, N. V. & Camp, J. V. (2007). Development of a nutrient database and distributions for use in a probabilistic risk benefit analysis of human seafood consumption. *Journal of Food Composition and Analysis*, 20, 662-670
- Sinha, A. K., Kumar, V., Makkar, H. P. S., Boeck, G. D. & Becker, K. (2011). Nonstarch polysaccharides and their role in fish nutrition – A review. *Food Chemistry*, 127, 1409-1426
- Singh, R. A., Rohr, F. & Frazier. (2014). Recommendations for the nutrition management of phenylalanine hydroxylase deficiency. *Genetics in Medicine*, 16, 121–131
- Sirakov, I., Velichkova, K., Stoyanova, S. & Staykov, Y. (2015). The importance of microalgae for aquaculture industry. *International Journal of Fisheries and Aquatic Studies*, 2 (4), 81-84
- Slaninova, A., Machova, J. & Syobodoya, Z. (2014). Fish kill caused by aluminium and iron contamination in a natural pond used for fish rearing: A case report. *Veterinarni Medicina*, 59 (11), 573-581
- Soliman, A. K. & Wilson, R. P. (1992a). Water-soluble vitamin requirements of tilapia: Riboflavin requirement of blue tilapia, Oreochromis aureus. Aquaculture, 104, 309-314
- Soliman, A. K. & Wilson, R. P. (1992b). Water-soluble vitamin requirements of tilapia: Pantothenic acid requirement of blue tilapia, *Oreochromis aureus*. *Aquaculture*, 104, 121-126
- Solomon, S. G., Ataguba, G. A. & Abeje, A. (2011). Water stability and floatation test of fish pellets using local starch sources and yeast (*Saccahromyces cerevisae*). *International Journal of Latest Trends in Agriculture and Food Sciences*, 1(1)
- Tacon, A. G. J. & Metian, M. (2008). Global overview on the use of fish meal and fish oil in industrially compounded aquafeeds: Trends and future prospects. *Aquaculture*, 285, 146-158



- Tagliabue, A., Bowie, P., Boyd, K., Buck, K. & Saito, M. (2017). The integral role of iron in ocean biogeochemistry. *Nature*, 543, 51
- Tartiel, M. B., Ibrahim E. M. & Zeinhom, M. M. (2008). Partial replacement of fish meal with dried microalga (*Chlorella* spp and *Scenedesmus* spp) in Nile tilapia (Oreochromis niloticus) diets. *Proc. Of the 8th Int. Conf. on Tilapia in Aquaculture*. Cairo, Egypt
- Tessier, A., Blin, C., Cottet, M., Kue, K., Panfili, J., Chanudet, V., Descloux, S. & Guillard, J. (2018). Life history traits of the exploited Nile Tilapia (*Oreochromis niloticus*- Cichlidae) in a subtropical reservoir. *Cybium*, 43 (1). 71-82
- Thirupathaiah, M., Samatha, C. H. & Sammaiah, C. (2012). Analysis of water quality using physico-chemical parameters in lower manair reservoir of Karimnagar district, Andhra Pradesh. *International Journal of Environmental Sciences*, 3 (1), 172-180
- Tian, H. Z., Zhy, C. Y., Gao, J. J., Cheng, K., Hao, J. M. & Wang, K. (2015). Quantitative assessment of atmospheric emissions of toxic heavy metals from anthropogenic sources in China: Historical trend, spatial distribution, uncertainties, and control policies. *Atmospheric Chemistry and Physics*, 15, 10127-10147
- Tibbetts, S. M., Melanson, R. J., Park, K. C., Banskota, A. H., Stefanova, R. & McGinn, P. J. (2015). Nutritional evaluation of whole and lipid-extracted biomass of the microalga *Scenedesmus* sp. isolated in Saskatchewan, Canada for animal feeds: Proximate, amino acid, fatty acid, carotenoid and elemental composition. *Current Biotechnology*, *4*, 530-546
- Tocher, D. R. (2003). Metabolism and functions of lipids and fatty acids in teleost fish. *Review in Fisheries Science*, *11*, 107–184
- Tower. L. (2005). Farming Tilapia: Life history and biology. Retrieved from https://thefishsite.com/articles/tilapia-life-history-and-biology
- Toyub, M. A., Miah, M. I., Habib, M. A. B. & Rahman, M. M. (2012). Growth performance and nutritional value of Scenedesmus Obliquus cultured in different concentrations of sweetmeat factory waste media. *Bangladesh Journal of Animal Science*, 37 (1), 86-93

- Ugoala, C., Ndukwe, G. I. & Audu, T. O. (2008). Comparison of fatty acids profile of some freshwater and marine fishes. *Internet Journal of Food Safety*, *10*, 9– 17
- Vaikosen, S. E., Nwokoro, S. O. & Orheruata, A. M. (2007). Yield and chemical composition of *Chlorella* species cultivated in pig, poultry and cow dungs in Southern Nigeria. *Agriculture and Environment*, 7, 229-235
- Velichkova, K. (2014). Effect of different nitrogen sources on the growth of microalgae *Chlorella vulgaris* cultivation in aquaculture wastewater. *Agricultural science and technology*, 6 (3), 337–340
- Vizcaino, A. J., Lopez, G., Saez, M. I., Jimenez, J. A., Barros, A., Hidalgo, L., Camacho- Rodriguez, J., Martinez, T. F., Ceron- Garcia, M. C. & Alarcon, F. J. (2014). Effects of the microalga *Scenedesmus almeriensis* as fishmeal alternative in diets for gilthead sea bream, *Sparus aurata*, juveniles. *Aquaculture*, 431, 34-43
- Vonshak, A. (1997). Appendix III–Growth media and conditions for Spirulina. Spirulina platensis (Arthrospira): Physiology, Cell-biology and Biotechnology, 218-219
- Walker, A. B. & Berlinsky, D. L. (2011). Effects of partial replacement of fishmeal protein by microalgae on growth, feed intake, and body composition of Atlantic cod. *North American Journal of Aquaculture*, 73, 76–83
- Wang, M. & Lu, M. (2016). Tilapia polyculture: A global review. *Aquaculture Research*, 47(8), 2363-2374
- Wang, Y., Hu, M., Cao, L., Yang, Y. & Wang, W. (2008). Effects of daphnia (*Moina micrura*) plus chlorella (*Chlorella pyrenoidosa*) or microparticle diets on growth and survival of larval loach (*Misgurnus* anguillicaudatus). Aquaculture International, 16, 361–368
- Watanabe, T., Satoh, S. & Takeuchi, T. (1988). Availability of minerals in fish meal to fish. Asian Fish Sience, 1 (2), 75-195

Watershed Academy Web. (2021). The Effect of Climate change on water Resource and Programs. Retrived from https://cfpub.epa.gov/watertrain/moduleFrame.cfm?parent_object_id=2456 &object_id=2459



- World Bank. (2013). Fish to 2030. Prospects for Fiseries and Aquaculture, Washington, DC
- Yee, L. T., Paka, D. D., Nyanti, L., Ismail, N. & Emang, J. J. J. (2012). Water Quality at Batang Ai hydroeletric reservoir (Sarawak, Malaysia) and implications for aquaculture. *International Journal of Applied Science and Technology*, 2 (6), 23-30
- Yusoff, A. (2015). Status of resource management and aquaculture in Malaysia. Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia: Challenges in Responsible Production of Aquatic Species. Proc. of the Int. Conf on Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia. Tigbauan, Iloilo, Philippines.pp. 53-65
- Zamal, H., Barua, P., Uddin, B. & Islam, K. S. (2008). Application of ipil-ipil leaf meal as feed Ingredient for monosex tilapia fry (*Oreochromis niloticus*) in terms of growth and economics aquaculture. *The Asia Magazine*, 5, 31-33
- Zhang, T. & Nakajima, M. (2014). Advances in applied biotechnology. Procs of the second Int. Conf on Applied Biotechnology. Springer. pp. 245-25



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